



# ECOLOGICAL RISK ASSESSMENT OF J.H. BAXTER & CO. EUGENE, OREGON PLANT SITE

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#### 1. INTRODUCTION

The J.H. Baxter Wood Preserving site in Eugene, Oregon (the "Site") (Figure 1), has been treating wood for several decades. The State of Oregon environmental cleanup law and administrative rules requires that characterisation of current and reasonably likely future risk posed by hazardous substances be based on baseline human health and ecological risk assessment. To address the requirement to perform a baseline ecological risk assessment (BERA), the Oregon Department of Environmental Quality (DEQ) has requested a Level II - screening ecological risk assessment be conducted for the vegetated area in the south west corner of the facility (fallow area), hereafter referred to as the area of potential concern (AOPC) (Figure 2). DEQ has stated that the fallow area is the AOPC at the Site with respect to ecological receptors.

#### 1.1. Site Description

Refer to the Phase II Remedial Investigation report.

#### **1.2.** Site History

Refer to the Phase II Remedial Investigation report.

#### **1.3.** Summary of Previous Ecological Investigations

A qualitative Environment Impact Assessment (EIA) was conducted as part of the Phase II Remedial Investigation for the Site in 1994. An ecological survey to determine if threatened or endangered species were present or likely to be present on the Site, was performed. Except for a small area in the southwest corner of the Site there was no habitat suitable for threatened or endangered species. At the time of the survey this

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corner of the Site was dominated by the Himilayan blackberry. Below the blackberry there was a band of reed canary grass, which circled a small pond. Beggar ticks dominated the bottom of the pond. No threatened or endangered species were found on the Site during the site survey.

#### 1.4. Assessment Objectives and Scope

The objective of the BERA is to evaluate the potential risk to ecological receptors in the AOPC from exposures to contaminants present at the Site. The performance of a BERA is governed by the State of Oregon environmental cleanup law ORS 465, and administrative rules OAR 340-122-080 and OAR 340-122-084. The BERA is being conducted under the guidance of DEQ and according to the *Guidance for Ecological Risk Assessment Level II - Screening* (DEQ, 1998). The BERA is designed to determine if constituents in the abiotic media of the AOPC pose a potential risk to ecological receptors and to assist in the development of risk-based remediation goal options. It will also provide information needed to evaluate and compare potential cleanup alternatives for future risk management decisions.

#### 2. SITE SURVEY

The site survey was conducted by Mr. Bruce Newhouse of Salix Associates of Eugene, Oregon. The AOPC is in the south west corner of the J.H Baxter site, and is approximately 3.5 acres in size. It is about 300 feet by 500 feet in dimensions. The land form is generally flat except for a "L" shaped berm which runs along the south and west side of the AOPC. Along the north edge and northeast corner is a low lying area that ponds water from early winter to mid summer. The AOPC is dominated by scrub shrub and emergent wetland habitats, and upland herb-dominated habitats. A berm which runs along the south and west sides of the AOPC is dominated by Himalayan blackberry

(Rubus discolor), an introduced pest. The intermittent pond area is dominated by creeping foxtail (Alopercurus geniculatus) and nodding beggars-ticks (Bidens cernua). Reed canary grass (Phalaris arundincacea) is encroaching around the perimeter, and a few small patches are scattered in the intermittent pond area. The survey report is found in Appendix A.

#### 3. PROBLEM FORMULATION

#### 3.1. Assessment Endpoints and Endpoint Species

## 3.1.1. Assessment Endpoints

An assessment endpoint is an explicit expression of a specific ecological receptor and an associated function or quality that is to be maintained or protected; that provides a clear connection between regulatory policy goals and risk assessment results. For this assessment, the policy goal was protection of all terrestrial species from any adverse impacts due to the presence of site-related contaminants. These considerations lead to definition of the following assessment endpoints: *Protection of mammalian, and avian, receptors within the foraging range of the Site from reproductive impairment resulting from exposure to contaminants of potential ecological concern (CPEC) in the locality of the facility, and protection of plant and soil invertebrate receptors within the locality of the facility impairment resulting from exposure to CPECs.* 

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#### 3.1.2. Endpoint Species

Specific ecological receptors (termed endpoint species) were selected from the site-specific species list developed form the site survey: based on their having substantial aesthetic, social, or economic value or are important in the biological functions or biodiversity of the system. These endpoint species are either themselves the object of protection or serve as surrogates for other ecological receptors requiring protection. For this assessment, endpoint species were selected on the basis of their: (a) documented presence at or in the locality of the site, (b) use in previous tissue residue studies and risk assessments, (c) sensitivity to the contaminant of concern, (d) ecological relevance, and (e) connection to policy goals.

The following endpoint species were selected: earthworms, Himalayan blackberry (Rubrus discolor), Reed canary grass (Phalaris arundincacea), deermouse (Peromyscus maniculatus), and the American Kestrel (Falco sparverius). The DEQ has concurred with the selection. The earthworm was selected to represent soil invertebrates, which are eaten by many carnivorous and omnivorous small mammals and birds including the American kestrel and deer mice. Earthworms are also in direct contact with CPECs in the soil. The Himalayan blackberry was selected to represent plants found on the site because of their great abundance on the site and because they are a source of food for many wildlife species including birds and deer mice. Reed canary grass (Phalaris arundincacea), found in water and wet places, was selected to represent plants growing in the intermittent ponded area of the AOPC because it was found in abundance, and it is a potential source of food for mammalian and avian species including deer mice. The American kestrel was selected because it is a top predator species that may potentially intake contaminants from various food sources. It represents the culmination of bioaccumulation processes in the food web (Figure 3). Higher-level predators tend to be ecological receptors of greatest concern because of this trophic bioaccumulation and their greater longevity. Although

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the majority of its diet is small mammals such as the deer mouse and small birds, the American kestrel will preferentially eat insects (EPA, 1993; Causti 1997) (Table 1).

## 3.2. Contaminants of Potential Ecological Concern (CPEC)

If the concentration of a contaminants of interest (COIs) in soil was less than 50 times its Level II Screening benchmark values (SBVs), as described in the Guidance for Ecological Risk Assessment (DEQ, 1998), and there were no observable significant effects to the health to the local population of each endpoint species, then the COI was not selected as a CPEC. The COI concentrations used for comparison were the maximum of the composite samples, as measured (SS98-12COMP, SD98-6COMP), and the calculated composite concentration (Tables 2 and 3). Four soils samples, that were not composites, were samples collected on February 3, 1998, and analyzed for phenols, polycyclic aromatic hydrocarbons (PAHs), arsenic, copper, chromium, and zinc. To compare the individual samples as measured to the composite samples taken at the area of concern, a composite concentration was calculated for each COI measured in the February 3, 1998, sampling event. The 90 percent UCL derived using the standard Bootstrap method were the calculated composite concentrations. The calculated composite concentration was referred to as "SS98-1-4 COMP" (Tables 4, 5, and 6).

The bootstrap method is a nonparametric statistical method (US EPA 1997). In the bootstrap procedure, repeated samples are drawn with replacement from the given set of observations. The process is repeated a large number of times, and each time an estimate of the mean is computed. The estimates thus obtained are used to calculate the standard error of the mean. The bootstrap UCL90 concentration is calculated as follows:

#### $UCL95 = x + z_{\alpha}\sigma_B$

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where:

$$\sigma_{B} = \sqrt{\frac{1}{N-1} \sum_{i=1}^{N} (x_{i} - x_{B})^{2}}$$

$\sigma_{\text{B}}$	Ξ	the bootstrap estimate of the standard error
x	=	the statistically unbiased estimate of the mean
$x_i$	_	the mean of the i <sup>th</sup> sample size n with replacement
x <sub>B</sub>		the bootstrap estimate of the population mean if the N
		estimates x <sub>i</sub> .
$Z_{\alpha}$	=	the standard normal critical values
N	=	number of times data re-sampled (1000)

(USEPA 1997).

When SBVs for COIs were not provided by DEQ, SBVs for COIs were derived by applying uncertainty factors to toxicity endpoints from the literature (Table 7). There are no DEQ SBVs for PAHs in soil except for benzo(a)pyrene with respect to mammals and fluoranthene with respect to plants. Toxicity endpoints from literature were used to screen the other PAHs in soil (Table 8). No PAHs were in excess of their literature based SBVs. PAHs in soil were not, therefore, carried forward as CPECs in this risk assessment. Individual chloro- and nitro-phenols in the soil of the AOPC were not detected and thus were not carried forward as CPECs (Table 9). Arsenic, copper and zinc were present in the soil at levels below 50 times their SBVs and were thus not carried forward as CPECs (Table 10). The level of chromium in the soil exceeded 50 times its SBV in soil with respect to soil invertebrates only (Table 4). However, the level of chromium in the soil (49.9 mg/kg) is less than the background concentration of chromium for the area (61 mg/kg). Therefore chromium was not retained as a CPEC.

found in soils at levels below 50 times their SBV; however, most were in excess of their SBV (Table 11). Dioxins and furans are persistent, lipophilic, highly toxic, and tend to bioaccumulate so that total exposure potentially increases from one trophic level to the next. For these reasons dioxin and furans are retained as CPECs in the AOPC soil.

If the concentration of a contaminants of interest (COI) in sediment was less than 10 times its' Level II Screening benchmark values (SBVs), as described in the Guidance for Ecological Risk Assessment (DEQ, 1998), and there were no observable significant effects to the health to the local population of each endpoint species, then the COI was not selected as a CPEC. All PAHs were present in the sediment at levels below 10 times their SBV (Table 12). PAHs were therefore not carried forward as CPECs. Individual chloro- and nitro - phenols in the sediment of the AOPC were not detected and thus were not carried forward as CPECs (Table 13). Arsenic, chromium, copper and zinc were present in the sediment at levels below 10 times their SBVs and were thus not carried forward as CPECs (Table 14). DEQ has no SBVs for dioxins and furans in sediment. Dioxins and furans are persistent, lipophilic, toxic and tend to bioaccumulate. For these reasons dioxin and furans are considered CPECs in the AOPC sediment (Table 15).

#### 3.3. Risk Hypotheses

The following are risk hypotheses for this risk assessment:

- Dioxin and furan and their congeners in the soil and sediment of the AOPC will not effect invertebrates and plants in the AOPC.
- Food chain accumulation and transfer of dioxin and furan and their congeners does not occur to the degree that allows for effects to the chicks of the American Kestrel utilising the site due to embryo exposures to dioxin and furan and congeners.



• Food chain accumulation and transfer of dioxin and furan and their congeners to deer mice does not occur to the degree that allows for effects to deer mice utilising the site.

## 3.4. Relevant and Complete Exposure Pathways

An exposure route is the pathway by which a chemical or physical agent comes in contact with a receptor (i.e., by ingestion, inhalation, dermal contact, etc.). Ecological receptors may be exposed to chemical contaminants either through direct (primary) and/or indirect (secondary) exposure routes. Only those pathways that are complete, and are expected to contribute substantially to exposures by ecologically important receptors, are addressed. The following exposure pathways were addressed in this risk assessment:

- Direct contact and ingestion of soil by soil invertebrates;
- Direct contact of plant roots with soil;
- Ingestion of soil by terrestrial vertebrates;
- Ingestion, by terrestrial vertebrates, of plants that have bioaccumulated CPECs from the soil;
- Ingestion, by small mammals, of soil invertebrates that may have bioaccumulated CPECs from the soil;
- Direct contact of plant roots with sediment;
- Direct contact and ingestion of sediment by aquatic invertebrates;
- Ingestion, by carnivorous birds, of terrestrial and aquatic invertebrates that may have bioaccumulated CPECs from the soil or sediments (this is an indirect exposure pathway); and
- Ingestion, by carnivorous birds, of small mammals that may have bioaccumulated CPECs from soil invertebrates and plants that may intern have bioaccumulated CPECs from the soil. This is an indirect exposure pathway.

The relevant and complete exposure pathways present in the AOPC are summarized in Figure 4.

#### 4. EXPOSURE ANALYSIS

The objective of exposure analysis is to estimate the concentration or dose of a contaminant received by an ecological receptor, taking into consideration a number of factors. These factors include the spatial distribution of contaminant concentrations relative to the spatial distribution of receptors. The exposure point value (EPV) is, for terrestrial species, the contaminant dose received by the receptor (applied dose) or, for aquatic species, the contaminant concentration in the media in which receptors are immersed (surface water or sediment). EPV estimation is a multi-step process, as described below.

## 4.1. Habitats and Receptors Considered

#### 4.1.1. Habitats

The AOPC is in the south west corner of the Site. It is approximately 3.5 acres in size. There are two distinct areas on the AOPC, a low lying area in the north east corner that ponds water from early winter to mid summer and whose dominant vegetation is reed canary grass, beggars ticks, and foxtail. The rest of the AOPC is generally flat except for an "L" shaped berm which runs along the south and west sides of the AOPC. This area of the AOPC is dominated by Himalayan blackberry. To the north and east of the AOPC is the active industrial portion J. H. Baxter Wood Preserving Facility. To the west of the site is another industrial site. The portion of this site adjacent to the AOPC is a vegetated field, part of which is used for log storage. To the south of the AOPC across the rail road tracks is vacant industrial zoned property that is covered with grasses and weeds. Using



the U.S. Environmental Protection Agency SITEPLUS map system a map was created showing the population distribution, national parks and recreation areas, wetlands, schools and hospitals on the lands within a 5 mile radius of the centroid of the J.H. Baxter site (Figure 7). Areas within the 5 mile radius were classified as industrial, commercial, residential, vacant, and roads and rail.

## 4.2. CPEC Environmental Concentrations (EC)

The calculation of the concentration of CPECs in sediment and soil followed that described in the work plan. Dioxins/furans in the composite sediment sample, which was comprised of five specimens, was carried forward into the risk assessment. (Table 15) The concentration of dioxins and furans in soil used in the risk assessment was the maximum of the following:

- 1) the measured concentration of dioxins and furans in a composite sample comprised of five specimens and sampled in the AOPC during the October 7, 1998 sampling event (Table 2 and 3), or
- 2) the measured concentration of dioxins and furans in a calculated composite sample comprised of four specimens and sampled in the AOPC during the February 3, 1998 sampling event (Table 2 and 3).

Dioxins and furans are believed to all have the same mode of action and effect (EPA, 1993a; Powell et al, 1998). Therefore to determine the maximum potential effect of dioxins and furans all congeners of 2,3,7,8-tetrachlorodibenzo-p-dioxin (2,3,7,8 TCDD) for which the soil and sediment was analyzed were converted to 2,3,7,8 - TCDD toxic equivalents (TCDD-EQ) using the generally accepted international toxicity equivalency factors (I-TEFs) (Table 11). To be conservative the TCDD-EQ for each 2,3,7,8-TCDD congener was then summed. The total TCDD-EQ value for dioxins and furans in soil and sediment, was assumed to be the concentration in the soil and sediment.

The background level of 2,3,7,8-TCDD has been reported as 8 parts per trillion (ppt) (EPA 1997). To be conservative it was assumed that the ambient level of TCDD-EQ in the City of Eugene is 32 ppt.

#### 4.3. Exposure Estimation Models

The EPV for the sedentary endpoint species, soil invertebrates, and plants were based on The EPV carnivorous avian and the local concentration of CPECs in the soil. omnivorous mammal endpoint species was based on the dietary intake of the selected species and the CPEC concentrations therein. The estimation of CPEC concentration in the dietary food of the avian and mammalian endpoint species was based on the The foods. the dietary **CPECs** by biouptake/bioaccumulation of bioaccumulation/biouptake of CPECs was estimated using validated mathematical models or using literature biouptake values. The soil-to-wet plant uptake factor for metal and organic CPECs and the log of the octanol-water partition coefficient (Kow), for organic CPECs, was taken from the Toxicity & Chemical-Specific Factors Data Base (Risk Assessment Program Oak Ridge National Laboratory, 1998) (Table 16). The bioaccumulation of organic CPECs by earthworms was calculated using the "Earthworm Model" (Jager and Hammers, 1997). The following equations are used to calculate the bioaccumulation of organic CPECs in earthworms:

Equation 1

$$BCF = \frac{F_{water} + F_{fat} + Kow}{\rho_{worm}}$$

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where:

BCF	-	bioconcentration factor for worm [m <sub>water</sub> <sup>3*kgwwt<sup>-1</sup>]</sup>
F <sub>water</sub>	=	volume fraction of water in worm $[m_{water}^{3} * m_{worm}^{-3}]$
F <sub>fat</sub>	=	volume fraction of fat in worm [m <sub>fat</sub> <sup>3</sup> *m <sub>worm</sub> <sup>-3</sup> ]
Kow	H	octanol-water partition coefficient $[m_{water}^{3} * m_{oct}^{-3}]$
$\rho_{worm}$		bulk density of worm [kg <sub>wwt</sub> *m <sub>worm</sub> <sup>-3</sup> ]

Equation 2

$$BAF = \frac{\frac{k_1}{Kp} + k_f}{k_2 + k_g + k_m}$$

where:

$$Koc = \frac{1.26 \times Kow^{0.81}}{1000}$$
$$Kp = Foc * Koc$$

BAF	-	bioaccumulation factor
Kp	=	solids-water partition coefficient in soil [m <sup>3</sup> /kg <sub>solids</sub> ]
Foc	= .	fraction of organic carbon in soil [kgoc/kgsolids]
Koc	=	organic carbon-water partition coefficient $[m_{water}^{3}/kg_{oc}]$
kı	=	0.35 (a constant when log Kow>4) diffusive uptake
		$[(mg * kg_{wwt})/(mg * m_{porew}^3)/d]$
k <sub>2</sub>	==	0.35/BCF diffusive losses [1/d]
k <sub>f</sub>	=	0.034 (uptake through food) [(mg * kg <sub>wwt</sub> )/(mg * $m_{porew}^{3}$ )/d]
kg	=	0.0058 (dilution by growth) [1/d]
k <sub>m</sub>	=	0 (losses due to metabolism = 0 due to lack of general
		knowledge) [1/d]

(Figure 5; Table 17).

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## 4.4. Exposure Point Value (EPV) Estimates

An exposure model is used to estimate the EPV. An exposure model must explicitly consider the spatial relationships between endpoint species, their habitat, and the distribution of contaminants, as well as habitat quality and temporal utilization of habitat. Simple non-spatial exposure models are often based on the assumption that contaminants are evenly distributed on the site or that an endpoint species forages randomly with respect to contamination on that portion of the site which constitutes habitat. In either case, an endpoint species is assumed to be exposed to mean concentrations.

However, because many sites are industrial or highly modified in nature, it is unlikely that all areas within their bounds will provide habitat suitable for endpoint species. For example, contaminant concentrations might be greatest near the centre of a site, but the habitat quality might be highest near the edges. Thus, if contaminant levels are related to habitat quality, the assumptions of a simple model would not hold. A more reasonable model and the model used in this risk assessment accounts for the proportional contribution of each area with a distinct combination of contaminant level and habitat quality, as follows:

Equation 3 
$$EPV_j = \sum_{k=1}^{q} \sum_{i=1}^{d} \left[ \left( \frac{IR_i \times C_{ijk}}{BW} \right) \times \left( \frac{\left( Hq_k / \sum_{k=1}^{q} Hq_k \right) \times Ha_k}{\sum_{k=1}^{q} Ha_k} \right) \right] \times AU \times S$$

where:

 $EPV_j = Exposure point value for a given endpoint species for j<sup>th</sup> contaminant (mg/[kg·d]);$ 

q = Number of habitat patches within local population boundary (unitless);
 d = Total number of i<sup>th</sup> media (e.g., food, water, soil);

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- $IR_i = Intake rate for i<sup>th</sup> medium (kg/d or L/d);$
- $C_{ijk} = 90^{th}$  percentile UCL concentration of  $j^{th}$  contaminant in  $i^{th}$  medium in  $k^{th}$  habitat patch (mg/kg or mg/L);

BW = Body weight of endpoint species (kg);

 $Hq_k$  = Relative habitat quality (based on expected residency) of  $k^{th}$  habitat patch for a given endpoint species (unitless);

 $Ha_k = Area of k^{th} habitat patch (m^2);$  and

AU = Area use factor (unitless).

S = Seasonality factor for small mammal (unitless).

Equation 3 assumes that individuals within the local population boundary use habitat in proportion to the habitat area and quality. Here EPV is the applied dose (mg/[kg·d]) experienced by an individual of the endpoint species. Since multiple food items are considered, Equation 3 includes a term to represent that item's fraction of IR<sub>i</sub> in the total intake, e.g., incidentally ingested soil may be only a small fraction (< 2%) of total food items.

For abiotic media such as soils or water,  $C_{ijk}$  equals the environmental concentration. In the absence of measured contaminant concentrations, the value of  $C_{ijk}$  in the tissues of consumed prey and forage has been estimated using the environmental concentrations in soil and sediment in conjunction with an appropriate intermedia transfer factor. Some endpoint species have migration, hibernation, or other behaviour patterns that result in less exposure throughout the year at a site. The seasonality factor (S) quantifies the frequency of exposure to contaminated media as a function of such behaviour patterns. This factor is defined as the fraction of the number of days per year an endpoint species is active within a habitat, so that  $1 \ge S > 0$ . Non-hibernating, non-migratory species will have a unitless default seasonality factor of 1. For those species that use the habitat island only as a stop-over point during their annual migration it will be necessary to estimate a S

value < 1. The S for the mammalian and avian endpoint species used in this assessment was assumed to be 1. An area use factor (AU) is included to account for the effect of a receptor's territory size or foraging area on frequency and duration of contact with contaminated media or prey. When a terrestrial receptor's territory size ( $A_{bird}$ ) or foraging area ( $A_{mammal}$ ) is equal to or less than the total contaminated area within the locality of the facility ( $A_{site}$ ), AU will have a unitless default value of 1.0. When this area exceeds the area of contamination, AU is calculated as the ratio of the contaminated area to foraging area and will have a value less than 1.0.

#### 4.4.1. Define Local Population Boundaries

Establishing a species-specific local population boundary sets a limit on the number of individual members of an endpoint species population that will be considered in the risk assessment. The boundary must be established with reference to the locality of the facility and must relate in some way to a biological feature of the species. The boundary cannot be so large as to include individuals that will have little probability of contacting a site-related contaminant or be so small as to exclude individuals who might reasonably contact such a contaminant. Its size will also necessarily have to vary with the endpoint species. With the recognition that the following approach has ecological limitations but practical advantages, population boundaries for sessile, mobile, and migratory species will be established as follows:

- For sessile terrestrial species (e.g., plants), the local population spatial boundary is assumed to be equal to the spatial boundaries of the locality of the facility.
- For sessile aquatic species (e.g., benthic invertebrates) in ponds or lakes within the locality of the facility, the local population spatial boundary is assumed to be equal to the spatial boundaries of the water body.



Individuals of mobile terrestrial and avian species usually travel varying distances, on a daily to seasonal basis, to find food, water, and shelter. The area encompassed by these travels is termed an individual's foraging home range (FHR). Studies of dispersal behaviour in mammals suggest that there is a low probability of an animal moving more than five FHR diameters in a straight line from its natal range (Waser, 1987). Thus for both terrestrial and avian mobile species, the local population spatial boundary diameter is assumed to be equal to five FHR diameters from the spatial boundaries of the locality of the facility. FHRs are assumed to be non-overlapping.

For the deer mouse and the American Kestrel, the local population spatial boundary was determined as follows:

$$D_h = 10 \times \sqrt{\frac{A_h}{\pi}}$$

#### **Equation 4**

where:

 $D_h$  = Local population boundary diameter for the *h*th endpoint species (m); and  $A_h$  = Foraging home range area for the *h*th endpoint species (m<sup>2</sup>).

The foraging home range and the local population boundary diameter for the selected mammalian endpoint species is found in Table 18 (Figures 6 and 7) (USEPA 1993; Causti et al 1997).

## 4.4.2. Habitat Size and Quality

Observation and mapping of habitats were used to estimate: (a) number (q), (b) approximate habitat spatial extent (Ha<sub>k</sub>), and (c) relative habitat quality (Hq<sub>k</sub>) (based on relative expected residency) of each habitat (or habitat patch) within the local population boundary, including the locality of the facility. Presumably, habitat patches with greater relative quality will increase the probability of exposure by attracting and holding an endpoint species more strongly and for a longer duration (i.e., raising its expected residency) than those with minimal habitat quality. The quality of habitat, with respect to the needs of a given endpoint species relative to all other existing species-specific habitat within the local population boundary, were rated as follows:

- unsuitable (0),:
- poor (0.25);
- average (0.5);
- good (0.75); or
- excellent (1).

The habitat quality for the mammalian and avian endpoint species was determined based on a site visit, the performance of a species habitat survey and knowledge of the habitat requirements of the endpoint species. The resulting  $Hq_k$  values were then normalised so that the sum of all such factors for a given locality equals one. Information on each endpoint species is found in Appendix B. Tables 19 and 20 contains the habitat quality rating for mammalian and avian endpoint species.

Five habitat patches were identified in the local boundary for deer mice. The five habitat patches identified for the deer mouse are the following:

- 1) grass and shrubs;
- 2) intermittent pond;

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- 3) grass;
- 4) grass-storage;
- and 5) industrial-active.

Seven habitats were identified in the local boundary for the American Kestrel. Because the size of the American kestrel local population boundary is so large only the two habitat patches within the AOPC 1) grass and shrubs, 2) intermittent pond, were specifically identified. The other five habitats areas are the following:

- 1)industrial (grass & shrubs);
- 2) commercial (grass & shrubs);
- 3) residential (grass & trees);
- 4) vacant (grass & shrubs); and
- 5) buildings active industrial sites and paved areas (Table 21).

These five habitat types were referenced in terms area of the percent of land each habitat occupies within the local population boundary. Information on land use was acquired from Clair Van Bloem of the Lane Council of Governments (1998). The information was based on information collect during 1994. It has been assumed the land allowed to grow fallow in the Eugene metropolitan area will soon end up with a combination of grass and shrubs. It was assumed that for residential land that 25% of the land was building or covered with pavement, gravel, or concrete. Sixty six percent of commercial land was assumed to be occupied by building or covered with pavement, concrete, or gravel. Half of all industrial land was assumed to be an active part of the facility, covered by building or paved. The type, size and relative quality of each habitat patch with respect to the mammalian endpoint species is listed in Tables 19, 20 and 21.

Areas covered with buildings, pavement, gravel, or concrete, and roads and rail lines, and active industrial sites were given a habitat rating of 0 and not carried any further into the EPV analysis.

#### 5. ECOLOGICAL RESPONSE ANALYSIS

#### 5.1. Receptor Toxicity Profiles

## 5.1.1. Dioxins and Furans

Dioxins and furans are ubiquitous in the environment. The background level of 2,3,7,8tetrachlorodibenzo-p-dioxin (2,3,7,8-TCDD) has been reported as 8 ppt (EPA 1997). Ecological receptors on the AOPC may potentially be exposed to elevated levels of dioxins and furans (Tables 4 and 6) present in AOPC soils and sediment. Dioxins and furans are polyhalogenated diaromatic hydrocarbons (PHDHs). 2,3,7,8-TCDD is the most toxic of these compounds. 2,3,7,8-TCDD is believed to exert many of its toxic effects by binding with the aryl hydrocarbon receptor (AhR), and subsequently entering the nucleus where this ligand AhR complex activates specific genes including CYP1A1, which encodes for protein cytochrome P4501A1 (Powell, D.C. et al, 1998). Enhanced expressions of this and other associated genes leads to a variety of responses attributed to TCDD. Other PHDHs structurally similar to 2,3,7,8-TCDD are known to act through the same mechanism of toxicity (Schecter, 1994). This allows the other PHDHs to be expressed relative to TCDD in the form of toxic equivalency factors. The TEFs can then be used to determine dioxin toxic equivalents (TEQs).

Dioxins and furans are known to cause a variety of adverse effects in avian species including embryo lethality, beak deformities, subcutaneous edema, hydropercardium, liver lesions and induction of P450 enzymes (Schecter 1994; Powell D.C., 1998). When chicken embryos were exposed to 2,3,7,8-TCDD injected into the air cell at the start of

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incubation the chick had a LD50 of 0.297 ug/kg egg (Hensel, 1997). When doublecrested cormorant were exposed to 2,3,7,8-TCDD injected into the air cell at the start of incubation the LD50 calculated from mortality at hatching was of 4 ug/kg egg (Powell D.C., 1998) and when exposed to PCB 126 the calculated LD 50 was 177 ug/kg. The LD50 for the congeners of 2,3,7,8-TCDD polychlorinated biphenyl (PCB) 126 was 65 ug/kg egg for American kestrels (Hoffman 1996). Hoffman reported a TEF of 0.05 for the conversion of PCB 126 to TCD-EQ. This is more than twice as large as the TEF (0.022) calculated for the cormorant by Powell (1998). The TEF of 0.05 was use to convert PCB 126 exposures to the American kestrel TCDD-EQ.

Dioxins and furans are known to cause a variety of adverse effects in mammalian species including hepatotoxicity, wasting syndrome, immunotoxicity, and dermatitis in guinea pigs (Decaprio, 1986), amylodosis of the kidney spleen and liver, and cleft palate in mice (Toth et al 1979; Dillman 1982) and tetragenic effects in rats (Smith et al 1976). The acute toxicity of 2,3,7,8-TCDD to experimental animals is quite variable. Studies by Hochstein et al (1988) and Aulerich et al (1988) indicated that mink is among the mammalian species most sensitive to 2,3,7,8-TCDD intoxication (EPA 1993). The 28 day LD50 of 4.2 ug/kg for the mink is more than the LD50 of 0.600 to 2.0 ug/kg for the guinea pig, but less than the LD50 of 22 to 45 ug/kg for the rat, the LD50 of 115 ug/kg for the rabbit, and LD50 of 114 to 284 for the mouse (EPA, 1993).

## 5.2. Ecological Benchmark Value Estimates

Per OAR 340-122-084(1)(h)(B)(ii), effects on species other than those classified as threatened or endangered are made only at the population level. The EBV for populations is defined as the median lethal dose or concentration ( $LD_{50}$  or  $LC_{50}$ ). If a  $LD_{50}$  or  $LC_{50}$ , was not available for endpoint species considered in this risk assessment, the EBV was

derived from other toxicological endpoints for those receptors or appropriate surrogates for those receptors, adjusted with uncertainty factors to equate to a  $LD_{50}$  or  $LC_{50}$ . The uncertainty factors process shown in Figure 3 of *the Guidance for Ecological Risk Assessment Level III - Baseline* (DEQ, 1998), is used to convert available toxicological endpoints to a  $LD_{50}$  or  $LC_{50}$  for an endpoint species.

The 2,3,7,8-TCDD EBV for the deer mouse is based on the LD 50 for 2,3,7,8-TCDD in the mouse (species unspecified) of .114 mg/kg/day (EPA, 1993). Using the methodology shown in Figure 3 of *the Guidance for Ecological Risk Assessment Level III - Baseline* (DEQ, 1998) LD50 for the mouse was converted to an LD 50 for the deer mouse. The mouse is assumed to be of the same genus as the deer mouse thus the LD50 for 2,3,7,8-TCDD in the mouse was multiplied by an uncertainty factor of 0.5 to account for the possibility of the mouse and deer mouse being a different species. The LD50 was multiplied by an additional uncertainty factor of 0.5 to account for the deer mouse not being a threatened or endangered species. EBV for 2,3,7,8-TCDD in the deer mouse used for the in this risk assessment was 0.029 mg/kg/day (Table 22).

The 2,3,7,8-TCDD EBV for the American kestrel was based on the LD 50 for polychlorinated biphenyl (PCB) 126 in the American kestrel of 0.065 mg/kg/day (Beyer et al, 1996). The conversion of PCB 126 to 2,3,7,8-TCDD was based on multiplying the LD50 for PCB 126 in the American kestrel by the toxic equivalency factor of 0.05 (Kubiak, 1991). The LD50 was multiplied by an additional uncertainty factor of 0.5 (DEQ, 1998) to account for the American kestrel not being threatened or endangered. The EBV used for 2,3,7,8-TCDD in the American kestrel in this risk assessment was 0.00163 mg/kg/day (Table 22).

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## 6. RISK CHARACTERIZATION

Risk characterization quantitatively defines the magnitude of potential risks to endpoint species under a specific set of circumstances. It is the process of applying numerical methods and professional judgement to determine whether acceptable risk levels for endpoint species are or could be exceeded as a result of exposure to site-related CPECs. Risk characterization involves two components: a quantitative risk estimate and a narrative risk description. Because no one piece of information can adequately define risks to complex ecological systems, a formal "weight-of-evidence" approach might be needed to compile and integrate various types of evidence indicating the degree of risk present for each CPEC and assessment endpoint.

## 6.1. Risk Estimation Methodology

The acceptable risk level for sessile species such as plants and some soil invertebrates is a toxicity quotient (TQ) of less than or equal to 1. The TQ = EPV/EBV and the toxicity index (TI) =  $\Sigma$ TQ. TQs are added for a given receptor only when they have the same mode of action and same effect. The EBV for populations of sessile species is the LD50 for that chemical with respect to the endpoint species.

The acceptable risk level (ARL) for populations of ecological receptors is a 10 percent chance, or less, that 20 percent or less of the total local population would have an exposure point value greater than the EBV for each contaminant of concern. Once an EPV distribution and a contaminant-specific ecological benchmark value, either as a point value or a distribution, have been established for each endpoint species, computation of the acceptable risk level (ARL) numerical criterion involves the following:

• Estimate an endpoint species local population abundance.

- Estimate the probability of an individual of an endpoint species experiencing an exposure in excess of the benchmark or p(EPV > EBV).
- Estimate the number of individuals in a local population of an endpoint species likely to experience p(EPV > EBV) ≥ 10%, using a cumulative binomial distribution function.

## 6.1.1. Estimation of Local Population Abundance

Because the definition of acceptable risk for a population is based on effects to a certain percentage of individuals, it is necessary to estimate the number of individuals of each endpoint species within the local population boundary. For sessile terrestrial species (e.g., plants), the local population spatial boundary is assumed to be equal to the spatial boundaries of the locality of the facility. For sessile aquatic species (e.g., benthic invertebrates) in ponds or lakes within the locality of the facility, the local population spatial boundaries of the water body. For transient and migratory species, local population abundance is defined as the number of individuals utilizing habitat within spatial boundaries of the locality over the course of a year. Population abundance for mammals used in the risk assessment was taken from the literature (U.S. EPA, 1993; Causti et al 1997). Estimates of population abundance are found in Table 21.

# 6.1.2. Probability of Exposure Exceeding the Benchmark

In general, risk is the relationship between an unfavourable consequence and the probability associated with the occurrence of that unfavourable consequence. For our purposes the "unfavourable consequence" is an ecological response in a local population. The "probability associated with the occurrence of the unfavourable consequence" is



p(EPV > EBV). An ecological response occurs when toxicological responses in individuals of an endpoint species, as a consequence of long-term (chronic) exposure to a substance, results in the actual or projected loss of a minimum viable local population of that species (Newton, 1988). Aspects of individual health, viability, and performance are important only insofar as they might influence the sustainability of the local population.

Estimation of p(EPV > EBV) may be accomplished through the use of a normal density function (Suter et al, 1986). If EBV is expressed as a single point value and the mean and standard deviation of its natural logarithms define EPV, then the probability of EPV > EBV may be determined as follows:

$$p \cong \phi_Z \left( \frac{x_{EPV} - \ln(EBV)}{s_{EPV}} \right)$$

#### Equation 5

where:

p = Probability of EPV > EBV (unitless);

 φ<sub>Z</sub> = Cumulative distribution function of a standard normal random variable (NORMSDIST function in MS-Excel®);

 $x_{EPV}$  = Mean of natural logarithms of EPV (mg/[kg·d] or mg/L);

 $s_{EPV}$  = Standard deviation of natural logarithms of EPV (unitless); and

EBV = Single point value of EBV (mg/[kg·d] or mg/L).

The probability of EPV > EBV for the wildlife endpoint species deer mouse, and American kestrel was calculated for exposure to dioxins and furans in terms of the total 2,3,7,8-TCDD TEQ, in AOPC soils and sediment, plants, and/or soil and aquatic invertebrates. Using the total TEQ in this case is appropriate because dioxin furans and its congeners have the same mode of action and effect for each endpoint species,

respectively. The probability of EPV > EBV due to exposure to dioxins and furans in AOPC soils and sediment by deer mouse (0, very small) or the American kestrel (0, very small) is less than 10% (Tables 19 and 20).

## 6.1.3. Number of Individuals Affected

The number of individuals of an endpoint species within the local population boundary with  $a \ge 10\%$  chance of experiencing EPV > EBV was estimated using a cumulative binomial distribution function defined as (Barnthouse et al, 1995):

**Equation** 6

$$e = CRITBINOM(n, p, \alpha)$$

**Equation 7** 

$$r = CEILING(0.20 \times n, 1)$$

where:

$e$ = Number of individuals with $\geq 10\%$ chance of experiencing EP	1 22 .
(CRITBINOM function in MS Excel®);	
n = Total number of individuals within the local population bound	dary;
p = Probability of EPV > EBV (Equation 6);	
$\alpha$ = Probability of individual experiencing EPV > EBV (0.1); and	Ĺ
r = Twenty percent of individuals within the local population box	undary
(rounded-up to whole integer with CEILING function in MS	Excel®).

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Solving Equation (6) yields an estimate of the number of individuals within the local population boundary that have  $a \ge 10\%$  chance of encountering EPV > EBV. Solving Equation (7) provides an estimate of 20% of the total local population.

The number of individuals of the endpoint species deer mouse, racoon, and black-tail deer, respectively, within their respective local population boundary with  $a \ge 10\%$  chance of experiencing EPV > EBV was calculated for exposure to dioxins and furans in terms of the total 2,3,7,8-TCDD TEQ, in AOPC soils and sediment, plants, and/or soil and aquatic invertebrates. The number of individual of a species where EPV > EBP due to exposure to dioxins and furans in AOPC soils and sediment by deer mouse is zero and by the American kestrel is zero (Table 19 and 20).

Soil and sediment invertebrates and plants do not have the Ah receptor and are therefore are not believed to be susceptible to the potential toxic effects of dioxins and furans.

#### 6.2. Risk Description

This is a qualitative narrative discussion of risks presented by the site and must include a discussion of any toxicological and ecological factors beyond those embodied in the quantitative risk estimates. Risk must be described for each CPEC-pathway-receptor combination, i.e., for each assessment endpoint.

Dioxins and furans, ubiquitous pollutants, are present in the site soils and sediment at levels in excess of generally accepted background levels of 8 ppt. Dioxins and furans have large octanol water partition coefficients, are hydrophobic in nature, and can be absorbed into the lipids of plants, invertebrates, and wildlife where they may bioaccumulate. Their high octanol water partition coefficient, also makes them bind very tightly to soil and sediment particles and are very insoluble in water. Soil and aquatic
invertebrates and plants are not susceptible to the aryl hydrocarbon receptor (AhR) mediated toxic effects of dioxins and furans because they do not appear to have the Ah receptor. However, they do absorb dioxins and furans and transfer them to primary and subsequently secondary and tertiary consumers. Deer mice on the AOPC become exposed to TCDD via ingestion of soil invertebrates, plants, and incidental ingestion of soil. Mice birds have the Ah receptor and thus are susceptible to Ah receptor mediated toxic effects.

The total 2,3,7,8-TCDD TEQs in the soils (1.034 parts per billion (ppb)) and sediments (0.866 ppb) is less than the EBV for 2,3,7,8-TCDD in the deer mouse of 57 ppb (Table 22). Mice were caught on the AOPC for the purpose of sampling their tissues. Six mice were caught in mice trap in two days. This indicates that mice are likely using the site for there home. The home range of the deer mouse is much smaller than the size of the AOPC, consequently deer mice spend a significant portion of their time on the AOPC. The are no obvious effects to the mice caught and therefore no reason to believe that they are being negatively effected by exposures to CPECs on the site. The potential risk to the deermouse due to exposures to dioxin and furan like compounds at levels present at the AOPC are low.

The total 2,3,7,8-TCDD TEQs in the soils (1.034 ppb) and sediments (0.866 ppb) is less than the 2,3,7,8-TCDD LD50 of 1.63 ppb (Table 22). With the home range of the American kestrel being 300 acres, it likely spends only a small portion of its time on the 3.5 acre AOPC (Appendix B). The LD50 used for the American kestrel is based on embryo exposure to dioxin and furan like compounds. The preferred nest for the kestrel is a woodpecker hole or a natural cavity in a tree. There are no trees on the AOPC, therefore nesting and subsequent embryo exposures to dioxin and furan like compounds at levels present at the AOPC are unlikely. The potential risk to the American kestrel due to exposures to dioxin and furan like compounds at levels present at the AOPC are low.

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# 7. UNCERTAINTY ANALYSIS

# 7.1. CPEC Selection and Quantification

The selection of the CPEC in the AOPC was based on collection of two composite samples of 5 specimens each and 4 individual samples. Should the distribution of contamination in the AOPC be very heterogeneous the sampling of 15 locations may not adequately inform one of contamination present. However, the contamination in the AOPC is due to the spray of wastewater over the area in the past. The spray was spread over the entire AOPC and thus contamination is likely reasonably homogeneous. It is unlikely that any CPECs were missed. The data allows for a very good estimate of the mean concentration. The maximum detected value for each CPECs was used in the risk assessment. Level of uncertainty is considered to be low to medium.

# 7.2. Receptor Selection

The receptors selected were from a list of those that were identified on the AOPC, that were believed to be abundant in case tissue sampling was necessary, were likely to come into contact with the site soils, and that fell into the food web of the higher level consumers selected as endpoint receptors of concern. Level of uncertainty is considered to be low.

# 7.3. Exposure Estimation

The uptake of dioxins and furans by the deer mouse and by the American kestrel was based on literature values for food consumption, amount and type. The concentration of dioxins in their food was estimated using a mathematical model and using literature uptake values. Uptake of organic constituents can be effected by the level of organic carbon in the soil. The home range size used in the exposure estimation was based on literature values. The types of habitats that each animal prefers was taken from the literature. The abiotic media was sampled; however, no tissue samples were analysed. The models used were conservative and likely to over-estimate the uptake of constituents from abiotic media. The level of uncertainty may be moderate to high, however, it is unlikely that exposure has been underestimated.

# 7.4. Response Estimation

Ecological benchmark values were more often than not unavailable for the endpoint species CPEC. This meant using uncertainty factors to adjust the toxicity value being used for another species. The TEF for PCB 126 was used to convert the LD50 for PCB 126 to a 2,3,7,8-TCDD toxic equivalence (TCDD -EQ) in the American kestrel. The level of uncertainty is moderate. The uncertainty in EBV values is difficult to reduce without performing site specific toxicity testing.

# 7.5. Risk Estimation

The uncertainty in the risk estimation is directly effected by the uncertainty in the exposure estimation, CPEC concentration, and response estimation that were, moderate to high, moderate and moderate, respectively. In each case conservative assumptions were made. The uncertainty in the risk estimation is moderate to high however the likelihood that the risk was under estimated is low.

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# 8. CONCLUSION

The COIs that became CPECs are dioxins and furans with respect to mammals and birds. The probability of EPV > EBV due to exposure to dioxins and furans in AOPC soils and sediment by deer mouse (0, very small) or the American kestrel (0, very small) is less than 10%. Therefore, the number of deer mice or American kestrel, respectively, where EPV > EBP due to exposure to dioxins and furans in AOPC soils and sediment is zero for both. The level of acceptable risk to these species has not been exceeded. The risk for the deer mouse and the American kestrel due to exposures is low.

# 9. RECOMMENDATIONS

The CPECs in the AOPC are highly unlikely to present significant risk to soil invertebrates, plants, avian species, and small mammals and further ecological assessment is not recommended.

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# Table 1 Selected Endpoint Species

Endpoint Species	Trophic Level
American Kestrel (Falco sparverius) Deer mouse (Permyscus maniculatus) Earthworm Himilayan blackberry (Rubus discolor) Reed canary grass (Phalaris arundinacea	-



## **Contaminants of Intrest** SS98-12 COMP SS98-1-4COMP Maximum Minimum Mean 2/3/98 Detected Detected 10/7/98 (mg/kg) (mg/kg) (mg/kg) (mg/kg) (mg/kg) Polycyclic Aromatic Hydrocarbons Naphthalene < 0.1 < 0.1 < 0.1 < 0.1 0.12 0.05 0.12 0.05 0.085 Acenaphthylene Acenaphthene < 0.1 < 0.1 < 0.1 < 0.1 0.011 0.041 0.011 0.041 0.026 Fluorene Phenanthrene 0.025 0.019 0.025 0.019 0.022 Anthracene 0.2 0.06 0.2 0.06 0.13 0.039 0.039 0.039 0.039 0.039 Fluoranthene 0.044 0.025 0.044 0.025 0.0345 Ругепе Benzo(a)anthracene 0.11 0.11 0.11 0.11 0.11 0.063 0.14 0.063 0.1015 0.14 Chrysene 0.033 0.0715 Benzo(b)fluoranthene 0.033 0.11 0.11 Benzo(k)fluoranthene 0.15 0.2 0.2 0.15 0.175 Benzo(a)pyrene 0.078 0.11 0.11 0.078 0.094 0.1045 0.089 0.089 0.12 Indeno(1,2,3,-cd)pyrene 0.12 0.058 0.025 0.058 0.025 0.0415 Dibenzo(a,h)anthracene Benzo(g,h,i)perylene 0.11 0.13 0.13 0.11 0.12 Phenols Phenol < 1 < 0.5 < 0.5 2-Chlorophenol < 1 < 0.5 2-Nitrophenol <1 2,4,-Dimethylphenol <1 < 0.5 < 0.5 2,4-Dichlorophenol <1 2,6-Dichlorophenol <1 < 0.5 < 0.5 4-Chloro-3-methylphenol <1 2,4,6,-Trichlorophenol < 1 < 0.5 < 0.5 2,4,5-Trichlorophenol <1 < 5.0 2,4-Dinitophenol <10 4-Nitrophenol <1 < 0.5 2,3,5,6-Tetrachlorophenol < 1.0 <2 2,3,4,6-Tetrachlorophenol <2 < 1.0 2-Methyl-4,6-dinitrophenol <1 < 0.5 Pentachlorophenol <1 < 0.5 Metals Arsenic (Total) 36 73.7 73.7 36 54.85 49.9 49.9 46 47.95 Copper (Total) 46 54.6 62.95 Chromium (Total) 71.3 54.6 71.3 199.9 199.9 128 163.95 Zinc (Total) 128

# Table 2 Composite Data and Summary Statistics Media: Soil

Contaminants of Intrest	SS98-12 COMP 10/7/98	SS98-1-4COMP 2/3/98	Maximum Detected	Minimum Detected	Mean
	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)
Dioxins/Furans <sup>#</sup>					
2378-TCDD	8.7	< 50	< 50	8.7	16.9
1,2,3,7,8-pentachlorodibenzo-p-dioxin (PeCDD) 1,2,3,4,7,8-hexachlorodibenzo-p-dioxin (HxCDD)	42.7 219	< 90	< 90	42.7	43.9
1,2,3,6,7,8-hexachlorodibenzo-p-dioxin (HxCDD)	1070	< 310 410	219 1070	< 310	192.0
1,2,3,7,8,9-hexachlorodibenzo-p-dioxin (HxCDD)	335	<270	335	410 < 270	740.0 235.0
1,2,3,4,6,7,8-heptachlorodibenzo-p-dioxin (HpCDD)	25770	7940	25770	7940	16855.0
octachlorodibenzo-p-dioxin (OCDD)	412130	160370	412130	160370	286250.0
2,3,7,8-tetrachlorodibenzofuran (TCDF)	22.7	<30	22.7	<30	18.9
1,2,3,7,8-pentachlorodibenzofuran (PeCDF)	55.6	< 50	55.6	< 50	40.3
2,3,4,7,8-pentachlorodibenzofuran (PeCDF)	38.6	< 50	38.6	< 50	31.8
1,2,3,4,7,8-hexachlorodibenzofuran (HxCDF)	343	< 190	343	< 190	219.0
1,2,3,6,7,8-hexachlorodibenzofuran (HxCDF)	112	< 160	112	< 160	96.0
2,3,4,6,7,8-hexachlorodibenzofuran (HxCDF)	171	< 190	171	< 190	133.0
2,3,7,8,9-HxCDF	13.1	< 210	< 210	13.1	59.1
,2,3,4,6,7,8-heptachlorodibenzofuran (HpCDF) ,2,3,4,7,8,9-heptachlorodibenzofuran (HpCDF)	6750	2080	6750	2080	4415.0
,2,3,4,7,0,9-nepractionorodinenzoruran (RpCDF) octachlorodibenzofuran (OCDF)	372 12730	< 420 7740	372 12730	< 420 7740	291.0 10235.0

# Table 2 Composite Data and Summary Statistics Media: Soil

a = Dioxins/Furans in ng/kg

KEYSTONE ENVIRONMENTAL

Contaminants of Intrest	SD98-6 (mg/kg)
· ·	
Polycyclic Aromatic Hydrocarbons	
Naphthalene	ND
Acenaphthylene	0.052
Acenaphthene	
Fluorene	ND
Phenanthrene	0.016
Anthracene	0.1
Fluoranthene	0.027
Pyrene	0.024
Benzo(g,h,i,)perylene	0.027
Chrysene	0.015
Benzo(a)anthracene	0.04
Benzo(b)fluoranthene	0.1
Benzo(k)fluoranthene	0.1
Benzo(a)pyrene	0.013
Dibenzo(a,h)anthracene	0.01
Indeno(1,2,3-cd)pyrene	0.027
Phenols	
Phenol	< 0.5
2-Chlorophenol	< 0.5
2-Nitrophenol	< 0.5
2,4,-Dimethylphenol	< 0.5
2,4-Dichlorophenol	< 0.5
2,6-Dichlorophenol	< 0.5
4-Chloro-3-methylphenol	< 0.5
2,4,6,-Trichlorophenol	< 0.5
2,4,5-Trichlorophenol	< 0.5
2,4-Dinitophenol	< 5.0
4-Nitrophenol	< 0.5
2,3,5,6-Tetrachlorophenol	< 1.0
2,3,4,6-Tetrachlorophenol	< 1.0
2-Methyl-4,6-dinitrophenol	< 0.5
Pentachloropheno1	< 0.5

# Table 3 Composite Data and Summary Statistics Media: Soil

Contaminants of Intrest	SD98-6 (mg/kg)
Metals	
Arsenic	58.9
Chromium(III)	40.8
Chromium(VI)	40.8
copper	84.2
zinc	288
Dioxins/Furans <sup>a</sup>	
2378-TCDD	12,9
1,2,3,7,8-pentachlorodibenzo-p-dioxin (PeCDD)	96.7
1,2,3,4,7,8-hexachlorodivenzo-p-dioxin (HxCDD)	556
1,2,3,6,7,8-hexachlorodibenzo-p-dioxin (HxCDD)	1360
1,2,3,7,8,9-hexachlorodibenzo-p-dioxin (HxCDD)	668
1,2,3,4,6,7,8-heptachlorodibenzo-p-dioxin (HpCDD)	25830
octachlorodibenzo-p-dioxin (OCDD)	173880
2,3,7,8-tetrachlorodibenzofuran (TCDF)	14.4
1,2,3,7,8-pentachlorodibenzofuran (PeCDF)	33.2
2,3,4,7,8-pentachlorodibenzofuran (PeCDF)	34.4
1,2,3,4,7,8-hexachlorodibenzofuran (HxCDF)	209
1,2,3,6,7,8-hexachlorodibenzofuran (HxCDF)	129
2,3,4,6,7,8-hexachlorodibenzofuran (HxCDF)	236
1,2,3,7,8,9-HxCDF	12.9
1,2,3,4,6,7,8-heptachlorodibenzofuran (HpCDF)	2890
1,2,3,4,7,8,9-heptachlorodibenzofuran (HpCDF)	181
octachlorodibenzofuran (OCDF)	4870

# Table 3 Composite Data and Summary StatisticsMedia: Soil

.

a = Dioxins/Furans in ng/kg



Table 4 Calculated Composite Concentration for Phenols in Soil Sampled February 3, 1998

Contaminant of Interest	SS98-1	SS98-2	S-86SS	SS98-4	Maximum	Minimum
	2/3/98	2/3/98	2/3/98	2/3/98	Detected	Detected
	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)
Phenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2-Chlorophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2-Nitrophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2.4Dimethylphenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2.4-Dichlorophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2.6-Dichlorophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
4-Chloro-3-methylphenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2.4.6Trichlorophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2.4.5-Trichlorophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2,4-Dinitophenol	< 5.0	< 5.0	< 5.0	< 5.0	< 5.0	< 5.0
4-Nitrophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
2.3.5.6-Tetrachlorophenol	< 1.0	< 1.0	<1.0	< 1.0	< 1.0	< 1.0
2.3.4.6-Tetrachlorophenol	< 1.0	< 1.0	< 1.0	< 1.0	< 1.0	< 1.0
2-Methyl-4.6-dinitrophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5
Pentachlorophenol	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5

Table 5 Calculated Composite Concentration for Metals in Soil Sampled February 3, 1998

Contaminant of Interest	SS98-1	SS98-2	SS98-3	SS98-4	Maximum	Minimum	Mean	UCL95
	2/3/98	2/3/98	2/3/98	2/3/98	Detected	Detected		(SS98-1-4COMP)
	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg) (mg	(mg/kg)	(mg/kg)
Arsenic (Total)	80.5	72.7	13.8	43.6	80.5		52.65	
Copper (Total)	62.8	44.6	41.6	38.5	62.8		46.875	
Chromium (Total)	38.8	21.7	61.9	27.7	61.9		37.525	
Zinc (Total)	231	176	110	114	231		157.75	

Table 6

Calculated Composite Concentration for Polycyclic Aromatic Hydrocarbons in Soil Sampled February 3, 1998

Contaminant of Interest	SS98-1	SS98-2	SS98-3	SS98-4	Maximum	Minimum	Меап	UCL95
	2/2/98	2/2/98	2/2/98	2/2/98	Detected	Detected		(SS98-1-4 COMP)
	(me/ke)	(me/ke)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)
Manhthalana	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1		< 0.1
A concuptivulance	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1		0.05
A constructions	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1		<0.1
Fluctuation	< 0.01	0.01010	0.0105	0.0104	0.0104	0.005	0.0077	0.011
Dhenanthrene	0.021	0.011	0.017	0.017	0.021	0.011	0.0165	0.019
	0.058	0.021	0.055	0.055	0.058	0.021	0.04725	0.06
Tillua aveile Tilluanathane	0.039	0.025	0.038	0.038	0.039	0.025	0.035	0.039
	< 0.05	< 0.05	< 0.05	< 0.05	< 0.05	< 0.05	<u></u>	0.025
rylene Darrofolouthrocene	0.04	0-027	0.115	0.115	0.115	0.027	0.07425	0.11
	0.032	0.027	0.066	0.066	0.066	0.027	0.04775	0.063
Curysourc Dearcollyfinoranthene	0.075	0.051	0.213	0.213	0.213	0.051	0.138	0.11
Deuro(U)filoranthene	0.039	0.032	0.115	0.115	0.115	0.032	0.07525	0.2
Danzo(c)nimane	0.025	0.023	0.094	0.094	0.094	0.023	0.059	0.11
Indano(1 ) 2 . adhurane	0.074	0.052	0.212	0.212	0.212	0.052	0.1375	0.089
Titon that the there are a second to the second sec	< 0.05	< 0.05	< 0.05	< 0.05	< 0.05	< 0.05		0.025
Dubenzo(a, i)nervlene	0.047	0.035	0.142	0.142	0.142	0.035	0.0915	0.13
Benzo(g,n,1)perylene	1 th O.O	0.00 C						

Constituent	Terrestrial Receptor	Endpoint	Ltierature Toxicity Value (mg/kg)	SBV (mg/kg)	Reference
· · · · · · · · · · · · · · · · · · ·		<u></u>	(		
Acenaphthylene	terrestrial	NOEC	95	6	· a
Fluorene	terrestrial	NOEC	95	· 6	a
Phenanthrene	terrestrial	NOEC	95	6	Denneman and van Gestral, 1990
Anthracene	terrestrial	NOEC	95	6	a
Fluoranthene	terrestrial	NOEC	95	6	a
Pyrene	terrestrial	NOEC	2500	156	Ь
Benzo(g,h,i,)perylene	terrestrial	· NOEC	2500	156	b .
Chrysene	terrestrial	NOEC	2500	156	ь
Benzo(a)anthracene	terrestrial	NOEC	2500	156	Ъ
Benzo(b)fluoranthene	terrestrial	NOEC	2500	156	b
Benzo(k)fluoranthene	terrestrial	NOEC	2500	156	ъ
Benzo(a)pyrene	terrestrial	NOEC	2500	156	Denneman and van Gestral, 1990
Dibenzo(a,h)anthracene	terrestrial	NOEC	2500	156	b
Indeno(1,2,3-c,d)pyrene	terrestrial	NOEC	2500	156	b
Benzo(b+k)fluoranthene	terrestrial	NOEC	2500	156	b
Indeno(1,2,3-c,d)pyrene	terrestrial	NOEC	2500	156	b

Table 7. Screening Benchmark Value (SBV) for Soil

NOEC (no-observed-effect-concentration) - the highest concentration of a test material to which organisms are exposed that does not cause any observed or statistically significant effect to the organism.

a - value represents NOEC for phenanthrene and is used as surrogate

b - value represents NOEC for benzo(a)pyrene and is used as surrogate

Denneman, C.A.J. and van Gestral, C.A.M., 1990. Soil Contamination and Soil Ecosystems: Proposal for C- (test) Values Based on Ecotoxicological Risk, National Institute of Public Health and Environmental Protection, RIVM, the Netherlands, April, 1990



Table 8 A Comparison of 50 Times the Screening Benchmark Value to the Concentration of Polycyclic Aromatic Hydrocarbons (PAHs) in Soil for the Selection of Contaminants of Potential Ecological Concern (CPEC)

x 50 (mg/kg) (mg/kg) (x 50 (mg/kg) (mg/kg) ( 6 6 6 6 6 6 6 6 6 6 6 156 156 156 156 1	SBV <sup>*</sup> plants SBV plants SBV inverts <sup>b</sup> SBV inverts	SBVbirds	SBVbirds	<b>SBV</b> mammals	SBVmammals	Soil	CPEC"
(mg/kg)         (mg/kg)         (mg/kg)         (mg/kg)         (mg/kg)         (mg/kg)         (           ne         6         6         6         6         1500         1500         1500           e         6         6         6         6         6         6         1560         1560         1560         1560         1560         156<	x 50		x 50		· x 50	Concentration	
ne 6 6 6 1500 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6	(mg/kg) (mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	
ne 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6							
anthracence     156     30     1500       6     6     6     6       6     6     6     6       6     156     156     156       156     156     156     156       anthene     156     156     156       anthene     156     156     156       anthracene     156     156     156       ne     7     156     156		6		9		0.12	°Z
6     6       6     6       6     6       6     6       6     6       6     156       156     156       156     156       156     156       156     156       anthene     156       156     156       anthene     156       156     156       156     156       156     156       156     156       156     156       156     156	30 1500	6		9		0.041	°N N
berylene     6     6       6     6     6       6     156     156       156     156     156       anthene     156     156       ne     7     156		9		9		0.025	0N0
berylene     156     6       156     156     156       156     156     156       anthene     156     156       ne     7     156		6		6		0.2	No
berylene     156     156       156     156     156       156     156     156       anthene     156     156       anthene     156     156       anthene     156     156       anthene     156     156       ne     7     156       ne     7     156		9		6		0.039	No
k,hi,)perylene 156 156 e 156 156 156 anthracene 156 156 156 Aluoranthene 156 156 156 Aluoranthene 156 156 An honthracene 156 156 156 156 156		156		156		0.044	°N
156 156 156 156 156 156 156 156 156 156		156		156		0.11	°Z
1.20 156 156 156 156 156 156 156 156		156		156		0.14	٩
1.00 1.56 1.56 1.56 1.56 1.56 1.56 1.56		156		156		0.11	Ño
156 156 156 156 156 156 156		156		156		0.2	No
156 156 156 156 156 156 156 156 156 156		156		156		0.11	No
inscene 156 156				7	350	0.153	°N
				156		0.058	°N
		120		156	-	0.13	No
_		001		1.00			2

There are no screening benchmark values for PAHs in soil except for benzo(a)pyrene with respect to mammals and Fluoroanthene with respect to soil invertebrates therefore PAHs will be carried forward int the risk assessment with respect to all but these endpointsthese endpoints

a - SBV = Screening Benchmark Value

b - inverts = invertebrates

\* Benzo(k)fluoranthene sediment SBV used as surrogate for Benzo(b)fluoranthene EPV - Exposure Point Value

Sample ID Date Sampled	Soil Concentration (mg/kg)	CPEC
Phenol	<1	no
2-Chlorophenol	<1	no
2-Nitrophenol	<1	no
2,4,-Dimethylphenol	< 1	no
2,4-Dichlorophenol	<1	no
2,6-Dichlorophenol	< 1	no
4-Chloro-3-methylphenol	< 1	no
2,4,6,-Trichlorophenol	<1	no
2,4,5-Trichlorophenol	<1	no
2,4-Dinitophenol	< 10	no
4-Nitrophenol	< 1	no
2,3,5,6-Tetrachlorophenol	<2	no
2,3,4,6-Tetrachlorophenol	<2	no
2-Methyl-4,6-dinitrophenol	<1	no
Pentachlorophenol	<1	no

Table 9. A Comparison of 50 Times the Screening BenchmarkValue to the Concentrations of Phenols in Soil for the Selectionof Contaminants of Potential Ecological Concern (CPEC)



Table 10. A Comparison of 50 Times the Screening Benchmark Valueto the Concentration of Selected Metals in Soilfor the Selection of Contaminants of Potential Ecological Concern

Contaminants		SBV <sup>a</sup> plants	SBV <sup>a</sup> plants SBV plants	<b>SBV</b> inverts <sup>b</sup>	SBVinverts	SBVbirds	SBVbirds	SBVmammals	SBVmammals	Soil	CPEC
	Background		× 50		x 50		x 50		x 50	Concentration	
	)	(mg/kg) <sup>c</sup>	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	
Arcanin	10	01	500	60	3000	126	13000	80	400	73.7	<sup>N</sup> o
	25	2 -	9	40	20			33000	165000		No
	10	-	2		ì	40	2450	30	1500		No
Chromium(VI)						f :	0010	\$	0000		N.S.
Conner	41	100	5000	20	2500	605	00005	184	0076		
Zinc	104	50	2500	200	10000	1284	64200	3800	190000	199.9	No
a - SBV = Screening Benchmark Value	ning Benchma	urk Value									

a - 5B v = 5 creening Bench b - inverts = invertebrates

c - mg/kg = milligrams per kilogram EPV - Exposure Point Value

for the Selection of Contaminants of Potential Ecological Concern (CPEC) Table 11. A Comparison of 50 Times the Screening Benchmark Value to the Concentration of Dioxin/Furan and Congeners in Soil

CPEC Υc Y cs Yes Yes Yes Yes Yes ŝ Ϋ́cs 8 **ICDD-EQ** og/kg(ppt) 412.13 034.49 257.7 21.35 21.9 2.27 2.78 34.3 11.2 67.5 3.72 12.73 33.5 19.3 17.1 1.31 107 8.7 Concentration pg/kg(ppt) 8.7 42.7 219 1070 335 25770 412130 22.7 55.6 38.6 13.1 6750 372 12730 343 112 171 Soil SBVmammals x 50 pg/kg(ppt) 5000 100000 00000 5000 50000 50000 10000 5000 50000 50000 50000 5000 5000 5000 5000 500 SBVmammafs pg/kg(ppt) 100<sup>d</sup> 200<sup>c</sup> 100<sup>d</sup> 100<sup>d</sup> 100<sup>d</sup> 100<sup>d</sup> 1000<sup>d</sup> 2000° 100<sup>d</sup> 100<sup>d</sup> 1000<sup>d</sup> 1000<sup>d</sup> 20<sup>4</sup> SBVbirds x 50 pg/kg(ppt) 500000 100000 500000 50000 500000 50000 000000 10000 50000 50000 50000 10000 50000 50000 50000 400 5000 SBV<sup>b</sup>birds pg/kg(ppt) 1000<sup>4</sup> 10000<sup>4</sup> 100000<sup>4</sup> 1000<sup>d</sup> 10000<sup>d</sup> 10000<sup>d</sup> 2060<sup>d</sup> 1000 10004 1000 1000 200<sup>d</sup> 1000<sup>4</sup> 100° 200<sup>d</sup> **‰** I-TEF 0.001 0.01 0.01 0.1 0.01 0 0.05 ŝ 0.1 0.1 0.1 0.1 100.0 0.1 0.1 0.5 ,2,3,4,6,7,8-heptachlorodibenzo-p-dioxin (HpCDD) ,2,3,4,7,8-hexachlorodibenzo-p-dioxin (HxCDD) ,2,3,6,7,8-hexachlorodibenzo-p-dioxin (HxCDD) ,2,3,7,8,9-hexachlorodibenzo-p-dioxin (HxCDD) .2,3,7,8-pentachlorodibenzo-p-dioxin (PeCDD) ,2,3,4,6,7,8-heptachlorodibenzofuran (HpCDF) ,2,3,4,7,8,9-heptachlorodibenzofuran (HpCDF) ,3,4,6,7,8-hexachlorodibenzoftıran (HxCDF) ,2,3,4,7,8-hexachlorodibenzofuran (HxCDF) ,2,3,6,7,8-hexachlorodibenzofuran (HxCDF) ,2,3,7,8-pentachlorodibenzofuran (PeCDF) ,3,4,7,8-pentachlorodibenzofuran (PeCDF) ,3,7,8-tetrachlorodibenzofuran (TCDF) ctachlorodibenzo-p-dioxin (OCDD) octachlorodibenzofuran (OCDF) Contaminants of Intrest 1,2,3,7,8,9-HxCDF **Total TCDD-EQ** 1378-TCDD

Plants and soil invertebrates do not have the Ath receptor and are thus believed to have a relatively low toxicity with respect to dioxin and dioxin like compounds. a - I-TEF = International Toxicity Equivalents for mammals

c - SBVs taken for the "Guidance for Ecological Risk Assessment Level II Screening Benchmark Values (Origon Department of Environmental Quality, 1998) b - SBV = Screening Benchmark Value

d - SBVs for dioxin like congeners derived by multiplying the SBV for 2,3,7,8-TCDD by its I-TEF

EPV - Exposure Point Value

**KEYSTONE** ENVIRONMENTAL

Contaminants of Intrest	SBV <sup>b</sup> (mg/kg)	SBV x 10	Sediment Concentration (mg/kg)	CPEC
Manhthalana	0.032	0.32	ND	No
Naphthalene Acenaphthylene	0.33	3.3	0.052	No
Fluorene	0.034	0.34	ND	No
Phenanthrene	0.56	5.6	0.016	No
Anthracene	0.032	0.32	0.1	No
Fluoranthene	0.064	0.64	0.027	No
Pyrene	0.57	5.7	0.024	No
Benzo(g,h,i,)perylene	0.29	2.9	0.027	No
Chrysene	0.5	5	0.015	No
Benzo(a)anthracene	0.26	2.6	0.04	No
Benzo(b)fluoranthene	0.24	2.4*	0.1	No
Benzo(k)fluoranthene	0.24	2.4	0.1	' No
Benzo(a)pyrene	0.35	3.5	0.013	No
Dibenzo(a,h)anthracene	0.23	2.3	0.01	No
Indeno(1,2,3-cd)pyrene	0.078	0.78	0.027	No

# Table 12. A Comparison of 10 Times the Screening Benchmark Value to the Concentration of Polycyclic Aromatic Hydrocarbons (PAHs) in Sediment for the Selection of Contaminants of Potential Ecological Concern (CPEC)

\* Benzo(k)fluoranthene sediment SBV used as surrogate for Benzo(b)fluoranthene

a - SBV = Screening Benchmark Value

Sample ID	Sediment	CPEC
-	Concentration	
Date Sampled	(mg/kg)	
Phenol	< 0.5	no
2-Chlorophenol	< 0.5	no
2-Nitrophenol	< 0.5	no
2,4,-Dimethylphenol	< 0.5	no
2,4-Dichlorophenol	< 0.5	no
2,6-Dichlorophenol	< 0.5	no
4-Chloro-3-methylphenol	< 0.5	no
2,4,6,-Trichlorophenol	< 0.5	no
2,4,5-Trichlorophenol	< 0.5	no
2,4-Dinitophenol	< 5.0	no
4-Nitrophenol	< 0.5	, no
2,3,5,6-Tetrachlorophenol	< 1.0	no
2,3,4,6-Tetrachlorophenol	< 1.0	no
2-Methyl-4,6-dinitrophenol	< 0.5	no
Pentachlorophenol	< 0.5	no

# Table 13. A Comparison of 10 Times the Screening Benchmark Value to the Concentrations of Phenols in Sediment for the Selection of Contaminants of Potential Ecological Concern (CPEC)

Table 14. A Comparison of 10 Times the Screening Benchmark Valueto the Concentration of Selected Metals in Sedimentfor the Selection of Contaminants of Potential Ecological Concern (CPEC)

Contaminants of Intrest	SBV <sup>b</sup> (mg/kg)	SBV x 10	Sediment Concentration (mg/kg)	CPEC
Arsenic	12	120	58.9	No
Chromium(III)	56	2800	40.8	No
Chromium(VI)			40.8	
conner	28	1400	84.2	No
zinc	159	79500	288	No

a - SBV = Screening Benchmark Value

c - mg/kg = milligrams per kilogram

Table 15. A Comparison of 10 Times the Screening Benchmark Value to the Concentration of Dioxin/Furan and Congeners in Sediment for the Selection of Contaminants of Potential Ecological Concern (CPEC)

of Intrest Dioxin/Furan 2378-TCDD 1,2,3,7,8-pentachlorodibenzo-p-dioxin (PeCDD) 1,2,3,6,7,8-hexachlorodibenzo-p-dioxin (HxCDD) 1,2,3,6,7,8-hexachlorodibenzo-p-dioxin (HxCDD) 1,2,3,7,8,9-hexachlorodibenzo-p-dioxin (HxCDD) 0.1 1,2,3,7,8,9-hexachlorodibenzo-p-dioxin (HxCDD) 0.1 0.1	Concentration (ppt)		
un chlorodibenzo-p-dioxin (PeCDD) achlorodibenzo-p-dioxin (HxCDD) achlorodibenzo-p-dioxin (HxCDD) achlorodibenzo-p-dioxin (HxCDD)		(pot)	
chlorodibenzo-p-dioxin (PeCDD) achlorodibenzo-p-dioxin (HxCDD) achlorodibenzo-p-dioxin (HxCDD) achlorodibenzo-p-dioxin (HxCDD)			
chlorodibenzo-p-dioxin (PeCDD) achlorodibenzo-p-dioxin (HxCDD) achlorodibenzo-p-dioxin (HxCDD) achlorodibenzo-p-dioxin (HxCDD)	12.9	12.9	Yes
	96.7	48.35	Yes
 		55.6	Yes
 	1360	136	Yes
â	668	66.8	Yes
	1 25830	258.3	Yes
octachlorodibenzo-p-dioxin (OCDD)	1 173880	173.88	Yes
2.3.7.8-tetrachlorodibenzofuran (TCDF) 0.1	14.4	1.44	Yes
1.2.3.7.8-pentachlorodibenzofuran (PeCDF) 0.05		1.66	Yes
2.3.4.7.8-pentachlorodibenzofuran (PeCDF) 0.5		17.2	Yes
1.2.3.4.7.8-hexachlorodibenzofuran (HxCDF) 0.1		20.9	Yes
1.2.3.6.7,8-hexachlorodibenzofuran (HxCDF) 0.1	129	12.9	Yes
2.3.4.6.7.8-hexachlorodibenzofuran (HxCDF) 0.1		23.6	Yes
1.2.3.7.8.9-HxCDF 0.1		1.29	Yes
1 2 3 4 6 7 8-hentachlorodibenzofuran (HpCDF) 0.01	1 2890	28.9	Yes
1 2 3 4.7.8.9-hentachlorodibenzofuran (HpCDF) 0.01	1 181	1.81	Yes
Actachlorodibenzoftran (OCDF) 0.001	11 4870	4.87	Yes
Total		866.4	

a - I-TEF = International Toxicity Equivalents for mammals b - SBV = Screening Benchmark Value

Table 16 Literature Soil to Plant, Sediment to Invertebrate, and Small Mammal Uptake factors

Chemical	Soil-to-Plant Sediment-to- Wet Uptate Invertebry	Sediment-to- Invertebrate Uptake	Log Kow	Small Mammal Uptake	Molecular Reference Weight	Reference
HpCDD, 2,3,7,8-	2.40E-04	· · ·		-	425.31	B
ocdd	6.20E-05					63
OCDF	7.90E-05					63
PeCDD, 2,3,7,8-	1.30E-03					63
PeCDF, 1,2,3,7,8-	8.90E-04					69
PeCDF, 2,3,4,7,8-	7.50E-04					, tə
TCDD, 2,3,7,8-	8.80E-04	0.99	6.54	1.77	322	a, D, C, G
TCDF, 2,3,7,8-	3.20E-03					69

Log Kow - Logrithim of the octanol-water partition coefficient

a - Toxicity and Chemical specific factors Data Base (Rrisk Assessment Program Oak Ridge National Laboratory

b - Sample et al Development and Bioaccumulation Model for Earthworms ES/ER/TM-200 ORNL Oak Ridge, Tenn.
 c - Suter G.W. 1997. Guidance For Ecological Risk Estimation Methods

d - Sample et al, 1998 Development and validation of Bioaccumulation Models for Small Mammals

Table 17 The Earthworm Bioaccumulation Factor (BAF)



BCF	3.467E+03	
Рмогш	8.400E-01 1.000E-02 3.467E+06 1.000E+03 3.467E+03	3388441.56
Kow	3.467E+06	, k_m
Ffat	1.000E-02	$\frac{k_1}{Kp} + k_f$ $\frac{k_2}{k_2 + k_g + k_m}$
Fwater	8.400E-01	$BAF = \frac{1}{k}$
Chemical	2,3,7,8 -TCDD	

2.95E+01 BAF 3.47E+03 BCF 0.00E+00 k<sub>m</sub> 5.80E-03 يى ا 1.01E-04  $\mathbf{k}_2$ 2.50E+00 Kp 3.40E-02 **k**f 3.50E-01 Ā 2,3,7,8 -TCDD Chemical



Table 18 Foraging Home Range

Species	Foraging Home Range	FHR Diameter (m)	Home Range FHR Diameter (m) Local Population Boundary Preferred Habitat Type Area (m <sup>2</sup> )	Preferred Habitat Type	Area (m²)
4	$(A_{\rm h}) ({\rm m}^2)$		Diameter (D <sub>h</sub> ) (m <sup>2</sup> )		
Deer morree	006	34	0/1	Any	1.88E+05
American Kestrel	12	1236	6180	grasslands, deserts, junipe	1.28E+08
				woodlands, meadows	

	Area #1	Area #2	Area #3	Area #4
Endpoint Species > Deer Mouse	grass/shrubs	Intermittent Pond	d grass	gass/storage
Lahitat (L)	1	2	4	ŝ
	0.75	0.75	0.75	0.25
	0,300	0.300	0.300	0.100
Relative Habitat Quality (Inqu)	12138	2023	66714	38243
Habitat Area (metresz)				119118
Total Area (metres/)	1.034E-03	8.660E-04	2.000E-05	2.000E-05
Geometric Mean of Soul Concentration (Culty)	5.120E-04	5.120E-04	1.000E-05	1,000E-05
Standard Deviation of Soli Concentrations	9.099E-07	7.621E-07	1.760E-08	1.760E-08
Plant Concentration (Dassed on uptake factor = 0.00-1)	3.05E-02	2.55E-02	5.90E-04	5.90E-04
Invertebrate Concentration (Dased on uptake Lactor			1	1
Area use Factor (AU)	• -	0.4	1	
Seasonality factor	2.200E-02	2.200E-02	2.200E-02	2.200E-02
Body Wergnt (kingrams) Intake Rate (kg/d)	4.180E-03	4,180E-02	4,180E-03	4.180E-03
	an 1703 b	50-2055 C	4.873E-06	9.311E-07
Exposure Point Value (EPV) (mg/kg/d)	נטרתנסנ.			7.723E-05
Total Area (mg/kg/d) Natural Logarithm of EPV	066.6-	-10.573	-12,232	-13.887
				-11.671
Mean of Natural Logarithm of EPV area of concern Standard Deviation of Mean of Natural Logarithm of EPV (calculated using Crystal Ball)	ing Crystal Ball)			0.204
p(EPV>EBV) for area of concern and associated areas				0.0E+00
Number of Individuals With > 10% Chance of EPV > EBV				0.0015+00
Twenty Percent of Individuals Within the Local Population Boundary				÷

	Area #1 grass 1	Area #2 wetland	Area #3 Industrial	Area #4 Commercial	Area #5 Residential	Area #6 Vacant	
Endpoint Species > American Kestrel							
	-	2	4	4	4	5	
Habitat (k)	0.75	0.75	0.5	0.5	0.25	0.75	
Habitat Quality	001.0	0.300	005.0	0.300	0.300	0.100	
Relative Habitat Quality (Hqk)	85121	2023	2165788	11160325	35485358	35669659	
Habitat Area (metres2)						84495291	
Total Area (metres2)	1.03E-03	8.66E-04	3.200E-05	3.200E-05	3.200E-05	3.200E-05	
Geometric Mean of Soil Concentration (c.j.w)	5.120E-04	5,120E-04	1.000E-05	1.000E-05	1.000E-05	1.0005-05	
Standard Deviation of Soli Concentrations	3 050E-02	2.55E-02	9.440E-04	9.440E-04	9.440E-04	9.440E-04	
Invertebrate (based on uptake factor = 23.2)	5.10E-05	5.10E-05	5.105-05	5.10E-05	5.10E-05	5.10E-05	
Small mammai (pased on uptake tactor - 2007)	1.446-04	2.39E-05	2.56E-02	1.32E-01	4.20E-01	4.22E-01	
Area use Factor (AU)	1	1	-	1	-	I	
Seasonality factor	0.1200	0.1200	0.12	0.12	0.12	0.12	
Body Weight (kliograms) Intake Rate (kg/d)	0.035	0.035	0.035	0.035	0,035	0.035	
		C1 7623 2	, 108E-08	6 634E-07	6.707E-06	2.259E-06	
Exposure Point Value (EPV) (mg/kg/d)	11-3//57	61-3ccr.c				9.654E-06	
Total Area (mg/kg/d)			343 61	ACC 11-	-11.912	-13.001	
Natural Logarithm of EPV	-24.463	-28.223	cnc·/ 1+				
						-18.222	
Mean of Natural Logarithm of EPV area of concern Standard Deviation of Mean of Natural Logarithm of EPV (calculated using Crystal Ball)	stal Ball)					0.152	
p(EPV>EBV) for area of concern and associated areas						0.02+00	

Table 20: American Kestrel Results of Exposure Model Calculations for Dioxin

0

Number of Individuals With > 10% Chance of EPV > EBV Twenty Percent of Individuals Within the Local Population Boundary

# Table 21 Population Abundance

Habitat	¥	Area (ha)	k Area (ha) Area (m')		Endpoint Density (#/ha) Estimated Species Abundance	Estimated Abundance
ornaes/shruth	-	1.214	12138	Deer Mouse	12.7 - 45.5	18 - 64
grass an uc intermittent nond	2	0.202	2023	Deer Mouse	12.7 - 45.5	4 - 13
unvanuevan pour		6.671	66714	Deer Mouse	12.7 - 45.5	85 - 304
grassismuu 1 U. Rovter active site	4	5.956	59561	Deer Mouse	12.7 - 45.5	76 - 271
grass/storage	Ś.	3.824	38243	Deer Mouse	12.7 - 45.5	49 - 174
		1.214	12138	American Kestrel	0.0005 - 0.0012	0-0
grassi suu uo intermittent nond	5	0.202	2023	American Kestrel	0.0005 - 0.0012	0-0
	. "	216.579	2165788	2165788 American Kestrel	0.0005 - 0.0012	0-1
	14	1116.032	11160325	1160325 American Kestrel	0.0005 - 0.0012	1-1
Donidantrial (78,0%)*	Ŷ	3548 536	35485358	35485358 American Kestrel	0.0005 - 0.0012	2-4
Vacant (29%)*	9	3566.966	35669659	35669659 American Kestrel	0.0005 - 0.0012	2-4
			-			

building and/or paved; and 25% of residential land was building and/or paved. Paved areas buildings, active industrial sites, roads, and rail is considered to have a habitat rating of 0 and was not included. 32% of the \*The the areas by percentage for each area as communicated by Clair Van Bloem were industrial = 3.5% 50% of industrial land was actively used, building, and/or paved; that 66% of the commercial land was commercial = 26.5%, residential = 38.5%, vacant = 29%, roads/rail/water = 2%. It was assumed that land fell into this category.

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# Table 22 Expsosure Benchmark Values (EBV) for Selected Terrestrialand Aquatic Species (LC50 or LD50)

Species	2,3,7,8-TCDD (mg/kg-day)	Reference
Deer Mouse <sup>b</sup> American Kestrel Earthworm Reed Canary grass Himilayan blackberry	2.850E-02 1.63E-03 NA NA NA	EPA 1993 Hoffman et al 1991








Automotion formation formation formation formation and a second to a second to a second to a second to a second



FIGURE 7 – Local Population Boundary for the American Kestrel



#### APPENDIX A

#### Species and Habitat Survey

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#### Inventory and Habitat Assessment for Rare Plant Species for the J. H. Baxter Site and Reference Site Eugene, Oregon

#### Introduction

Salix Associates was requested by Keystone Environmental to inventory plant and animal species of the J. H. Baxter site, and assess habitats for possible occurrence of rare species. A visit to a nearby reference site also was requested. The site visits occurred on September 16 and 18, 1998.

#### Site Descriptions

The Baxter site is located on the south side of Roosevelt Street in west Eugene, about half way between Seneca and Bertelsen streets (Attachment A). The site is mostly developed with heavy industrial operations, primarily related to treating wood with preservatives. The subject portion of the site (hereinafter called "Site 1") is in the southwest corner, and is approximately 3.5 acres in size. It measures about 300' in a north-south direction, and 500' in an east-west direction. The land form is generally flat, however an L shaped berm several feet in height is present along the south and west sides of the site. (The exact height of the berm is difficult to determine because of thick cover of Himalaya blackberry.) The source of material in the berm is not known to the site manager, but likely originated from the excavated retention pond directly to the east. Along the north edge and in the northeast corner is a low-lying area that ponds water through the winter and into the spring. The area was dry at the time of field surveys. A small ditch, also dry at the time of the surveys, is located along the south property line. A chain link fence is located along the west and south property lines bordering the subject site. A railroad line runs east-west immediately south of the site, and industrial uses border on the west, north and east sides.

The reference site ("Site 2") begins approximately 1/4 mile to the west, on the southwest corner of Bertelsen and Roosevelt streets, and runs westward another 1/4 mile. It is about 16.5 acres in size, and also is flat except for a berm and two drainage channels. The large berm runs towards the west from the old home site, which is just southwest of the northeast corner. Drainageways several feet wide and of unknown depth run along the west and south boundaries of the site. Both drainageways contained water at the time of the field surveys.

#### Habitat Assessment

Site 1 is dominated by scrub-shrub and emergent wetland habitats, and upland herb-dominated habitats. Scrub-shrub dominates the berms along the west and south portions. The dominant species on the berms is Himalaya blackberry (Rubus discolor), an introduced pest. (Species seen at the site are listed in Attachment B.)

The low emergent wetland in the north/northeast portion of Site 1 is dominated by creeping foxtail (Alopecurus geniculatus) and nodding beggars-tick (Bidens cernua). These plants are of low stature, averaging one to two decimeters in height. Reed canarygrass (Phalaris arundinacea) is encroaching around the perimeter, and a few small patches are scattered in the wetland. According to the site manager, the area has standing water through late spring and into early summer. A few Piper's willows (Salix hookeriana) are along the northern edge of the emergent wetland.

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Between the berms along the south and west sides, and this wetland, is an uneven area dominated by reed canarygrass (averaging approximately a meter in height or more), occasional patches of Himalaya blackberry, and other weeds. This area comprises the majority of the site. It is likely that the area is transitional between wetlands and uplands.

Overall, the site has been disturbed extensively, and now is overwhelmingly dominated by non-native weeds. The habitats have some value for insects, small mammals and songbirds. Occasionally, other medium sized mammals (in addition to nutria), and raptors (such as American kestrels or sharp-shinned hawks) and shorebirds (such as killdeer and common snipe) could be expected to use the area.

Site 2 is much larger and more complex than Site 1. Large scale disturbance also has occurred on this site, as evidenced by the presence of straight drainage channels, a large berm, large fields of planted pasture grasses, and a former home site. Species seen at the site are listed in Attachment B.

The large, mostly open areas to the south and east of the former home site contain a variety of emergent and scrub-shrub wetland patches. Overall, the area is dominated by tall fescue (Festuca arundinacea), a weedy pasture grass. Some patches of emergent wetlands have significant populations of native species such as tufted hairgrass (Deschampsia cespitosa, particularly toward the southeast corner of the site). Patches of scrub-shrub wetlands near the central and south central portion of the site are dominated by Piper's willow (Salix hookeriana). Channelized drainages on the west and south borders probably are perennial waterways, or nearly so. Both drainages contained water at the time of the field surveys after a long summer drought period, and both have banks dominated by non-native grasses. The only forested habitat on the site is a grove of ornamental trees surrounding the former home site. Significant use by transient campers was noted in this area, primarily by the presence of garbage, and evidence of parked vehicles and trampling.

#### **Rare Plant and Animal Species**

Rare federally or state listed species which occur in the west Eugene area usually are found in undisturbed or slightly disturbed habitats. Most rare plant species in the area are associated with native wet prairies. Potentially occurring rare species are listed in the table in Attachment C, as are other rare species tracked by the Oregon Natural Heritage Program. Occurrence of any of these species on the Baxter site is highly unlikely because of the high level of disturbance of the habitat, dominance of the site by aggressive nonnative species (Himalaya blackberry and reed canarygrass), and the small size and isolated character. Occurrence of several of these species is possible on the reference site because of overall higher quality of habitat (in the open field areas), larger size, and greater connectivity to other west Eugene open spaces.

Salix Assoc	ates 2525 Potter, E	Eugene, OR 97405 • 541.343.2364	• fax 541.341.1752
FROM :	PHONE NO.	: 6404945 <sup>- 3</sup>	P04
OCT-29-1998 15:24	JALIX ASSOCIATES	503 341 17	752 P.04

#### ATTACHMENTS

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- A Location maps
- B Species lists Baxter Site Reference Site
- C Potentially Occurring Rare Species



P.06 503 341 1752

FROM

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ATTACHMENT B

# Salix Associates

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Preliminary Plant and Animal Species List for the JH Baxter Site, Eugene, OR September 16, 1998 (9:45-11 a.m.; warm, sunny; preceded by record long drought period) Dick Brainerd and Bruce Newhouse

D = dominant

#### PLANTS

General site list: NW, W, S, SE portions Agrostis capillaris Aira caryophyllea Anaphalis margaritacea Arrhenatherum elatius Avena sativa Bromus diandrus Bromus sterilis Centaureum erythraea Cirslum arvense Crataegus douglasii Crepis setosa Cytisus scoparius Daucus carota Equisetum arvense Fraxinus latifolia Holcus lanatus Hypochaeris radicata Juncus effusus Lactuca saligna Leontodon taraxacoides ssp. taraxacoides Leucanthemum vulgare Lupinus rivularis Madia elegans Madia sativa Melllotus albus Parentucellia viscosa Phalaris arundinacea -D Poa sp. Rubus discolor - D Rumex crispus Senecio jacobaea Trifolium pratense Vulpia myuros

### Small wet ditch along S property line

Anaphalis margaritacea Bidens cernua Cirsium arvense

Cirsium vulgare Crataegus douglasii Echinochloa crus galli - D Epilobium cilianum ssp. watsonii Festuca arundinacea Holcus lanatus Lolium perenne Salix hookeriana Toxicodendron diversiloba

Large vernal pool in NE corner, including periphery

Agrostis exarata Alopecurus geniculatus - D Bidens cernua - D Carex densa Carex stipata Cirsium vulgare Dipsacus fullonum Gnaphalium sp. Lotus corniculatus Lythrum portula Mentha pulegium Panicum capillaris Phalaris arundinacea Polygonum hydropiperoldes Populus balsamifera ssp. trichocarpa Salix hookeriana Salix lucida ssp. lasiandra

#### ANIMALS

Birds American kestrel barn swallow black-capped chickadee common snipe (overhead) European starling (adjacent to S) greater yellowlegs (retention pond to E) killdeer (overhead) sparrow, unidentified (brief look; in RUBDIS thicket; probably song sparrow; possibly golden-

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crowned or Lincoln's spotted towhee (adjacent to S)

#### Mammals

FROM :

nutria (scat/trail) house mouse (trapped by G. Orth) deer mouse (trapped by G. Orth)

#### Reptiles northwestern garter snake

Insects

blue (butterfly: unidentified species; female) Carolina locust juba skipper moth (unidentified, buff colored, ~3cm wingspan) sulphur (butterfly; probably orange or common) ост-29-1990 15:26

PØ8

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J. H. Baxter Reference Site: SW corner of Bertelsen and Roosevelt, Eugene, OR September 16 and 18, 1998: D. Brainerd, B. Newhouse, J. Tilson

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D = dominant

East and west fields, and large berms Asclepias speciosa Aira caryophyllea Allium sp. Alopecurus pratensis - D Arum italicum Aster hallii Beckmannia syzigachne Bidens frondosa Briza minor Bromus of, secalinus Bromus sitchensis Carex obnupla Carex unilateralis Centaureum erythraea Convolvulus arvensis Crataegus douglasii Crataegus monogyna Crepis selosa Cynosurus echinatus Cytisus scoparius Dactylis glomerata Daucus carola - D Deschampsia cespitosa - D Deschampsia danthonioldes Dianthus armeria Dipsacus fullonum Epilobium brachycarpum Epiloblum ciliatum ssp. watsonli Festuca arundinacea - D Fraxinus latifolia Gnaphalium sp. Holcus lanatus - D Hordeum brachyantherum Hypericum perforatum Hypochaeris radicata Juncus patens Kickxia elatine Lactuca serriola Lathyrus latifolms Leontodon taraxacoides ssp. taraxacoldes Lotus denticulatus Lotus purshianus Madia elegans Madia glomerata Madia sativa Mentha pulegium Parentucellia viscosa - D Phalaris arundinacea - D Phleum pratense Poa pratensis Populus balsamifera ssp. trichocarpa Rosa multiflora Rubus discolor - D Rubus laciniatus Rumex crispus Rumex salicifolius Salix hookeriana - D Senecio jacobea Tragopogon sp. Trifolium pratense Vicia cf. hirsuta Vicia cracca Ditch along west boundary Bromus sitchensis Cirsium arvense Crataegus douglasii Dipsacus fullonum Echinochloa crus-galli -D Epilobium ciliatum ssp. watsonii Festuca arundinacea - D

Hordeum brachyantherum

Hypericum perforatum

Hypochaeris radicata

Phalaris arundinacea-D

Juncus effusus

Lemna minor

Phleum pratense

Poa pratensis

Plantago lanceolata

Rubus discolor -D

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OCT-29-1998

503 341 1752 P.09 P09

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15:26

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#### Rumer sp. Salix hookeriana

#### Ditch along south boundary

All spp. except Lemna rooted on bank. Agrostis capillaris Carex stipata Cirsium arvense Holcus lanatus Juncus effusus Lemna minor - D Melissa officinalis Poa sp. Polystichum munitum Torilis arvensis

Homesite area: NE corner

Asclepias speciosa Agrostis capillaris - D Cirsium arvense Cirsium vulgare Corylus cf. avellana Crataegus douglas! Deschampsia cespitosa Festuca arundinacea - D Juglans nigra Lactuca serriola Malus sylvestris Phleum pratense Prunus cerasiformis Pyrus communis Rubus discolor - D Salix babylonica

Birds

American kestrel barn swallow black-capped chickadee common snipe (o-head) European starling greater yellowlegs house finch killdeer (o-h) mailard savannah sparrow spotted towhee (to S)

song sparrow

Mammals nutria (scat, trail, grazed vegetation) raccoon scat

Reptiles (Northwestern?) Garter snake

Insects Carolina locust juba skipper sulphur butterfly tan (female?) "blue" unid., sm., white moth (-3cm dia)

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# Occurrence of Rare Vascular Plants, Amphibians, Birds, Mammals and Reptiles at the J. H. Baxter and Reference Sites, Lane County, Willamette Valley Province

known to occur in Lane County in the Willamette Valley Province are included on the list. Habitat and survey information from Salix Associates and: This list was excerpted from "Rare, Threatened and Endangered Plants and Animals of Oregon" (March 1998, Oregon Natural Heritage Program). Species listed below do not include invertebrates, non-vascular plants and fungi, for which very little information is known in most cases. Species Corkran and Thoms 1996, Cauli et al. 1997, Leonard et al. 1993, Marshall 1996, Storm and Leonard eds. 1995, Verts and Carraway 1998.

Key to "At JHB/R Site?": 1 = Baxter site, 2 = Reference site; C = confirmed on site; P = possible on site, but not confirmed; U = unlikely on site.

Scentific	Fed. State Shine Shite	State	ONUT	Best Survey Time		THB/R	
VASCTITAR PLANTS						11 20	
	SoC	LT	-1	8/1-9/15	Native upland prairie, usually just above jurisdictional weitance.	11. 21	
Asid cuius Ador vialis	SoC	LT	-	7/1-8/30	Upland forest edges or operanges in woods, occasionally in part	2, 'n r	
						1U, 2P	
singly and successful singly and successful successful successful successful successful successful successful s	1	1	2	5/15-6/15	Vernally moist areas, usually with inche compound ves	116 213	
Cimicifuga elata	SoC	0	1	51/6-1/9	North slopes with PSEMEN/ACEMAC (rarely PSEMEN/ALNRUB); moist site herbs (DICFOR, PROHOO,	1 c + 2	AT.
				-	HYDTEN, ADIALE)		[A(
Erizeron decumbens var. decumbens	Prop.	LE		6/15-7/30	Native to disturbed, wetland to upland, prairies, pastures, tree farms, roadsides.	10, 11	.HM
	ш   					1U, 2P	EN
Hurkelja congesta ssp. congesta	SoC	<u>ں</u>	-	6/15-7/30	Native prairie, moist to uplane		r C
							-

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Lathyrus holochlorus			4	16/12-2//31	Hedgerows, edges of woods, usually upland, occasionally moist.	11, 20
Lomatium bradshawii	TE	LE	÷	4/1-5/15	Usually native, wet prairie. Occasionally in pastures.	1U, 2P
Lupinus sulphureus ssp. kincaidii	PT	LT		51/6-1/9	Wetland to upland, rative to disturbed, prairie to hedgerow, occ. under RUBDIS.	IU, 2P
Moatta howellii	SoC	U	-	11/15- 4/31	Survey time variable depending on rainfall. Prefers flat or gently sloping areas with shallow standing water (e.g., vernal pools) or flow; often in gravel areas (parking lots, old roads, gravel piles), or occasionally native wet prairie. Often with POAANN.	JU, 2P
Sidalcea cannestris	i	U	4	<i>S/</i> 15-7/15	Hedgerows, fences, edges of woods Upland to moist	1U, 2P
Sidalcea cusickii	1	1	*	5/15-7/15	Hedgerows, fearces, edges of woods. Moist to upland.	1U, 2P.
AMPHIBIANS						
clouded salamander	-	su	м	warm, wct	Edges of Valley, foothills. In decomposing logs and stumps. Occasionally in rock crevices.	10, 20
unthen red-leved frog	SC SC	SV	m	3	Streams, ponds, swamps and adjacent forests.	10, 20
Oreson spotted froe	U	sc	*	3	Marshes, wet meadows, ponds, slow streams.	10, 20
Cascade seep salamander		sc	ŧĒ	÷.	West edge of Valley, foothills. Cold streams and adjacent forests.	10,2V
BIRDS						
grasshopper sparrow	 	SP	ŧ	4/15-6/30	Undisturbed grassfaulds.	10, 20
w. Oregon little willow flycatcher	SoC	λS		2	Thickets of willows or other shurb species, usually near water.	1U, 2P
streaked horned kark	1	8	<b></b>	3	Large, open fields.	1U, 2U
vellow-breasted chat	1	Š	4	5/15-6/30	Tree and shrub thickets, usually near water.	1U, 2P

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		1	3	4/15-6/30	Associated with oak forests.	n, 1
arcom weodpocket		5	*C		Grasslands, fields, often with scattered shubs.	10, 2P
Oregon vesper sparrow		ىد SV	4	3	ats. Occasional on	(U, 2P
western mesidowlark	1	sc	4	3	Open fields and grasslands, usually with scattered shrubs or small	iU, 2P
					rices.	
MAMMALS					financial and a financial ones fields. Day roosts in	1U, 2U
palind bat	1	SV	<u>م</u>	summer	Brushy, rocky areas, cuess or would be crevices in structures, caves, lrees, etc.	
	Soc	SU		sprng- fail	Associated with streams in deciduous forest, and conifer forest.	11, 20
while-tootcd voic Davids western bie-carted bat	SoC	sc	2*	Summer	Associated with conferous forests. Day roosts in shallow roof	10, 20
	5	(IS	4	2	Associated with conferous forests. Day roosts in buildings,	1U, 2U
long-cared bat					carces, hollowy trees, under ortiges, the outs, in your and	10.20
frinted hat	SoC	SV	3*	÷	th forests.	111.22
Viena hat	SoC		4	;	Associated with water. Day roosts in buildings, caves, used	111 20
western gray squirrel	1	su	m	any	Usually associated with Oregon white oak lorests and savauses.	
REPTILES						IU, 2P
northwestern poord turtle	SC	sc	5#	4/1-9/30	Ponds, axoows, mow moved a second forest: acc. in grasslands.	10, 21
sharptail snake	1	S	4	3/1-11/15	Most areas in connervue or upper and a fill of the	10, 20
	1	SV	4	5/1-9/30	Usually associated with warm, rocky musicos.	

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#### APPENDIX B

**Endpoint Species Description** 



#### Appendix B Endpoint Species Description

**Deer Mouse** (Perymyscus maniculatus) (Taken from Distribution, Habitat and Natural History Atlas of Oregon Wildlife (Causti, 1997))

The deer mouse is found from coast to coast from central Canada to southern Mexico. It is absent only from southeastern United States and Coastal Mexico. About the only habitat that deer mice do not use is open water. They are found through Oregon in every habitat type. Deer mice are capable of breeding throughout the year. However, populations along the Oregon coast may have fewer litters from October to February. Deer mice are opportunistic Omnivores. The deer mouse weighs about 0.021 kilograms. They eat seeds, green vegetation, insects, berries and fungi. For the purposes of this risk assessment the were assumed to eat 72.2% vegetation and seeds, 25.8% insects and soil invertebrates, and 2% soil (EPA 1993). They consume approximately 0.00418 kilograms/day of food. Their home range is about 0.09 hectares. Deer mice are a stable prey for about everything carnivorous.

American Kestrel (Taken from Distribution, Habitat and Natural History Atlas of Oregon Wildlife (Causti, 1997))

The American kestrel is widely distributed in the New World, except for dense tropical moist forest. It breeds from Alaska south through Canada, the United States, and Mexico, into Central America and farther south into South America. The American kestrel uses a wide variety of open and semi-open habitats, including grassland, desert, juniper woodlands, meadows and clearcuts in forests, marshes, agricultural fields, and even urban areas. Their preferred nests are a woodpecker hole or a natural cavity in a tree, but an American kestrel will make do with covered rock ledges, or nest boxes. The breeding

season begins in April and the young are fledged by August. A clutch of 4 or 5 are incubated 29 - 30 days by the female. Young are tended by both parents and are independent in 4 or 5 weeks. The American kestrel will feed insects when available but when insects are seasonally low they will feed on small mammals and sometimes birds. Studies on feeding by the American kestrel indicate that it eats about 55% wildlife and 45% insects. For the purposes of the risk assessment it was assumed that they also incidentally consume 2% of their diet by weight of soil. The American kestrel density varies with food supply. Two studies found territory size of 109 and 130 hectares. For the purposes of this risk assessment it was assumed that their territory size (home range) is 120 hectares. The American kestrel weighs about 0.12 kilograms and eats about .0348 kilograms of food each day (EPA 1993).

Himalayan Blackberry (Rubus discolor) (Taken from Plants of Coastal British Columbia including Washington, Oregon, and Alaska. (Pojar and Mackinnon 1994))

The Himalayan blackberry is an Asian species of blackberry introduced from India via England and widely naturalized, in disturbed sites and streamside areas. It is the most common introduced blackberry in this area and a favourite of berry pickers. They are erect to sprawling; stout stems erect, then arching, then trailing along the ground and rooting at the ends. They have stout reformed prickles and often form dense impenetrable thickets. The leaves are more or less evergreen, trifoliate to 5-foliate, smooth above and covered with white hairs below. They produce edible blackberries.

Reed Canary Grass (Taken from Plants of Coastal British Columbia including Washington, Oregon, and Alaska. (Pojar and Mackinnon 1994))

Reed canary grass is a robust perennial, 0.7 - 2 metres tall, with long, scaly, pinkish rhizomes and hollow stems. The leaves are roughened; sheaths open, margins

overlapping; ligules 4-10 millimetres long, usually tattered and turned backwards. They have glumes about 4-5 centimetres long. They prefer wet places in disturbed sites including clearings, along ditches, marshy spots and depressions, stream-banks and along the edges of wetlands, they are scattered but often locally abundant.

Earthworms (Taken from Toxicological Benchmarks for Contaminants of Potential Concern for Effects on Soil and Litter Invertebrates and Heterotrophic Processes: 1977 Revision (Efroymson et al 1977)

Earthworms are probably the most important soil invertebrates in promoting soil fertility. Their feeding and borrowing activities break down organic matter and release nutrients and improve aeration, drainage, and aggregation of soil. Earthworms are also am important component of the diet of many higher animals. Earthworms are known to take up many organic and inorganic contaminants.

One of the most common species of earthworm is Eisenia fetida, a non-borrowing organism found in organic rich environments. Another species Lumbricus rubellus is a shallow-burrowing lumbricid active is surface and litter horizons of pastures and grasslands. It may forage for food, such as dead roots in the subsurface horizon, and dig deep borrows in which to rest during periods of environmental stress. A third species Octalasium cyaneum is a burrowing lumbricid species that lives in the soil and feeds on dead roots. It is common in pastureland where it creates deep horizontal burrows.

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