

California Environmental Protection Agency
AIR RESOURCES BOARD

**CALIFORNIA NON-METHANE ORGANIC GAS
TEST PROCEDURES**

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Monitoring and Laboratory Division, Southern Laboratory Branch
Mobile Source Division
9528 Telstar Avenue
El Monte, California 91731

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Part A

GENERAL APPLICABILITY AND REQUIREMENTS

1. These test procedures shall apply to all 1993 and subsequent model-year transitional low-emission vehicles (TLEV), low-emission vehicles (LEV), ultra-low-emission vehicles (ULEV), and super-ultra-low-emission vehicles (SULEV) certifying to non-methane organic gas (NMOG) emission standards.
2. This document sets forth the analysis and calculation procedures that shall be performed to determine NMOG mass emissions. The document consists of the following parts:
 - A. General Applicability and Requirements
 - B. Determination of Non-Methane Hydrocarbon Mass Emissions by Flame Ionization Detection
 - C. Determination of Alcohols in Automotive Source Samples by Gas Chromatography (Method No. 1001)
 - D. Determination of C₂ to C₅ Hydrocarbons in Automotive Source Samples by Gas Chromatography (Method No. 1002)
 - E. Determination of C₆ to C₁₂ Hydrocarbons in Automotive Source Samples by Gas Chromatography (Method No. 1003)
 - F. Determination of Aldehyde and Ketone Compounds in Automotive Source Samples by High Performance Liquid Chromatography (Method No. 1004).
 - G. Determination of NMOG Mass Emissions

Appendix 1 List of Light-End and Mid-Range Hydrocarbons

Appendix 2 Definitions and Commonly Used Abbreviations

Appendix 3 References

Alternative procedures may be used if shown to yield equivalent results and if approved in advance by the Executive Officer of the Air Resources Board.

3. The analyses specified in the table below shall be performed to determine mass emission rates of NMOG in grams per mile (g/mi) or milligrams per mile (mg/mi) for vehicles operated on the listed fuel:

| Fuel | NMHC by FID | NMHC by GC | Alcohols | Carbonyls |
|----------|-------------|------------|----------|-----------|
| Alcohol | X | | X | X |
| CNG | | X | | X |
| Diesel | X | | | X |
| Gasoline | X | | | X |
| LPG | X | | | X |

The specified analyses shall be performed in accordance with the following parts of this document:

| | | |
|----------------------|---------|--|
| NMHC by FID-- | Part B. | Determination of Non-Methane Hydrocarbon Mass Emissions by Flame Ionization Detection |
| NMHC by GC-- | Part D. | Determination of C ₂ to C ₅ Hydrocarbons in Automotive Source Samples by Gas Chromatography (Method No. 1002); and |
| | Part E. | Determination of C ₆ to C ₁₂ Hydrocarbons in Automotive Source Samples by Gas Chromatography (Method No. 1003) |
| CARBONYLS-- | Part F. | Determination of Aldehyde and Ketone Compounds in Automotive Source Samples by High Performance Liquid Chromatography (Method No. 1004) |
| ALCOHOLS -- | Part C. | Determination of Alcohols in Automotive Source Samples by Gas Chromatography (Method No. 1001) |

4. For those manufacturers that choose to develop reactivity adjustment factors unique to a specific engine family, exhaust NMOG emissions shall be fully speciated. NMHC emissions shall be analyzed in accordance with parts D and E (Method Nos. 1002 and 1003). In addition, aldehydes and ketones, alcohols, and ethers shall be analyzed according to parts F, C, and E (Method Nos. 1004, 1001, and 1003). Analysis for alcohols shall be required only for vehicles that are operated on fuels containing alcohols.
5. For natural gas-fueled vehicles, the methane concentration in the exhaust sample shall be measured with a methane analyzer. A GC combined with a FID is used for direct measurement of methane concentrations. SAE Recommended Practice J1151 is a reference on generally accepted GC principles and analytical techniques for this application. A density of 18.89 g/ft³ shall be used to determine the methane mass emissions. The methane mass emissions shall be multiplied by the appropriate methane reactivity adjustment factor and then added to the reactivity-adjusted NMOG emissions as specified in the “California Exhaust Emission Standards and Test Procedures for 1988-2000 Model Passenger Cars, Light-Duty Trucks, and Medium-Duty Vehicles” and in the “California Exhaust Emission Standards and Test Procedures for 2001 and Subsequent Model Passenger Cars, Light-Duty Trucks, and Medium-Duty Vehicles.”
6. The mass of NMOG emissions shall be calculated in accordance with part G, “Determination of NMOG Mass Emissions”. The mass of NMOG emissions in g/mile or mg/mile shall be calculated by summing the mass of NMHC determined by the FID, the mass of aldehydes and ketones, and the mass of alcohols.

PART B

DETERMINATION OF NON-METHANE HYDROCARBON MASS EMISSIONS BY FLAME IONIZATION DETECTION

1. INTRODUCTION

- 1.1 This procedure describes a method for determining NMHC exhaust mass emissions from motor vehicles. Other applicable forms of instrumentation and analytical techniques which prove to yield equivalent results to those specified in this procedure may be used subject to the approval of the Executive Officer of the Air Resources Board.
- 1.2 All definitions and abbreviations are contained in Appendix 2 of these test procedures.

2. TOTAL HYDROCARBON MEASUREMENT

- 2.1 A FID is used to measure total hydrocarbon concentration in vehicle exhaust in accordance with the Code of Federal Regulations [Ref.1]. SAE Recommended Practices J254 [Ref. 2] and J1094a [Ref. 3] are references on generally accepted gas analysis and constant volume sampling techniques. For Beckman 400 FIDs only, implementation of the recommendations outlined in SAE paper 770141[Ref. 4] shall be required. Other FID analyzer models shall be checked and adjusted, if necessary, to minimize any non-uniformity of relative response to different hydrocarbons.

3. METHANE MEASUREMENT

- 3.1 A GC combined with a FID constitute a methane analyzer and shall be used for direct measurement of methane concentrations. The SAE Recommended Practice J1151[Ref. 5] is a reference on generally accepted GC principles and analytical techniques for this specific application.

4. TOTAL HC FID RESPONSE TO METHANE

- 4.1 The FID is calibrated to propane and therefore tends to over respond to the methane portion of the vehicle exhaust sample during hydrocarbon analysis. In order to calculate the NMHC concentration, a methane response factor must be applied to the methane concentration (as measured by the methane analyzer) before it can be deducted from the total hydrocarbon concentration. To determine the total hydrocarbon FID response to methane, known methane in air concentrations traceable to NIST shall be analyzed by the FID. Several methane concentrations shall be analyzed by the FID in the range of the exhaust sample concentration. The total hydrocarbon FID response to methane is calculated as follows:

$$r_{\text{CH}_4} = \text{FID}_{\text{ppm}} / \text{SAM}_{\text{ppm}}$$

where:

- r_{CH_4} = FID methane response factor.
- FID_{ppm} = FID reading in ppmC.
- SAM_{ppm} = the known methane concentration in ppmC.

The FID response to methane shall be checked at each calibration interval.

5. NMHC MASS EMISSION PER TEST PHASE

5.1 The following calculations shall be used to determine the NMHC mass emissions for each phase of the Federal Test Procedure [Ref. 1].

5.2 Non-Alcohol Fueled Vehicles

5.2.1 $NMHC_e = FID\ THC_e - (r_{CH_4} * CH_{4e})$

NOTE: If $NMHC_e$ is calculated to be less than zero, then $NMHC_e = 0$.

5.2.2 $NMHC_d = FID\ THC_d - (r_{CH_4} * CH_{4d})$

NOTE: If $NMHC_d$ is calculated to be less than zero, then $NMHC_d = 0$.

5.2.3 $CO_e = (1 - (0.01 + 0.005 * HCR) * CO_{2e} - 0.000323 * R_a) * CO_{em}$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e .

a) For gasoline, $CH_{1.85}$, where $HCR = 1.85$:
 $CO_e = (1 - 0.01925 * CO_{2e} - 0.000323 * R_a) * CO_{em}$

b) For Phase 2 gasoline, $CH_{1.94}$, where $HCR = 1.94$:
 $CO_e = (1 - 0.01970 * CO_{2e} - 0.000323 * R_a) * CO_{em}$

c) For LPG, $CH_{2.64}$, where $HCR = 2.64$:
 $CO_e = (1 - 0.02320 * CO_{2e} - 0.000323 * R_a) * CO_{em}$

d) For CNG, $CH_{3.78}$, where $HCR = 3.78$:
 $CO_e = (1 - 0.02890 * CO_{2e} - 0.000323 * R_a) * CO_{em}$

$$5.2.4 \quad DF = \frac{100 * \left(\frac{x}{x + y/2 + 3.76 * (x + y/4 - z/2)} \right)}{CO_{2e} + (NMHC_e + CH_{4e} + CO_e) * 10^{-4}}$$

(where fuel composition is $C_xH_yO_z$ as measured for the fuel used.)

- a) For gasoline, $CH_{1.85}$, where $x = 1$, $y = 1.85$, and $z = 0$:
 $DF = 13.47 / [CO_{2e} + (NMHC_e + CH_{4e} + CO_e) * 10^{-4}]$
- b) For Phase 2 gasoline, $CH_{1.94}$, $x = 1$, $y = 1.94$ and $z = 0.017$:
 $DF = 13.29 / [CO_{2e} + (NMHC_e + CH_{4e} + CO_e) * 10^{-4}]$
- c) For LPG, $CH_{2.64}$, where $x = 1$, $y = 2.64$, and $z = 0$:
 $DF = 11.68 / [CO_{2e} + (NMHC_e + CH_{4e} + CO_e) * 10^{-4}]$
- d) For CNG, $CH_{3.78}$, where $x = 1$, $y = 3.78$, and $z = 0.016$:
 $DF = 9.83 / [CO_{2e} + (NMHC_e + CH_{4e} + CO_e) * 10^{-4}]$

5.3 Vehicles Operating on Fuels Containing Methanol

5.3.1 $NMHC_e = FID\ THC_e - (r_{CH_4} * CH_{4e}) - (r_{CH_3OH} * CH_3OH_e)$

NOTE: If $NMHC_e$ is calculated to be less than zero, then $NMHC_e = 0$.

5.3.2 $NMHC_d = FID\ THC_d - (r_{CH_4} * CH_{4d}) - (r_{CH_3OH} * CH_3OH_d)$

NOTE: If $NMHC_d$ is calculated to be less than zero, then $NMHC_d = 0$.

5.3.3 $CO_e = (1 - (0.01 + 0.005 * HCR) * CO_{2e} - 0.000323 * R_a) * CO_{em}$

NOTE: If a CO instrument which meets the criteria specified in CFR 40 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e .

- a) For M100 (100% methanol), CH_3OH , where $HCR = 4$:
 $CO_e = (1 - 0.03000 * CO_{2e} - 0.000323 * R_a) * CO_{em}$

- b) For M85 (85% methanol, 15% indolene), $CH_{3.41}O_{0.72}$, where $HCR = 3.41$:
 $CO_e = (1 - 0.02705 * CO_{2e} - 0.000323 * R_a) * CO_{em}$

$$5.3.4 \quad DF = \frac{100 * \left(\frac{x}{x + y/2 + 3.76 * (x + y/4 - z/2)} \right)}{CO_{2e} + (NMHC_e + CH_{4e} + CO_e + CH_3OH_e + HCHO_e) * 10^{-4}}$$

(where fuel composition is $C_xH_yO_z$ as measured for the fuel used.)

- a) For M100 (100% methanol), CH_3OH , where $x = 1$, $y = 4$, and $z = 1$:
 $DF = 11.57 / [CO_{2e} + (NMHC_e + CH_{4e} + CO_e + CH_3OH_e + HCHO_e) * 10^{-4}]$
- b) For M85 (85% methanol, 15% Indolene), $CH_{3.41}O_{0.72}$, where $x = 1$,
 $y = 3.41$, and $z = 0.72$:
 $DF = 12.02 / [CO_{2e} + (NMHC_e + CH_{4e} + CO_e + CH_3OH_e + HCHO_e) * 10^{-4}]$

5.4 Vehicles Operating on Fuels Containing Ethanol

$$5.4.1 \quad NMHC_e = FID \text{ THC}_e - (r_{CH_4} * CH_{4e}) - (r_{C_2H_5OH} * C_2H_5OH_e)$$

NOTE: If $NMHC_e$ is calculated to be less than zero, then $NMHC_e = 0$

$$5.4.2 \quad NMHC_d = FID \text{ THC}_d - (r_{CH_4} * CH_{4d}) - (r_{C_2H_5OH} * C_2H_5OH_d)$$

NOTE: If $NMHC_d$ is calculated to be less than zero, then $NMHC_d = 0$

$$5.4.3 \quad CO_e = (1 - (0.01 + 0.005 * HCR) * CO_{2e} - 0.000323 * R_a) * CO_{em}$$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e .

- a) For E100 (100% ethanol), C_2H_5OH , where $HCR = 3$:
 $CO_e = (1 - 0.02500 * CO_{2e} - 0.000323 * R_a) * CO_{em}$

$$5.4.4 \quad DF = \frac{100 * \left(\frac{x}{x + y/2 + 3.76 * (x + y/4 - z/2)} \right)}{CO_{2e} + (NMHC_e + CH_{4e} + CO_e + C_2H_5OH_e + HCHO_e) * 10^{-4}}$$

(where fuel composition is $C_xH_yO_z$ as measured for the fuel used.)

- a) For E100 (100% ethanol), C_2H_5OH , where $x = 1$, $y = 3$, and $z = 0.5$:
 $DF = 12.29 / [CO_{2e} + (NMHC_e + CH_{4e} + CO_e + C_2H_5OH_e + HCHO_e) * 10^{-4}]$

5.5 All Vehicles

$$5.5.1 \quad NMHC_{conc} = NMHC_e - NMHC_d * [1 - (1 / DF)]$$

NOTE: If $NMHC_{conc}$ is calculated to be less than zero, then $NMHC_{conc} = 0$

$$5.5.2 \quad NMHC_{mass_n} = NMHC_{conc} * NMHC_{dens} * VMIX * 10^{-6}$$

6. TOTAL WEIGHTED NMHC MASS EMISSIONS

6.1 All Vehicles

$$6.1.1 \quad \text{NMHC}_{\text{wm}} = 0.43 * \left(\frac{\text{NMHC}_{\text{mass1}} + \text{NMHC}_{\text{mass2}}}{D_{\text{phase1}} + D_{\text{phase2}}} \right) + 0.57 * \left(\frac{\text{NMHC}_{\text{mass3}} + \text{NMHC}_{\text{mass2}}}{D_{\text{phase3}} + D_{\text{phase2}}} \right)$$

7. SAMPLE CALCULATIONS

7.1 Given the following data for a gasoline vehicle, calculate the weighted NMHC mass emission.

| Test Phase | FID THC _e (ppmC) | FID THC _d (ppmC) | CH _{4e} (ppmC) | CH _{4d} (ppmC) | CO _{em} (ppm) | CO _{2e} (%) | VMIX (ft ³) | D _{phase n} (mile) | R _a (%) |
|------------|-----------------------------|-----------------------------|-------------------------|-------------------------|------------------------|----------------------|-------------------------|-----------------------------|--------------------|
| 1 | 41.8 | 8.6 | 7.53 | 5.27 | 147.2 | 1.19 | 2846 | 3.583 | 38 |
| 2 | 13.0 | 8.4 | 5.68 | 5.10 | 20.8 | 0.80 | 4856 | 3.848 | 38 |
| 3 | 15.4 | 8.9 | 6.16 | 5.20 | 36.7 | 1.04 | 2839 | 3.586 | 38 |

For Phase 1:

$$\begin{aligned} \text{NMHC}_e &= \text{FID THC}_e - (r_{\text{CH}_4} * \text{CH}_{4e}) \\ &= 41.8 \text{ ppmC} - (1.04 * 7.53 \text{ ppmC}) \\ &= 33.97 \text{ ppmC} \end{aligned}$$

$$\begin{aligned} \text{NMHC}_d &= \text{FID THC}_d - (r_{\text{CH}_4} * \text{CH}_{4d}) \\ &= 8.6 \text{ ppmC} - (1.04 * 5.27 \text{ ppmC}) \\ &= 3.12 \text{ ppmC} \end{aligned}$$

$$\text{CO}_e = (1 - 0.01925 * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e.

$$\begin{aligned} &= (1 - 0.01925 * 1.19\% - 0.000323 * 38\%) * 147.18 \text{ ppm} \\ &= 142.0 \text{ ppm} \end{aligned}$$

$$\text{DF} = 13.47 \div [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e) * 10^{-4}]$$

$$\begin{aligned} \text{DF} &= \frac{13.47}{1.19\% + (33.97 \text{ ppmC} + 7.53 \text{ ppmC} + 142.0 \text{ ppmC}) * 10^{-4}} \\ &= 11.15 \end{aligned}$$

$$\text{NMHC}_{\text{conc}} = \text{NMHC}_e - \text{NMHC}_d * [1 - (1 \div \text{DF})]$$

$$= 33.97 \text{ ppmC} - 3.12 \text{ ppmC} * [1 - (1 \div 11.15)]$$

$$= 31.13 \text{ ppmC}$$

$$\text{NMHC}_{\text{mass } n} = \text{NMHC}_{\text{conc}} * \text{NMHC}_{\text{dens}} * \text{VMIX} * 10^{-6}$$

$$= 31.13 \text{ ppmC} * 16.33 \text{ g/ft}^3 * 2846 \text{ ft}^3 * 10^{-6}$$

$$\text{NMHC}_{\text{mass } 1} = 1.45 \text{ g}$$

Similarly, for Phase 2: $\text{NMHC}_{\text{mass } 2} = 0.33 \text{ g}$
 and for Phase 3: $\text{NMHC}_{\text{mass } 3} = 0.27 \text{ g}$

Therefore,

$$\text{NMHC}_{\text{wm}} = 0.43 * \left(\frac{\text{NMHC}_{\text{mass } 1} + \text{NMHC}_{\text{mass } 2}}{D_{\text{phase } 1} + D_{\text{phase } 2}} \right) + 0.57 * \left(\frac{\text{NMHC}_{\text{mass } 3} + \text{NMHC}_{\text{mass } 2}}{D_{\text{phase } 3} + D_{\text{phase } 2}} \right)$$

$$\text{NMHC}_{\text{wm}} = 0.43 * \left(\frac{1.45 \text{ g} + 0.33 \text{ g}}{3.583 \text{ miles} + 3.848 \text{ miles}} \right) + 0.57 * \left(\frac{0.27 \text{ g} + 0.33 \text{ g}}{3.586 \text{ miles} + 3.848 \text{ miles}} \right)$$

$$\text{NMHC}_{\text{wm}} = 0.15 \text{ g/mile}$$

7.2 Given the following data for a vehicle operating on 85% methanol and 15% gasoline (M85), calculate the weighted NMHC mass emission.

| Test Phase | FID THC _e (ppmC) | FID THC _d (ppmC) | CH _{4e} (ppmC) | CH _{4d} (ppmC) | CH _{3OH_e} (ppm) | CO _{em} (ppm) | CO _{2e} (%) | VMIX (ft ³) | D _{phase n} (mile) | R _a (%) | HCHO _e (ppm) |
|------------|-----------------------------|-----------------------------|-------------------------|-------------------------|-------------------------------------|------------------------|----------------------|-------------------------|-----------------------------|--------------------|-------------------------|
| 1 | 88.5 | 5.5 | 17.76 | 2.82 | 72.9 | 303.2 | 1.28 | 2832 | 3.570 | 32 | 0.96 |
| 2 | 14.5 | 7.0 | 8.01 | 2.82 | 5.1 | 9.7 | 0.83 | 4827 | 3.850 | 32 | 0.10 |
| 3 | 21.8 | 7.7 | 10.13 | 2.93 | 7.4 | 18.2 | 1.13 | 2825 | 3.586 | 32 | 0.12 |

[For this example, CH_{3OH_d} was assumed to be 0.0 ppmC for all three background bag samples.]

For Phase 1:

$$\text{NMHC}_e = \text{FID THC}_e - (r_{\text{CH}_4} * \text{CH}_{4e}) - (r_{\text{CH}_3\text{OH}} * \text{CH}_3\text{OH}_e)$$

$$= 88.5 \text{ ppmC} - (1.04 * 17.76 \text{ ppmC}) - (0.66 * 72.9 \text{ ppmC})$$

$$= 21.92 \text{ ppmC}$$

$$\begin{aligned}
\text{NMHC}_d &= \text{FID THC}_d - (r_{\text{CH}_4} * \text{CH}_{4d}) - (r_{\text{CH}_3\text{OH}} * \text{CH}_3\text{OH}_d) \\
&= 5.5 \text{ ppmC} - (1.04 * 2.82 \text{ ppmC}) - (0.66 * 0.0 \text{ ppmC}) \\
&= 2.57 \text{ ppmC}
\end{aligned}$$

$$\text{CO}_e = (1 - 0.02705 * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e.

$$\begin{aligned}
&= (1 - 0.02705 * 1.28\% - 0.000323 * 32\%) * 303.2 \text{ ppm} \\
&= 289.6 \text{ ppm}
\end{aligned}$$

$$\text{DF} = 12.02 \div [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e + \text{CH}_3\text{OH}_e + \text{HCHO}_e) * 10^{-4}]$$

$$= \frac{12.02}{1.28\% + (21.92 \text{ ppmC} + 17.76 \text{ ppmC} + 289.6 \text{ ppmC} + 72.9 \text{ ppmC} + 0.96 \text{ ppmC}) * 10^{-4}}$$

$$= 9.10$$

$$\begin{aligned}
\text{NMHC}_{\text{conc}} &= \text{NMHC}_e - \text{NMHC}_d * [1 - (1 / \text{DF})] \\
&= 21.92 \text{ ppmC} - 2.57 \text{ ppmC} * [1 - (1 / 9.10)] \\
&= 19.63 \text{ ppmC}
\end{aligned}$$

$$\text{NMHC}_{\text{mass n}} = \text{NMHC}_{\text{conc}} * \text{NMHC}_{\text{dens}} * \text{VMIX} * 10^{-6}$$

$$\text{NMHC}_{\text{mass 1}} = 0.91 \text{ g}$$

$$\text{Similarly, Phase 2: } \text{NMHC}_{\text{mass 2}} = 0.0 \text{ g}$$

$$\text{and for Phase 3: } \text{NMHC}_{\text{mass 3}} = 0.10 \text{ g}$$

Therefore,

$$\text{NMHC}_{\text{wm}} = 0.43 * \left(\frac{\text{NMHC}_{\text{mass1}} + \text{NMHC}_{\text{mass2}}}{D_{\text{phase1}} + D_{\text{phase2}}} \right) + 0.57 * \left(\frac{\text{NMHC}_{\text{mass3}} + \text{NMHC}_{\text{mass2}}}{D_{\text{phase3}} + D_{\text{phase2}}} \right)$$

$$\text{NMHC}_{\text{wm}} = 0.43 * \left(\frac{0.91 \text{ g} + 0.00 \text{ g}}{3.570 \text{ miles} + 3.850 \text{ miles}} \right) + 0.57 * \left(\frac{0.10 \text{ g} + 0.00 \text{ g}}{3.586 \text{ miles} + 3.850 \text{ miles}} \right)$$

$$\text{NMHC}_{\text{wm}} = 0.06 \text{ g/mile}$$

Part C

DETERMINATION OF ALCOHOLS IN AUTOMOTIVE SOURCE SAMPLES BY GAS CHROMATOGRAPHY

METHOD NO. 1001

1. INTRODUCTION

- 1.1 This document describes a method of sampling and analyzing automotive exhaust for alcohols in the range of 1 to 1200 µg per 15 mL of solution. The “target” alcohols that shall be analyzed and reported by this method are methanol and ethanol. These alcohols, when measured in concentrations above the LOD, shall be reported.
- 1.2 This procedure is based on a method developed by the U. S. Environmental Protection Agency, (U.S. EPA) [Ref 6] which involves flowing diluted engine exhaust through deionized or purified water contained in glass impingers and analyzing this solution by gas chromatography .
- 1.3 All definitions and abbreviations are contained in Appendix 2 of these test procedures.

2. METHOD SUMMARY

- 2.1 The samples are received by the laboratory in impingers. Compound separation and analysis are performed using a GC. The sample is injected into the GC by means of a liquid autosampler. Separation of the sample mixture into its components is performed by a temperature-programmed capillary column. A FID is used for alcohol detection and quantification.
- 2.2 The computerized GC data system identifies the alcohol associated with each peak. The alcohol concentrations are determined by integrating the peak areas and using response factors determined from external standards.

3. INTERFERENCES AND LIMITATIONS

- 3.1 An interferent is any component present in the sample with a retention time similar to that of any target alcohol described in this method. To reduce interference error, proof of chemical identity may require periodic confirmations using an alternate method and/or instrumentation, e.g., GC/MS.

- 3.2 The concentration of the alcohols in the range of interest is stable for up to six days as long as the samples are sealed and refrigerated at a temperature below 40°F.

4. INSTRUMENTATION AND APPARATUS

- 4.1 For each mode of the CVS test, two sampling impingers, each containing a known amount of deionized or purified water (e.g. 15 mL for this procedure), are used to contain the sample.

4.1.1 A temperature-programmable GC, equipped with a DB-Wax Megabore column (30 m, 0.53 mm ID, 1.0 μ film thickness) and FID, is used. Other columns may be used, provided the alternate(s) can be demonstrated to be equivalent or better with respect to precision, accuracy and resolution of all the target alcohols.

4.1.2 A liquid autosampler is required.

4.1.3 A PC-controlled data acquisition system for quantifying peak areas is required.

5. REAGENTS AND MATERIALS

- 5.1 Methanol shall have a purity of 99.9 percent, or be high performance liquid chromatography grade, EM Science or equivalent.

5.2 Ethanol shall be absolute, ACS reagent grade.

5.3 ASTM Type I purified or Type II deionized water shall be used.

5.4 Stock solutions are prepared gravimetrically or volumetrically by diluting methanol and ethanol with deionized or purified water, e.g., for this method a typical stock solution contains approximately 10 mg/mL of each target alcohol. Stock solutions must be replaced at least every six months.

5.4.1 A **calibration standard** within the expected concentration range of the samples is prepared by successive dilutions of the stock solution with deionized or purified water; 3 to 50 μ g/mL is typical, depending on fuel type. Calibration standards must be replaced at least every week.

5.4.2 A **control standard** containing all target alcohols is prepared by successive dilutions of a stock solution different from that of Section 5.4.1. This standard, at an approximate concentration of the samples, is used to monitor the precision of the analysis of each target alcohol. Control standards must be replaced at least every week.

- 5.4.3 Standards used for linearity and LOD determinations (Section 8) are also prepared by successive dilutions of an appropriate level stock solution.
- 5.4.4 All standards should be refrigerated at a temperature below 40°F during storage.
- 5.5 Gas requirements.
 - 5.5.1 Air shall be “Zero” grade (<1 ppmC total hydrocarbon contamination) or better.
 - 5.5.2 Nitrogen shall have a minimum purity of 99.998 percent.
 - 5.5.3 Helium shall have a minimum purity of 99.995 percent.
 - 5.5.4 Hydrogen shall have a minimum purity of 99.995 percent.

6. PROCEDURE

- 6.1 Each of the graduated sampling impingers is filled with 15 mL of deionized or purified water.
- 6.2 The impingers are placed in an ice bath during the sample collection.
- 6.3 After sampling, the solution contained in each impinger is transferred to a vial and sealed.
 - 6.3.1 Samples shall be refrigerated at a temperature below 40°F if immediate analysis is not feasible, or if reanalysis at a later date may be required.
- 6.4 One microliter aliquots of unmodified samples are injected via autosampler into a GC. Suggested standard operating conditions for the GC are:

| | |
|---------------------|---|
| Column: | DB-Wax, 30 m, 0.53 mm ID, 1.0μ film thickness |
| Carrier gas flow: | Helium at 5 mL/min |
| Make-up gas flow: | Nitrogen at 25 mL/min |
| Detector: | FID, hydrogen at 30 mL/min and air at 300 mL/min |
| Injector: | Packed column injector with Megabore adapter insert; on-column injection |
| Column temperature: | 50°C (1 min), 50°C to 70°C (5°C/min), 70°C to 110°C (15°C/min), 110°C (4 min) |
| Data system: | PC-based data acquisition system |

- 6.5 Samples containing compounds having concentrations above the documented range of instrument linearity must be diluted and reanalyzed.
- 6.6 The peak integrations are corrected as necessary in the data system. Any misplaced baseline segments are corrected in the reconstructed chromatogram.
- 6.7 The peak identifications provided by the computer are checked and corrected if necessary.

7. CALCULATIONS

- 7.1 The concentration of each target alcohol, in $\mu\text{g/mL}$, is determined by the following calculation that compares the sample peak area with that of an external standard:

$$\text{Concentration } (\mu\text{g/mL})_{\text{sample}} = \text{Peak Area}_{\text{sample}} \times \text{Response Factor}$$

where the response factor (RF) is calculated during the calibration by:

$$\text{RF} = \frac{\text{Concentration}_{\text{standard}} (\mu\text{g/mL})}{\text{Peak Area}_{\text{standard}}}$$

- 7.2 This concentration is then used to calculate the total amount of alcohol in each impinger:

$$\text{Mass } (\mu\text{g}) = \text{Concentration } (\mu\text{g/mL}) \times \text{Impinger volume (mL)}$$

- 7.3 An internal standard method may also be used.

8. QUALITY CONTROL

- 8.1 Blank Run - A deionized or purified water blank is run each analysis day. All target alcohol concentrations from the blank analysis must be below the LOD before the analysis may proceed.
 - 8.1.1 If the blank shows a peak greater than the limit of detection (LOD) in the region of interest, the source of the contamination must be investigated and remedied.
- 8.2 Calibration Run - The calibration standard is analyzed each analysis day to generate the response factor used to quantify the sample concentrations.
- 8.3 Control Standard Run - The quality control standard is analyzed at least once each analysis day. Measurements of all target alcohols in the control standard must fall within the control limits to ensure the validity of the sample analyses that day. To

meet this requirement, it may be necessary to inspect and repair the GC, and rerun the calibration and/or control standards.

- 8.4 Control Charts - A quality control chart is maintained for each analyte in the control standard. The control charts, used on a daily basis, establish that the method is “in-control”. The following describes how to construct a typical control chart:
1. Obtain at least 20 daily control standard results;
 2. Calculate the control standard mean concentration and standard deviation for the target analyte; and
 3. Create a control chart for the target analyte by placing the concentration on the Y-axis and the date on the X-axis. Establish upper and lower warning limits at either two standard deviations (2s) or 5 percent, whichever is greater, above and below the average concentration. Establish upper and lower control limits at either three standard deviations (3s) or 5 percent, whichever is greater, above and below the average concentration.
 4. A control standard measurement is considered to be out-of-control when the analyzed value exceeds the control limit or two successive control standard measurements of the same analyte exceed the warning limit.
 5. If 20 control standard results are not yet available to create a control chart (e.g., the control standard was expended and replaced with a different concentration standard prior to obtaining 20 points with the new standard), measurements must be within 10% of the theoretical concentration.

The measured concentrations of all target analytes contained in the control standard must be within the control limits (“in-control”) for the sample results to be considered acceptable.

- 8.5 Duplicates - A duplicate analysis of one sample is performed at least once per analysis day. The relative percent difference (RPD) is calculated for each duplicate run:

$$RPD(\%) = \frac{|\text{Difference between duplicate and original measurements}|}{\text{Average of duplicate and original measurements}} * 100$$

For each compound, the allowable RPD depends on the average concentration level for the duplicate runs, as shown in the following table:

| <u>Average Measurement for Duplicate Runs</u> | | <u>Allowable RPD (%)</u> |
|---|-----------|--------------------------|
| 1 to 10 | times LOD | 100 |
| 10 to 20 | “ “ | 30 |
| 20 to 50 | “ “ | 20 |
| Greater than 50 | “ “ | 15 |

If the results of the duplicate analyses do not meet these criteria for all target alcohols, the sample may be reanalyzed. If reanalysis is not feasible or if the criteria are still not met on reanalysis, all sample results for that analysis day are invalid.

- 8.6 Linearity - A multipoint calibration to confirm instrument linearity is performed for all target alcohols for new instruments, after making instrument modifications that can affect linearity, and at least once every year. The multipoint calibration consists of at least five concentration or mass loading levels, each above the LOD, distributed over the range of expected sample concentration. Each concentration level is measured at least twice. A linear regression analysis is performed using concentration and area counts to determine the regression correlation coefficient (r). The r must be greater than 0.995 to be considered sufficiently linear for one point calibrations.
- 8.7 Limit of Detection - The LOD for the target alcohols must be determined for new instruments, after making instrument modifications that can affect the LOD and at least once every year. To make the calculations, it is necessary to perform a multipoint calibration consisting of at least four “low” concentration levels, each above the expected LOD, with at least five replicate determinations of the lowest concentration standard. A linear regression is performed and the standard deviation (in area counts) of the lowest concentration standard determined. The standard deviation is converted to concentration units using the slope of the linear regression:

$$s = s_a \div m$$

where m is the slope of the linear regression, s is the standard deviation (in concentration units) of the lowest concentration standard and s_a is the standard deviation (in area counts) of the lowest concentration standard.

The LOD must be calculated using the following equation [Ref. 12]:

$$\text{LOD} = t * s$$

where s is the standard deviation (in concentration units) of at least five replicate determinations of the lowest concentration standard and t is the t-factor for 99 percent confidence for a one-sided normal (Gaussian) distribution. The number of

degrees of freedom is equal to the number of replicates, minus one. An abbreviated t-table is:

| <u>Degrees of Freedom</u> | <u>t-value</u> |
|---------------------------|----------------|
| 4 | 3.7 |
| 5 | 3.4 |
| 6 | 3.1 |
| 7 | 3.0 |

The lowest standard must be of a concentration of one to five times the estimated LOD.

- 8.7.1 The maximum allowable LOD for each alcohol is 0.10 µg/mL. The calculated laboratory LOD must be equal to or lower than the maximum allowable LOD. All peaks identified as target compounds that are equal to or exceed the maximum allowable LOD must be reported. If the calculated laboratory LOD is less than the maximum allowable LOD, the laboratory may choose to set its reporting limit at the maximum allowable LOD, the calculated laboratory LOD, or any level in between.
- 8.7.2 For the purpose of calculating the total mass of all species, the concentrations of the compounds below the LOD are considered to be zero.

Part D

DETERMINATION OF C₂ TO C₅ HYDROCARBONS IN AUTOMOTIVE SOURCE SAMPLES BY GAS CHROMATOGRAPHY

METHOD NO. 1002

1. INTRODUCTION

- 1.1 This document describes a gas chromatographic method of analyzing C₂ to C₅ hydrocarbons (light-end hydrocarbons) in the ppbC range from automotive source samples. This method does not include sample collection procedures [Ref. 8]. The “target” hydrocarbons that shall be analyzed and reported by this method and Method 1003 are listed in Appendix 1. All compounds on this list, when measured in concentrations above the LOD, shall be measured and reported (“targeted”) by either Method 1002 or Method 1003. Each laboratory should divide the list into light-end (Method 1002) and mid-range (Method 1003) hydrocarbons in the manner that best suits the laboratory instrumentation. All compounds on the list not targeted by Method 1002 must be targeted by Method 1003.
- 1.2 All definitions and abbreviations are contained in Appendix 2 of these test procedures.

2. METHOD SUMMARY

- 2.1 This is a method intended for routine analysis.
- 2.2 The samples are received by the laboratory in Tedlar bags, which are sub-sampled into a GC for separation and analysis.
- 2.3 The gas chromatographic analysis is performed on an Alumina (Al₂O₃) PLOT column temperature programmed from 0°C to 200°C. An FID is used for detection and quantification.
- 2.4 The sample is injected into the GC by means of gas sampling valves. Separation of the sample hydrocarbon mixture into its components takes place in the chromatographic column. The chromatographic column and the corresponding operating parameters described in this method normally provide complete resolution of most target compounds.
- 2.5 The computerized GC data acquisition system identifies the hydrocarbons associated with each peak. The hydrocarbon concentrations are determined by integrating the peak areas and using response factors determined from NIST-traceable standards.

3. **INTERFERENCES AND LIMITATIONS**

- 3.1 An interferent is any component present in the sample with a retention time very similar to that of any target hydrocarbon described in this method. To reduce interference error, proof of chemical identity may require periodic confirmations using an alternate method and/or instrumentation, e.g., GC/MS, PID, different column, etc.
- 3.2 To maximize sample integrity, sample bags should not leak or be exposed to bright light or excessive heat. Sampling bags must be shielded from direct sunlight to avoid photochemically induced reactions of any reactive hydrocarbons. The compound 1,3-butadiene, resulting mostly during cold-start testing is unstable. Therefore all cold-start samples must be analyzed within 8 hours; all other samples must be analyzed within 24 hours, although analysis within 8 hours is recommended.

4. **INSTRUMENTS AND APPARATUS**

- 4.1 Tedlar bags, 2 mil in thickness, nominally 5 to 10 liters in capacity and equipped with quick-connect fittings, are used to contain the samples.
- 4.2 For manual sub-sampling into a GC, a ground glass syringe is used to transfer gaseous samples from Tedlar bags to the GC sample inlet. For automated systems, a sample loop is used to transfer gaseous samples from the Tedlar bag to the sample inlet of the GC. Sample aliquot size is chosen based on considerations of instrument sensitivity and/or linearity.
- 4.3 A temperature-programmable GC equipped with a gas sampling valve system, a FID, and accessories is required.
- 4.4 An Alumina PLOT column (50 m x 0.32 mm) is used. A wax precolumn is recommended to prevent water damage to the PLOT column. Other columns may be used, provided the alternate(s) can be demonstrated to be equivalent or better with respect to precision, accuracy and resolution of all the target hydrocarbons.
- 4.5 A sample trap capable of being cryogenically cooled may be used.
- 4.6 Data acquisition software is used to integrate peak areas to determine hydrocarbon concentrations.

5. REAGENTS AND MATERIALS

- 5.1 Helium shall have a minimum purity of 99.995 percent. Higher purity helium may be required to achieve the LOD required by Section 8.7.1.
- 5.2 Hydrogen shall have a minimum purity of 99.995 percent.
- 5.3 Air shall be "Zero" grade (<1 ppmC total hydrocarbon contamination) or better.
- 5.4 Nitrogen shall have a minimum purity of 99.998 percent.
- 5.5 Calibration Standard - The quantitative calibration standard for all target hydrocarbons shall be propane at a concentration level between 0.25 and 1 ppm-mole and within the calculated linearity of the method. (See Section 8.6.) This propane standard must be a NIST-certified SRM or secondary NIST-traceable standard. A secondary standard is obtained by a comparison between a SRM and a candidate standard.
- 5.6 Control Standard - A quality control standard, containing at least ethene, propane, n-butane, and 2-methylpropene with concentrations between 0.2 and 3 ppmC based on a propane standard, is used for the following purposes:
 1. Daily update of control charts, and
 2. Daily determination of marker retention time windows.
- 5.7 A high concentration standard (higher than the calibration standard), containing the target hydrocarbons listed in Section 5.6, is used for linearity determinations. The high concentration standard must have concentrations verified against a NIST-traceable propane standard. (See Section 5.5 for the definition of NIST-traceable.) This verification can be performed at the laboratory performing the analysis.
- 5.8 A low concentration standard (one to five times the estimated LOD), containing the target hydrocarbons listed in Section 5.6, is used for LOD determinations. The low concentration standard must have concentrations verified against a NIST-traceable propane standard. (See Section 5.5 for the definition of NIST-traceable.) This verification can be performed at the laboratory performing the analysis.
 - 5.8.1 In lieu of a low concentration standard, a higher concentration standard may be diluted.
- 5.9 Liquid nitrogen may be required to cool the cryogenic sample trap and column oven where applicable.

6. PROCEDURE

- 6.1 The gaseous sample is analyzed for the target hydrocarbons C₂ through C₅.
- 6.2 Suggested standard operating conditions for the gas chromatograph are:
- | | |
|------------------------------|---|
| Helium carrier gas velocity: | 30 cm/sec at 200 ⁰ C |
| Nitrogen make-up gas flow: | sufficient such that the total flow of helium plus nitrogen is 30 mL/min |
| Hydrogen gas flow: | 30 mL/min |
| Air flow: | 300 mL/min |
| Sample valve temperature: | 150 ⁰ C (PLOT column) |
| Column temperature: | 0 ⁰ C (hold 7 min), 10 ⁰ C/min to 200 ⁰ C (hold 15 min) |
| Detector temperature: | 250 ⁰ C |
| Injector temperature: | 150 ⁰ C |
- 6.3 For automated systems, connect the samples to the GC and begin the analytical process.
- 6.4 Introduce the sample into the carrier gas stream through the injection valve.
- 6.5 Each separated component exits from the column into the FID where a response is generated.
- 6.6 Concentrations of hydrocarbons are calculated using data acquisition/ processing software that uses calibration data from the NIST-traceable propane calibration standard.
- 6.7 For compounds having concentrations above the documented range of instrument linearity, a smaller aliquot must be taken (for manual systems, a smaller syringe or smaller loop; for automated systems, a smaller loop) or the sample must be diluted.
- 6.8 The peak integrations are corrected as necessary in the data system. Any misplaced baseline segments are corrected in the reconstructed chromatogram.
- 6.9 The peak identifications provided by the computer are checked and corrected if necessary.
- 6.10 Target compounds that coelute are reported as the major component, as determined by the analysis of several samples by GC/MS or other methods. An exception to this is m- and p-xylene, where GC/MS data and fuel profiles are used to determine the relative contribution of each component to the peak. This method was used to determine the m- and p-xylene MIR value given in Appendix 1.

- 6.11 The Alumina PLOT column is programmed to 200°C to assure all compounds are eluted before the next run.

7. CALCULATIONS

- 7.1 The target hydrocarbon concentrations, in ppbC, are calculated by the data system using propane as an external standard.

$$\text{Concentration}_{\text{sample}} (\text{ppbC}) = \text{Peak Area}_{\text{sample}} \times \text{Response Factor}$$

where the response factor (RF) is calculated during daily calibration by:

$$\text{RF} = \frac{\text{Concentration of NIST-traceable propane standard, ppbC}}{\text{Area of propane peak}}$$

8. QUALITY CONTROL

- 8.1 Blank Run - A blank (pure nitrogen or helium) is run each analysis day. All target hydrocarbon concentrations from the blank analysis must be below the LOD before the analysis may proceed. As an alternative to a daily blank run, a daily partial blank check in tandem with a weekly blank run may be used. A partial blank check is defined as a check of the calibration standard run for contamination over all but the propane region of the chromatograph. The calibration standard must consist of only propane and make-up gas, with the concentration of all organic compounds except methane and propane below 2 percent of the propane standard concentration. The weekly blank run will provide a check on contamination in the propane region of the chromatograph.

8.1.1 If the blank shows a peak greater than the limit of detection (LOD) in the region of interest, the source of contamination must be investigated and remedied.

- 8.2 Calibration Run - The calibration standard is analyzed each analysis day to generate the response factor used to quantify the sample concentrations.
- 8.3 Control Standard Run - The quality control standard is analyzed at least once each analysis day. Measurements of all compounds specified in Section 5.6 must fall within the control limits to ensure the validity of the sample analyses that day. To meet this requirement, it may be necessary to inspect and repair the GC, and rerun the calibration and/or control standards.
- 8.4 Control Charts - A quality control chart is maintained for each component of the control standard listed in Section 5.6. The control charts, used on a daily basis, establish that the method is “in- control.” The following describes how to construct a typical control chart:

1. Obtain at least 20 daily control standard results;
2. Calculate the control standard mean concentration and standard deviation for the target hydrocarbon; and
3. Create a control chart for the target hydrocarbon by placing the concentration on the Y-axis and the date on the X-axis. Establish upper and lower warning limits at either two standard deviations (2s) or 5 percent, whichever is greater, above and below the average concentration. Establish upper and lower control limits at either three standard deviations (3s) or 5 percent, whichever is greater, above and below the average concentration.
4. A control standard measurement is considered to be out-of-control when the analyzed value exceeds the control limit or two successive control standard measurements of the same analyte exceed the warning limit.
5. If 20 control standard measurements are not yet available to create a control chart (e.g., the control standard was expended and replaced prior to obtaining 20 points with the new standard), measurements must be within 10% of the certified concentration. If the control standard is not a NIST standard, the cylinder should be certified by the laboratory against a NIST standard.

The measured concentrations of all target hydrocarbons contained in the control standard must be within the control limits (in-control) for the sample results to be considered acceptable.

- 8.5 Duplicates - A duplicate analysis of one sample is performed at least once per analysis day. The relative percent difference (RPD) is calculated for each duplicate run:

$$\text{RPD (\%)} = \frac{|\text{Difference between duplicate and original measurements}|}{\text{Average of duplicate and original measurements}} \times 100$$

For each compound specified in Section 5.6, the allowable RPD depends on the average concentration level for the duplicate runs, as shown in the following table:

| <u>Average Measurement for the Duplicate Runs</u> | | | <u>Allowable RPD (%)</u> |
|---|-------|-----|--------------------------|
| 1 to 10 | times | LOD | 100 |
| 10 to 20 | “ | “ | 30 |
| 20 to 50 | “ | “ | 20 |
| Greater than 50 | “ | “ | 15 |

If the results of the duplicate analyses do not meet these criteria for all compounds specified in Section 5.6, the sample may be reanalyzed. If reanalysis is not feasible or if the criteria are still not met on reanalysis, all sample results for that analysis day are invalid.

- 8.6 Linearity - A multipoint calibration to confirm instrument linearity is performed for the target hydrocarbons in the control standard for new instruments, after making instrument modifications that can affect linearity, and at least once every year unless a daily check of the instrument response indicates that the linearity has not changed. To monitor the instrument response, a quality control chart is constructed, as specified in Section 8.4, except using calibration standard area counts rather than control standard concentrations. When the standard area counts are out-of-control, corrective action(s) must be taken before analysis may proceed. The multipoint calibration consists of at least five concentration or mass loading levels (using smaller or larger volume sample sizes of existing standards is acceptable), each above the LOD, distributed over the range of expected sample concentration. Each concentration level is measured at least twice. A linear regression analysis is performed using concentration and average area counts to determine the regression correlation coefficient (r). The r must be greater than 0.995 to be considered sufficiently linear for one-point calibrations.
- 8.7 Limit of Detection – The LOD for the target hydrocarbons in the control standard must be determined for new instruments and after making instrument modifications that can affect linearity and/or sensitivity and at least once every year unless a daily check of the instrument response indicates that the LOD has not changed. To monitor the instrument response, a quality control chart is constructed, as specified in Section 8.4, except using calibration standard area counts rather than control standard concentrations. When the calibration standard area counts are out-of-control, investigation and/or corrective action(s) must be taken. To make the calculations, it is necessary to perform a multipoint calibration consisting of at least four “low” concentration levels, each above the LOD, with at least five replicate determinations of the lowest concentration standard. A linear regression is performed and the standard deviation is converted to concentration units using the slope of the linear regression:

$$s = s_a \div m$$

where m is the slope of the linear regression, s is the standard deviation (in concentration units) of the lowest concentration standard and s_a is the standard deviation (in area counts) of the lowest concentration standard.

The LOD must be calculated using the following equation [Ref. 12]:

$$\text{LOD} = t * s$$

where s is the standard deviation (in concentration units) of at least five replicate determinations of the lowest concentration standard and t is the t-factor for 99 percent confidence for a one-sided normal (Gaussian) distribution. The number of degrees of freedom is equal to the number of replicates, minus one. An abbreviated t-table is:

| <u>Degrees of Freedom</u> | <u>t-value</u> |
|---------------------------|----------------|
| 4 | 3.7 |
| 5 | 3.4 |
| 6 | 3.1 |
| 7 | 3.0 |

The lowest standard must be of a concentration of one to five times the estimated LOD.

- 8.7.1 The maximum allowable LOD for each compound is 5 ppbC. The calculated laboratory LOD must be equal to or lower than the maximum allowable LOD. All peaks identified as target compounds that are equal to or exceed the maximum allowable LOD must be reported. If the calculated laboratory LOD is less than the maximum allowable LOD, the laboratory may choose to set its reporting limit at the maximum allowable LOD, the calculated laboratory LOD, or any level in between.
- 8.7.2 For the purpose of calculating the total mass of all species, the concentrations of all compounds below the LOD are considered to be zero.
- 8.8 Method No. 1002/Method No. 1003 Crossover Check - For each sample, a compound shall be measured by both Method No. 1002 and Method No. 1003. The crossover compound shall be a compound that can reasonably be expected to be found and measured by both methods in the laboratory performing the analysis. The results obtained by the two methods should be compared and an acceptance criteria set for the relative percent difference.

Part E

DETERMINATION OF C₆ TO C₁₂ HYDROCARBONS IN AUTOMOTIVE SOURCE SAMPLES BY GAS CHROMATOGRAPHY

METHOD NO. 1003

1. INTRODUCTION

- 1.1 This document describes a gas chromatographic method of analyzing C₆ to C₁₂ hydrocarbons (mid-range hydrocarbons) in the ppbC range from automotive source samples. This method does not include sample collection procedures [Ref. 7]. The target hydrocarbons that shall be analyzed and reported by this method and Method 1002 are listed in Appendix 1. All compounds on this list, when measured in concentrations above the LOD, shall be measured and reported (“targeted”) by either Method 1002 or Method 1003. Each laboratory should divide the list into light-end (Method 1002) and mid-range (Method 1003) hydrocarbons in the manner that best suits the laboratory instrumentation. All compounds on the list not targeted by Method 1003 must be targeted by Method 1002.
- 1.2 All definitions and abbreviations are contained in Appendix 2 of these test procedures.

2. METHOD SUMMARY

- 2.1 This is a method intended for routine analysis.
- 2.2 The samples are received by the laboratory in Tedlar bags, which are sub-sampled into a GC for separation and analysis.
- 2.3 The gas chromatographic analysis is performed through a temperature-programmed capillary column. A FID is used for detection.
- 2.4 The sample is injected into the GC by means of gas sampling valves. Separation of the sample hydrocarbon mixture into its components takes place in the chromatographic column. The chromatographic column and the corresponding operating parameters described in this method normally provide complete resolution of most target hydrocarbons.
- 2.5 The computerized GC data acquisition system identifies the hydrocarbons associated with each peak. The hydrocarbon concentrations are determined by integrating the peak areas and using a response factor determined from NIST-traceable standards.

3. INTERFERENCES AND LIMITATIONS

- 3.1 An interferent is any component present in the sample with a retention time similar to that of any target hydrocarbon described in this method. To reduce interference error, proof of chemical identity may require periodic confirmations using an alternate method and/or instrumentation, e.g., GC/MS, PID, different column, etc.
- 3.2 The concentration of hydrocarbons in the range of interest is stable for at least 24 hours in the Tedlar sampling bags, provided the sample bags do not leak and are not exposed to bright light or excessive heat. Sampling bags must be shielded from direct sunlight to avoid photochemically induced reactions of any reactive hydrocarbons. Samples must be analyzed within 24 hours.

4. INSTRUMENTATION AND APPARATUS

- 4.1 Tedlar bags, 2 mil in thickness, nominally 5 to 10 liters in capacity and equipped with quick-connect fittings, are used to contain the samples.
- 4.2 For manual sub-sampling into a GC, a ground glass syringe is used to transfer gaseous samples from Tedlar bags to the GC sample inlet. For automated systems, a sample loop is used to transfer gaseous samples from the Tedlar bag to the sample inlet of the GC. Sample aliquot size is chosen based on considerations of instrument sensitivity and/or linearity.
- 4.3 The GC is equipped with a FID, and a gas sampling valve system.
- 4.4 A non-polar capillary column [e.g., J&W DB-1, 60 m x 0.32 mm ID, film thickness 1.0 μ] is used. Other columns may be used, provided the alternate(s) can be demonstrated to be equivalent or better with respect to precision, accuracy and resolution of all the target hydrocarbons.
- 4.5 A sample trap capable of being cryogenically cooled may be used.
- 4.6 A computer-controlled data acquisition system is required for quantifying peak areas.

5. REAGENTS AND MATERIALS

- 5.1 Helium shall have a minimum purity of 99.995 percent. Higher purity helium may be required to achieve the LOD required by Section 8.7.1.
- 5.2 Hydrogen shall have a minimum purity of 99.995 percent.
- 5.3 Air shall be "Zero" grade (<1 ppmC total hydrocarbon contamination) or better.

- 5.4 Nitrogen shall have a minimum purity of 99.998 percent.
- 5.5 Calibration Standard - The quantitative calibration standard for all target hydrocarbons shall be propane at a concentration level between 0.25 and 1 ppm-mole and within the calculated linearity of the method. (See Section 8.6.) This propane standard must be a NIST-certified SRM or secondary NIST-traceable standard. A secondary standard is obtained by a comparison between a SRM and a candidate standard.
- 5.6 Control Standard - A quality control standard, containing at least n-hexane, n-octane, n-decane, benzene, toluene, and m- or p-xylene with concentrations between 0.2 and 1 ppmC based on a propane standard, is used for the following purposes:
1. Daily update of control charts, and
 2. Daily determination of marker retention time windows.
- 5.7 A high concentration standard (higher than the calibration standard), containing the target hydrocarbons listed in Section 5.6, is used for linearity determinations. The high concentration standard must have concentrations verified against a NIST-traceable propane standard. (See Section 5.5 for the definition of NIST-traceable.) This verification can be performed at the laboratory performing the analysis.
- 5.8 A low concentration standard (one to five times the estimated LOD), containing the target hydrocarbons listed in Section 5.6, is used for LOD determinations. The low concentration standard must have concentrations verified against a NIST-traceable propane standard. (See Section 5.5 for the definition of NIST-traceable.) This verification can be performed at the laboratory performing the analysis.
- 5.8.1 In lieu of a low concentration standard, a higher concentration standard may be diluted.
- 5.9 Liquid nitrogen may be required to cool the cryogenic trap and column oven where applicable.

6. PROCEDURE

6.1 Typical operating conditions.

6.1.1 Suggested operating conditions for the manual GC are:

| | |
|---|--|
| Helium carrier gas velocity: | 30 cm/sec at 200°C |
| Nitrogen make-up gas flow: | sufficient such that the total flow of helium plus nitrogen is 30 mL/min |
| Hydrogen gas flow (for FID): | 30 mL/min |
| “Zero” air gas flow (for FID): | 300 mL/min |
| Autozero FID at: | 0.0 min |
| Range 11, Attenuation 8 (or another suitable value) | |
| Sample valve temperature: | 150°C |
| Injector temperature: | 150°C |
| Column entrance port temperature: | 95°C |
| Detector temperature: | 250°C |
| Column temperature: | Initial temperature 0°C; 10°C/min to 200°C |

6.1.2 Suggested operating conditions for the automated GC are:

| | |
|---|---|
| Helium carrier gas velocity: | 30 cm/sec at 200°C |
| Nitrogen make-up gas flow: | sufficient such that the total flow of helium plus nitrogen is 30 mL/min |
| Hydrogen gas flow (for FID): | 30 mL/min |
| “Zero” air gas flow (for FID): | 300 mL/min |
| Range 12, attenuation 8 (or another suitable value) | |
| Sample valve temperature: | 150°C |
| Detector temperature: | 300°C |
| Injector temperature: | 150°C |
| Column temperature: | Initial temperature -50°C (5 min), 5°C/min to 50°C, 10°C/min to 200°C |

6.2 Data Reduction

6.2.1 The results are calculated from the FID responses.

6.2.2 The results are examined to see that the peaks are correctly integrated.

6.2.3 After running a particularly “dirty” sample, the analyst should run a blank before proceeding to the next sample as there may be sample carryover, or flush the sampling system with air.

6.2.4 The peak identifications provided by the computer are reviewed and, if necessary, corrected using the following procedure and criteria:

1. The relative retention indices from GC/MS analyses are used to help confirm peak identifications.
 2. The primary peak identification is done by the computer using the relative retention times based on reference calibration runs.
 3. Confirm that the relative peak heights of the sample run (“fingerprint”) match the typical fingerprint seen in past sample runs.
 4. Compare the relative retention times of the sample peaks with those of reference runs.
 5. Any peak with a reasonable doubt is labeled 'Unidentified'.
- 6.2.5 For compounds having concentrations above the documented range of instrument linearity, a smaller aliquot must be taken (for manual systems, a smaller syringe or smaller loop; for automated systems, a smaller loop) or the sample must be diluted.
- 6.2.6 The concentrations of the hydrocarbons are calculated using data acquisition/ processing software which uses calibration data from a NIST-traceable propane calibration standard.
- 6.2.7 Target compounds that coelute are reported as the major component, as determined by the analysis of several samples by GC/MS or other methods. An exception to this is m- and p-xylene, where GC/MS data and fuel profiles are used to determine the relative contribution of each component to the peak. This method was used to determine the m- and p-xylene MIR value given in Appendix 1.

7. CALCULATIONS

- 7.1 The target hydrocarbon concentrations, in ppbC, are calculated by the data system using propane as an external standard.

$$\text{Concentration}_{\text{sample}} \text{ (ppbC)} = \text{Peak Area}_{\text{sample}} * \text{Response Factor}$$

where the Response Factor (RF) is calculated during daily calibration by:

$$\text{RF} = \frac{\text{Concentration of NIST – traceable propane standard, ppbC}}{\text{Area of propane peak}}$$

8. QUALITY CONTROL

- 8.1 Blank Run - A blank (pure nitrogen or helium) is run each analysis day. All target hydrocarbon concentrations from the blank analysis must be below the LOD before the analysis may proceed. As an alternative to a daily blank run, a daily partial blank check in tandem with a weekly blank run may be used. A partial blank check is defined as a check of the calibration standard run for contamination over all but the propane region of the chromatograph. The calibration standard must consist of only propane and make-up gas, with the concentration of all organic compounds except methane and propane below 2 percent of the propane standard concentration. The weekly blank run will provide a check on contamination in the propane region of the chromatograph.
- 8.1.1 If the blank shows a peak greater than the limit of detection (LOD) in the region of interest, the source of the contamination must be investigated and remedied.
- 8.2 Calibration Run - The calibration standard is analyzed each analysis day to generate the response factor used to quantify the sample concentrations.
- 8.3 Control Standard Run - The quality control standard is analyzed at least once each analysis day. Measurements of all compounds specified in Section 5.6 must fall within the control limits to ensure the validity of the sample analyses that day. To meet this requirement, it may be necessary to inspect and repair the GC, and rerun the calibration and/or control standards.
- 8.4 Control Charts - A quality control chart is maintained for each component of the control standard listed in Section 5.6. The control charts, used on a daily basis, establish that the method is "in-control". The following describes how to construct a typical control chart:
1. Obtain at least 20 daily control standard results,
 2. Calculate the control standard mean concentration and standard deviation for the target hydrocarbon, and
 3. Create a control chart for the target hydrocarbon by placing the concentration on the Y-axis and the date on the X-axis. Establish upper and lower warning limits at either two standard deviations (2s) or 5 percent, whichever is greater, above and below the average concentration. Establish upper and lower control limits at either three standard deviations (3s) or 5 percent, whichever is greater, above and below the average concentration.
 4. A control standard measurement is considered to be out-of-control when the analyzed value exceeds the control limit or two successive control standard measurements of the same analyte exceed the warning limit.
 5. If 20 control standard measurements are not yet available to create a control chart (e.g., the control standard was expended and replaced prior

to obtaining 20 points with the new standard), measurements must be within 10% of the certified concentration. If the control standard is not a NIST standard, the cylinder should be certified by the laboratory against a NIST standard.

The measured concentrations of all target hydrocarbons contained in the control standard must be within the control limits (in-control) for the sample results to be considered acceptable.

- 8.5 Duplicates - A duplicate analysis of one sample is performed at least once per analysis day. The relative percent difference (RPD) is calculated for each duplicate run:

$$RPD(\%) = \frac{|\text{Difference between duplicate and original measurements}|}{\text{Average of duplicate and original measurements}} * 100$$

For each compound specified in Section 5.6, the allowable RPD depends on the average concentration level for the duplicate runs, as shown in the following table:

| <u>Average Measurement for Duplicate Runs</u> | | <u>Allowable RPD (%)</u> |
|---|-----------|--------------------------|
| 1 to 10 | times LOD | 100 |
| 10 to 20 | “ “ | 30 |
| 20 to 50 | “ “ | 20 |
| Greater than 50 | “ “ | 15 |

If the results of the duplicate analyses do not meet these criteria for all compounds specified in Section 5.6, the sample may be reanalyzed. If reanalysis is not feasible or if the criteria are still not met on reanalysis, all sample results for that analysis day are invalid.

- 8.6 Linearity - A multipoint calibration to confirm instrument linearity is performed for all target hydrocarbons in the control standard for new instruments, after making instrument modifications that can affect linearity, and at least once every year unless a daily check of the instrument response indicates that the linearity has not changed. To monitor the instrument response, a quality control chart is constructed, as specified in Section 8.4, except using calibration standard area counts rather than control standard concentrations. When the standard area counts are out-of-control, corrective action(s) must be taken before analysis may proceed. The multipoint calibration consists of at least five concentration or mass loading levels (using smaller or larger volume sample sizes of existing standards is acceptable), each above the LOD, distributed over the range of expected sample concentration. Each concentration level is measured at least twice. A linear regression analysis is performed using concentration and average area counts to

determine the regression correlation coefficient (r). The r must be greater than 0.995 to be considered sufficiently linear for one point calibrations.

- 8.7 Limit of Detection - The LOD for the target hydrocarbons in the control standard must be determined for new instruments and after making instrument modifications that can affect linearity and/or sensitivity and at least once every year unless a daily check of the instrument response indicates that the LOD has not changed. To monitor the instrument response, a quality control chart is constructed, as specified in Section 8.4, except using calibration standard area counts rather than control standard concentrations. When the calibration standard area counts are out-of-control, investigation and/or corrective action(s) must be taken. To make the calculations, it is necessary to perform a multipoint calibration consisting of at least four "low" concentration levels, each above the LOD, with at least five replicate determinations of the lowest concentration standard. A linear regression is performed and the standard deviation (in area counts) of the lowest concentration standard determined. The standard deviation is converted to concentration units using the slope of the linear regression:

$$s = s_a \div m$$

where m is the slope of the linear regression, s is the standard deviation (in concentration units) of the lowest concentration standard and s_a is the standard deviation (in area counts) of the lowest concentration standard.

The LOD must be calculated using the following equation [Ref. 12]:

$$\text{LOD} = t * s$$

where s is the standard deviation (in concentration units) of at least five replicate determinations of the lowest concentration standard and t is the t-factor for 99 percent confidence for a one-sided normal (Gaussian) distribution. The number of degrees of freedom is equal to the number of replicates, minus one. An abbreviated t-table is:

| <u>Degrees of Freedom</u> | <u>t-value</u> |
|---------------------------|----------------|
| 4 | 3.7 |
| 5 | 3.4 |
| 6 | 3.1 |
| 7 | 3.0 |

The lowest standard must be of a concentration of one to five times the estimated LOD.

- 8.7.1 The maximum allowable LOD for each compound is 5 ppbC. The calculated laboratory LOD must be equal to or lower than the maximum allowable LOD. All peaks identified as target compounds that are equal to

or exceed the maximum LOD must be reported. If the calculated laboratory LOD is less than the maximum allowable LOD, the laboratory may choose to set its reporting limit at the maximum allowable LOD, the calculated laboratory LOD, or any level in between.

- 8.7.2. For the purpose of calculating the total mass of all species, the concentrations of all compounds below the LOD are considered to be zero.
- 8.8 Method No. 1002/Method No. 1003 Crossover Check - For each sample a compound shall be measured by both Method No. 1002 and Method No. 1003. The crossover compound shall be a compound that can reasonably be expected to be found and measured by both methods in the laboratory performing the analysis. The results of the two analyses should be compared and an acceptance criteria set for the relative percent difference.

Part F

**DETERMINATION OF ALDEHYDE AND KETONE COMPOUNDS
IN AUTOMOTIVE SOURCE SAMPLES
BY HIGH PERFORMANCE LIQUID CHROMATOGRAPHY**

METHOD NO. 1004

1. INTRODUCTION

- 1.1 This document describes a method of analyzing automotive engine exhaust for aldehyde and ketone compounds (carbonyls) using impingers, containing acidified 2,4-dinitrophenylhydrazine (DNPH)-absorbing solution, or DNPH-impregnated cartridges. Carbonyl masses ranging between 0.02 to 200 µg are measured by this method. The “target” carbonyls that shall be analyzed and reported by this method are listed in Appendix 1. All of these carbonyl compounds, when measured in concentrations above the LOD, shall be reported.
- 1.2 This procedure is derived from a method used by Hull [Ref. 10].
- 1.3 All definitions and abbreviations are contained in Appendix 2 of these test procedures.

2. METHOD SUMMARY

- 2.1 The samples are received by the laboratory in sample collection cartridges or impingers. (See Section 4.2.) The absorbing solution (2,4-DNPH) complexes the carbonyl compounds into their diphenylhydrazone derivatives. The cartridges are then eluted with 5 mL acetonitrile.
- 2.2 Separation and analysis are performed using a HPLC with an ultraviolet detector.

3. INTERFERENCES AND LIMITATIONS

- 3.1 An interferent is any detectable compound present in the sample with a retention time very similar to that of any target carbonyl described in this method. To reduce interference error, proof of chemical identity may require periodic confirmations using an alternate method and/or instrumentation, e.g., alternative HPLC columns or mobile phase compositions.
- 3.2 If samples are not analyzed the same day as received, they must be refrigerated at a temperature below 40°F. Impinger solutions must first be transferred to glass bottles and sealed. Refrigerated samples are stable for up to 30 days.

- 3.3 When using the DuPont Zorbax or Supelco's Supelcosil columns described in Section 4.1.5, methyl ethyl ketone (MEK) and butyraldehyde tend to coelute.
- 3.4 When using the Delta Bond column described in Section 4.1.5, formaldehyde tends to coelute with an unknown, non-carbonyl compound, and the tolualdehyde isomers tend to coelute. The guard column for the Delta Bond column must also be changed frequently in order to prevent the coelution of butyraldehyde and methacrolein and to prolong the life of the column.

4. **INSTRUMENT AND APPARATUS**

- 4.1 The HPLC analytical system consists of the following:
 - 4.1.1 Dual high pressure pumps.
 - 4.1.2 Automated gradient controller or pump module controller.
 - 4.1.3 Temperature controller module for the column oven.
 - 4.1.4 A liquid autosampler.
 - 4.1.5 The primary system incorporates two DuPont Zorbax ODS or Supelco's Supelcosil columns in tandem and a guard column, (2 cm long packed with LC18 5 µm pellicular beads). The secondary system incorporates a Delta Bond AK (4.6 mm ID x 200 mm) packed column with a guard column 2 cm long packed with LC18 5 µm pellicular beads or equivalent). Other columns may be used, provided the alternate(s) can be demonstrated to be equivalent or better with respect to precision, accuracy and resolution of all target carbonyls.
 - 4.1.6 An ultraviolet/visible (UV/VIS) detector.
 - 4.1.7 Data system for peak integration.
- 4.2 Sample collection containers are glass impingers or DNPH-impregnated cartridges.

5. **REAGENTS AND MATERIALS**

- 5.1 Acetonitrile, HPLC grade, (Burdick and Jackson or equivalent).
- 5.2 Water, HPLC grade, (Burdick and Jackson or equivalent).
- 5.3 2,4-DNPH, purified, Radian Corporation or equivalent. Unpurified DNPH must be recrystallized twice from acetonitrile. The recrystallized DNPH is checked for contaminants by injecting a dilute solution of DNPH in contaminant-free acetonitrile into the HPLC.
- 5.4 Sulfuric acid, or perchloric acid, analytical reagent grade, (Baker Analyzed or equivalent).

- 5.5 The carbonyl/2,4-dinitrophenylhydrazone (DNPH) complexes [Ref. 11] listed in Table F1 may be purchased (e.g., Radian Corporation, in 1.2 mL ampules) or prepared in the laboratory. In-house standards must be recrystallized at least three times from 95 percent ethanol.

TABLE F-1
PROPERTIES OF CARBONYL/2,4-DNPH COMPLEXES

| Complex | Molecular Weight (g/mole) | Melting Point °C |
|---------------------|---------------------------|------------------|
| formaldehyde | 210.15 | 165-166 |
| acetaldehyde | 224.18 | 152-153 |
| acrolein | 236.19 | 165 ¹ |
| acetone | 238.20 | 125-127 |
| propionaldehyde | 238.20 | 144-145 |
| butyraldehyde | 252.23 | 119-120 |
| hexanaldehyde | 280.28 | |
| 106-107 | | |
| benzaldehyde | 286.25 | 240-242 |
| methyl ethyl ketone | 252.53 | 117-118 |
| methacrolein | 250.21 | 200-201 |
| crotonaldehyde | 250.21 | 185-188 |
| valeraldehyde | 266.26 | 107-108 |
| m-tolualdehyde | 300.27 | 212 |

- 5.6 Stock Calibration Standard - A stock calibration standard is prepared by diluting the target carbonyl/2,4-DNPH complexes with acetonitrile. A typical stock calibration standard contains 3.0 µg/mL of each target carbonyl compound. Stock calibration standards of other concentrations may also be used.
- 5.7 Working Standard - A working standard is prepared when required by diluting the stock calibration solution, making sure that the highest concentration of the standard is above the expected test level. Typically, the 3.0 µg/mL stock is diluted five times with acetonitrile in a volumetric flask to yield a 0.6 µg/mL solution.
- 5.8 Control Standard - A quality control standard, containing all target carbonyls/2,4-DNPH complexes within the typical concentration range of real samples, is analyzed to monitor the precision of the analysis of each target carbonyl. The control standard may be purchased, prepared in the laboratory from a stock solution different from the calibration standard, or prepared by batch mixing old samples, spiking it with a stock solution of target compounds, and

¹This compound has been known to decompose.

stirring for a minimum of 2 hours. If necessary, the solution is filtered using filter paper to remove precipitation. All target compounds except acrolein have been found to be stable in the control standard.

- 5.9 Standards used for linearity and LOD determinations (Section 8) may be purchased or prepared by dilutions of an appropriate level stock solution.

6. PROCEDURE

- 6.1 For systems collecting the samples via impingers, an absorbing solution is prepared by dissolving 0.11 - 0.13 grams of recrystallized DNPH in 1 L of HPLC grade acetonitrile. The absorbing solution should be prepared at least every two weeks. Each batch of acetonitrile used in this procedure is checked for oxygenated impurities by adding it to a contaminant-free dilute solution of DNPH and analyzing by HPLC.
- 6.1.1 In the laboratory, pipette 15 mL of the DNPH absorbing solution into each of the 30 mL impingers for each emission test. Add 0.1 mL of 2.85 N sulfuric acid or 0.15 mL of 3.8 M perchloric acid to each impinger.
- 6.2 For systems collecting the samples via cartridges, DNPH-impregnated cartridges shall be sealed and refrigerated, at a temperature less than 40°F, upon receipt from manufacturer, until ready for use.
- 6.2.1 At the exhaust volumes being sampled (1 L/min), a back-up cartridge may be required for CVS phase 1 but no back-up cartridge is needed for CVS phases 2 and 3.
- 6.3 After sampling uncap and place all impingers in preheated water at 70-80°C for 30 minutes (min) to complete derivatization. Heating is not required when using perchloric acid.
- 6.3.1 For cartridges, remove the caps and extract with 5 mL acetonitrile, running the extract into glass storage bottles.
- 6.4 Remove the impingers from the water bath and cool to room temperature. Replace any lost solvent by adding acetonitrile to the 15-mL mark.
- 6.4.1 Replacing lost solvent is not required when using an internal standard method (Section 7.4).
- 6.5 Transfer the solution from each impinger/cartridge to glass vials and seal with new septum screw caps.

- 6.6 Place the vials containing blank, working standard, control standard, and samples into the autosampler for subsequent injection into an HPLC. Suggested standard operating conditions for the HPLC are:

Primary System:

Columns: 4.6 mm ID x 250 mm x 1/4 inches OD Dupont Zorbax ODS or Supelco Supelcosil - two columns in series, Guard column - 2 cm long packed with C18 5 μ m pellicular

Column temperature: 40°C

Detector: UV/VIS at 360 nm

Sample volume: 10 μ L

Solvent A: acetonitrile

Solvent B: 10 percent (volume/volume) methanol in water

Flow: 1 mL/min

Program: 50 percent A, 50 percent B 0 (initial time)
65 percent A, 35 percent B 0 to 2 min
100 percent A, 0 percent B 2 to 22 min
50 percent A, 50 percent B 22 to 37 min

Under the above configuration, methyl ethyl ketone and butyraldehyde tend to coelute. In order to report these compounds, it is necessary to analyze the samples using a secondary system. The tolualdehyde isomers (m-, p-, and o-) are separated using this configuration. The reporting of tolualdehyde is addressed in Section 7.4.

Secondary System:

Columns: Delta Bond AK C18 (4.6 mm ID x 200 mm x 1/4 in OD) packed column, Guard column - 2 cm long packed with C18 5 μ m pellicular

Column temperature: 40°C

Detector: UV/VIS at 360 nm

Sample volume: 10 μ L

Solvent A: acetonitrile

Solvent B: acetonitrile in water, 35 percent (v/v)

Flow: 1.5 mL/min

Program: 0 percent A, 100 percent B 0 (initial time)
23 percent A, 77 percent B 0 - 5.5 min
46 percent A, 54 percent B 5.5 - 13 min
0 percent A, 100 percent B 13 to 30 min

This secondary system is not used to report all compounds because formaldehyde tends to coelute with a non-carbonyl compound. If this coelution is resolved, the secondary system may be used alone to analyze all carbonyl compounds. The tolualdehyde isomers, however, are not separated with this configuration. The reporting of tolualdehyde is addressed in Section 7.4.

Data System: The outputs from the UV/VIS detector are sent to a PC-controlled data acquisition system.

- 6.7 The peak integrations are corrected as necessary in the data system. Any misplaced baseline segments are corrected in the reconstructed chromatogram.
- 6.8 Samples containing compounds having concentrations above the documented range of instrument linearity must be diluted and reanalyzed.

7. CALCULATIONS

- 7.1 For each target carbonyl, the carbonyl mass is calculated from its carbonyl/2,4-DNPH mass.
- 7.2 The mass of each carbonyl compound, per impinger or cartridge, is determined by the following calculation:

$$\text{Mass}_{\text{sample}} = \text{Peak Area}_{\text{sample}} * \text{Response Factor} * \text{Impinger (or Cartridge) volume (mL)} * B$$

where B is the ratio of the molecular weight of the carbonyl compound to its 2,4-dinitrophenylhydrazone derivative and where the response factor (RF) for each carbonyl is calculated during the calibration by:

$$\text{RF} = \frac{\text{Concentration}_{\text{standard}} (\mu\text{g 2,4-DNPH species/mL})}{\text{Peak Area}_{\text{standard}}}$$

- 7.3 For tolualdehyde, the sum of all isomers present is reported as m-tolualdehyde.
 - 7.3.1 Under the conditions of the primary system in Section 6.6, the isomers are separated. The m-tolualdehyde response factor is applied to each peak and the sum reported as m-tolualdehyde.
 - 7.3.2 Under the conditions of the secondary system in Section 6.6, the isomers coelute. The m-tolualdehyde response factor is applied to the single tolualdehyde peak. This concentration is reported as m-tolualdehyde.
- 7.4 An internal standard method may also be used.

8. QUALITY CONTROL

8.1 Blank Runs

8.1.1 Reagent Blanks - The solvents used are of the highest HPLC grade and are tested for impurities when a new lot number is used. If this lot number is found to be acceptable, (no carbonyls present at concentrations at or above the LOD), daily blank analysis is not performed.

8.1.2 Carbonyl/2,4-DNPH Purity - The carbonyl/ 2,4- DNPHs are checked for purity by their melting points and their chromatograms (See Table F-1). Analysis of the solution of carbonyl/2,4-DNPH must yield only the peak of interest. No contaminant peaks above the LOD should be observed.

8.1.3 Field Blanks – One cartridge is analyzed as a field blank for each emission test. If the field blank shows a peak greater than the limit of detection (LOD) in the region of interest, the source of the contamination must be investigated and remedied.

8.1.4 Cartridge Blanks - At least one cartridge per batch is analyzed as a batch blank. If the cartridge blank shows a peak greater than the limit of detection (LOD) in the region of interest, the source of the contamination must be investigated and remedied.

8.2 Calibration Run - The calibration standard is analyzed each analysis day to generate the response factors used to quantify the sample concentrations.

8.3 Control Standard Run - The quality control standard is analyzed at least once each analysis day. Measurements of all target compounds in the control standard, except acrolein, must fall within the control limits to ensure the validity of the sample analyses that day. To meet this requirement, it may be necessary to rerun the calibration and control standards, and inspect and repair the HPLC.

8.4 Control Charts - A quality control chart is maintained for each component of the control standard. The control charts, used on a daily basis, establish that the method is “in- control.” The following describes how to construct a typical control chart:

1. Obtain at least 20 daily control standard results,
2. Calculate the control standard mean concentration, and standard deviation(s) for the target analyte, and
3. Create a control chart for the target analyte by placing the concentration on the Y-axis and the date on the X-axis. Establish an upper warning limit and a lower warning limit at two standard deviations (2s) above and below the average concentration. Establish an upper control limit and a lower

- control limit at three standard deviations (3s) above and below the average concentration.
4. Due to the low variability of the carbonyl control standard measurements, a control standard measurement is considered to be out-of-control when the analyzed value exceeds either the 3s limit, or the range of $\pm 10\%$ of the mean control measurement, whichever is greater, or if two successive control standard measurements of the same analyte exceed the 2s limit.
 5. If 20 control standard measurements are not yet available to create a control chart (e.g., the control standard was expended and replaced prior to obtaining 20 points with the new standard), measurements must be within 10% of the assay (purchased) or theoretical (prepared in-house) concentration.

The measured concentrations of all target analytes contained in the control standard must be within the control limits (in-control) for the sample results to be considered acceptable. No control requirements have been established for acrolein, since it has been shown to degrade over time.

- 8.5 Duplicates - A duplicate analysis of one sample is performed at least once per analysis day. The relative percent difference (RPD) is calculated for each duplicate run:

$$\text{RPD}(\%) = \frac{|\text{Difference between duplicate and original measurements}|}{\text{Average of duplicate and original measurements}} * 100$$

For each compound, the allowable RPD depends on the average concentration level for the duplicate runs, as shown in the following table:

| <u>Average Measurement for Duplicate Runs</u> | | <u>Allowable RPD (%)</u> |
|---|-----------|--------------------------|
| 1 to 10 | times LOD | 100 |
| 10 to 20 | “ “ | 30 |
| 20 to 50 | “ “ | 20 |
| Greater than 50 | “ “ | 15 |

If the results of the duplicate analyses do not meet these criteria for all target carbonyls, the sample may be reanalyzed. If reanalysis is not feasible or if the criteria are still not met on reanalysis, all sample results for that analysis day are invalid.

- 8.6 Linearity - A multipoint calibration to confirm instrument linearity is performed for all target analytes for new instruments, after making instrument modifications that can affect linearity, and at least once every year. The multipoint calibration consists of at least five concentration or mass loading levels (using smaller or larger volume sample sizes of existing standards is acceptable), each above the LOD, distributed over the range of expected sample concentration. Each concentration level is measured at least twice. A linear regression analysis is performed using concentration and average area counts to determine regression correlation coefficient (r). The r must be greater than 0.995 to be considered linear for one point calibrations.
- 8.7 Limit of Detection - The LOD for the target analytes must be determined for new instruments, after making instrument modifications which can affect the LOD and at least once per year. To make the calculations, it is necessary to perform a multipoint calibration consisting of at least four “low” concentration levels, each above the LOD, with at least five replicate determinations of the lowest concentration standard. A linear regression is performed and the standard deviation (in area counts) of the lowest concentration standard determined. The standard deviation is converted to concentration units using the slope of the linear regression:

$$s = s_a \div m$$

where m is the slope of the linear regression, s is the standard deviation (in concentration units) of the lowest concentration standard and s_a is the standard deviation (in area counts) of the lowest concentration standard.

The LOD must be calculated using the following equation [Ref. 12]:

$$\text{LOD} = t * s$$

where s is the standard deviation (in concentration units) of at least five replicate determinations of the lowest concentration standard and t is the t-factor for 99 percent confidence for a one-sided normal (Gaussian) distribution. The number

of degrees of freedom is equal to the number of replicates, minus one. An abbreviated t-table is:

| <u>Degrees of Freedom</u> | <u>t-value</u> |
|---------------------------|----------------|
| 4 | 3.7 |
| 5 | 3.4 |
| 6 | 3.1 |
| 7 | 3.0 |

The lowest standard must be of a concentration of one to five times the estimated LOD.

- 8.7.1 The maximum allowable LOD is 0.0075 µg/mL. The calculated laboratory LOD must be equal to or lower than the maximum allowable LOD. All peaks identified as target compounds that are equal to or exceed the maximum allowable LOD must be reported. If the calculated laboratory LOD is less than the maximum allowable LOD, the laboratory may choose to set its reporting limit at the maximum allowable LOD, the calculated laboratory LOD, or any level in between.
- 8.7.2 For the purpose of calculating the total mass of all species, the concentrations of the compounds below the LOD are considered to be zero.

Part G
DETERMINATION OF NMOG MASS EMISSIONS

1. INTRODUCTION

- 1.1 NMOG mass emissions consist of non-methane hydrocarbons and oxygenated hydrocarbons.
- 1.2 All definitions and abbreviations are set forth in Appendix 2 of these test procedures.

2. NMOG WEIGHTED MASS EMISSIONS

- 2.1 NMOG weighted mass (wm) emissions shall be calculated as follows:

$$\text{NMOG}_{\text{wm}} = \sum \text{NMHC}_{\text{wm}} + \sum \text{ROH}_{\text{wm}} + \sum \text{RHO}_{\text{wm}}$$

Non-methane hydrocarbon weighted mass emissions (NMHC_{wm}) can be determined by either FID or GC. If the FID method is used to calculate NMHC_{wm}, refer to Part B of these test procedures entitled, "Determination of Non-Methane Hydrocarbon Mass Emissions by Flame Ionization Detection." If the GC method is used to calculate NMHC_{wm}, refer to section 3, "Speciated Hydrocarbon Mass Emissions Calculation" contained herein. Carbonyl weighted mass emissions (RHO_{wm}) are to be calculated according to section 5 "Carbonyl Mass Emissions Calculation" contained herein. For alcohol fueled vehicles, alcohol weighted mass emissions (ROH_{wm}) are to be calculated according to section 4 "Alcohol Mass Emissions Calculation" contained herein.

3. SPECIATED HYDROCARBON MASS EMISSIONS CALCULATION

3.1 INTRODUCTION

Vehicular exhaust emissions are measured according to the FTP [Ref. 1]. For each of the three phases of the FTP, a tedlar bag of 0.5 ft³ capacity is used to collect a dilute exhaust sample. A fourth 0.5 ft³ tedlar bag is used to collect a composite dilution air (background) sample from all three phases of the FTP. All bag samples are analyzed according to Method No. 1002 (Part D of these test procedures) and Method No. 1003 (Part E of these test procedures) to determine the dilute exhaust and dilution air concentrations of individual hydrocarbon compounds. The measured hydrocarbon compound concentrations are used in the following equations to calculate the weighted mass emissions of each hydrocarbon compound.

3.2 HC MASS EMISSIONS CALCULATION PER TEST PHASE

3.2.1 $HC_{mass\ n} = (HC_{conc} * HC_{dens} * VMIX * 10^{-6}) / (\text{Carbon No.})$

3.2.2 $HC_{conc} = HC_e - (HC_d * (1 - (1 / DF)))$

NOTE: If HC_{conc} is calculated to be less than zero, then $HC_{conc} = 0$.

3.3. WEIGHTED HC MASS EMISSIONS CALCULATION

3.3.1 $HC_{wm} = 0.43 * \left(\frac{HC_{mass1} + HC_{mass2}}{D_{phase1} + D_{phase2}} \right) + 0.57 * \left(\frac{HC_{mass3} + HC_{mass2}}{D_{phase3} + D_{phase2}} \right)$

3.4. SAMPLE CALCULATION

3.4.1 Exhaust emissions from a gasoline vehicle are collected in three dilute exhaust sample bags and one dilution air (background) sample bag during the FTP. Gas chromatography is used to determine the benzene concentration of each bag sample. Calculate the weighted benzene mass emissions based on the following data:

| Test Phase | HC _e (ppbC) | HC _d (ppbC) | FID THC _e (ppmC) | CH _{4e} (ppmC) | CO _{2e} (%) | CO _{em} (ppm) | R _a (%) | VMIX (ft ³) | D _{phase n} (mile) |
|------------|------------------------|------------------------|-----------------------------|-------------------------|----------------------|------------------------|--------------------|-------------------------|-----------------------------|
| 1 | 500 | 25 | 98 | 6 | 1.20 | 280 | 28 | 2846 | 3.584 |
| 2 | 100 | 25 | 22 | 4 | 0.95 | 87 | 25 | 4854 | 3.842 |
| 3 | 120 | 25 | 29 | 5 | 1.07 | 101 | 24 | 2840 | 3.586 |

For Phase 1:

DF = $13.47 / [CO_{2e} + (NMHC_e + CH_{4e} + CO_e) * 10^{-4}]$
(see section 6, DF Calc.)

NMHC_e = FID THC_e - (r_{CH₄} * CH_{4e})
 = 98 ppmC - (1.04 * 6 ppmC)
 = 92 ppmC

CO = $(1 - (0.01 + 0.005 * HCR)) * CO_{2e} - 0.000323 * R_a) * CO_{em}$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e.

= $(1 - (0.01925) * 1.2\% - 0.000323 * 28\%) * 280\text{ ppm}$

$$\begin{aligned}
 &= 271 \text{ ppm} \\
 \text{DF} &= 13.47 / [1.2\% + (92 \text{ ppmC} + 6 \text{ ppmC} + 271 \text{ ppm}) * 10^{-4}] \\
 &= 10.89
 \end{aligned}$$

$$\begin{aligned}
 \text{HC}_{\text{conc}} &= \text{HC}_e - (\text{HC}_d * (1 - (1 / \text{DF}))) \\
 &= 500 \text{ ppbC} - (25 \text{ ppbC} * (1 - (1 / 10.89))) \\
 &= 477 \text{ ppbC}
 \end{aligned}$$

$$\begin{aligned}
 \text{Mol. Wt. of C}_6\text{H}_6 &= (6 * 12.01115) + (6 * 1.00797) \\
 &= 78.11472 \text{ g/mole}
 \end{aligned}$$

$$\begin{aligned}
 \text{HC}_{\text{dens}} &= (\text{Mol. Wt.} * \text{conversion of liter to ft}^3) / (\text{Mol. Vol.}) \\
 &= (78.11472 \text{ g/mole} * 28.316 \text{ liter/ft}^3) / 24.055 \text{ liter/mole} \\
 &= 91.952 \text{ g/ft}^3
 \end{aligned}$$

$$\text{HC}_{\text{mass n}} = (\text{HC}_{\text{conc}} * \text{HC}_{\text{dens}} * \text{VMIX} * 10^{-6}) / (\text{Carbon No.})$$

$$\begin{aligned}
 \text{HC}_{\text{mass 1}} &= (477 \text{ ppbC} * 91.952 \text{ g/ft}^3 * 2846 \text{ ft}^3 * 10^{-6}) / 6 \\
 &= 20.8 \text{ mg}
 \end{aligned}$$

$$\text{Similarly, for Phase 2:} \quad \text{HC}_{\text{mass}} = 5.7 \text{ mg}$$

$$\text{and for Phase 3:} \quad \text{HC}_{\text{mass}} = 4.2 \text{ mg}$$

Therefore,

$$\text{HC}_{\text{wm}} = 0.43 * \left(\frac{\text{HC}_{\text{mass1}} + \text{HC}_{\text{mass2}}}{\text{D}_{\text{phase1}} + \text{D}_{\text{phase2}}} \right) + 0.57 * \left(\frac{\text{HC}_{\text{mass3}} + \text{HC}_{\text{mass2}}}{\text{D}_{\text{phase3}} + \text{D}_{\text{phase2}}} \right)$$

$$\text{HC}_{\text{wm}} = 0.43 * \left(\frac{20.8 \text{ mg} + 5.7 \text{ mg}}{3.584 \text{ miles} + 3.842 \text{ miles}} \right) + 0.57 * \left(\frac{4.2 \text{ mg} + 5.7 \text{ mg}}{3.586 \text{ miles} + 3.842 \text{ miles}} \right)$$

$$\text{HC}_{\text{wm}} = 2.3 \text{ mg/mile (benzene weighted mass emissions)}$$

4. ALCOHOL MASS EMISSIONS CALCULATION

4.1. INTRODUCTION

Vehicular emissions are measured according to the (FTP [Ref. 1]). For each of the three phases of the FTP, a set of two impingers is used to collect alcohol emissions in the dilute exhaust. A fourth set of two impingers is used to collect a composite dilution air (background) alcohol sample from all three phases of the FTP. All impingers are analyzed according to Method No. 1001 to determine the alcohol concentration in each impinger. The measured alcohol concentrations are used in the following equations to calculate the weighted mass emissions of alcohol compounds.

4.2. ALCOHOL MASS EMISSIONS CALCULATION PER TEST PHASE

$$4.2.1 \quad ROH_{\text{mass } n} = (ROH_{\text{conc}} * ROH_{\text{dens}} * VMIX * 10^{-6}) / (\text{Carbon No.})$$

$$4.2.2 \quad ROH_{\text{conc}} = ROH_e - (ROH_d * (1 - (1 / DF)))$$

NOTE: If ROH_{conc} is calculated to be less than zero, then $ROH_{\text{conc}} = 0$.

$$4.2.3 \quad ROH_e = (Imass_e / Ivol_e) * (\text{Mol. Vol.} / \text{Mol. Wt.})$$

$$4.2.4 \quad Imass_e = (Iconc_{e1} + Iconc_{e2}) * Dens_{ROH} * Ivol_r$$

$$4.2.5 \quad Ivol_e = Ivol_{em} * (293.16^\circ\text{K} / Itemp_e) * (P_B / 760 \text{ mm Hg})$$

$$4.2.6 \quad ROH_d = (Imass_d / Ivol_d) * (\text{Mol. Vol.} / \text{Mol. Wt.})$$

$$4.2.7 \quad Imass_d = (Iconc_{d1} + Iconc_{d2}) * Dens_{ROH} * Ivol_r$$

$$4.2.8 \quad Ivol_d = Ivol_{dm} * (293.16^\circ\text{K} / Itemp_d) * (P_B / 760 \text{ mm Hg})$$

4.3. WEIGHTED ALCOHOL MASS EMISSIONS CALCULATION

$$ROH_{\text{wm}} = 0.43 * \left(\frac{ROH_{\text{mass1}} + ROH_{\text{mass2}}}{D_{\text{phase1}} + D_{\text{phase2}}} \right) + 0.57 * \left(\frac{ROH_{\text{mass3}} + ROH_{\text{mass2}}}{D_{\text{phase3}} + D_{\text{phase2}}} \right)$$

4.4 SAMPLE CALCULATION

4.4.1 Alcohol emissions from an M85 fueled vehicle are collected in three sets of dilute exhaust impingers and one set of dilution air impingers during the FTP. Gas chromatography is used to determine the methanol concentration in each impinger. Calculate the weighted methanol mass emissions based on the following data:

| Test Phase | Ivol _r (mL) | Iconc _{e1} (µg/mL) | Iconc _{e2} (µg/mL) | Ivol _{em} (liter) | Iconc _{d1} (µg/mL) | Iconc _{d2} (µg/mL) | Ivol _{dm} (liter) | Itemp _e (°K) | Itemp _d (°K) |
|------------|------------------------|-----------------------------|-----------------------------|----------------------------|-----------------------------|-----------------------------|----------------------------|-------------------------|-------------------------|
| 1 | 15 | 2.24 | 0.05 | 3.90 | 0.07 | 0.01 | 13.50 | 295 | 294 |
| 2 | 15 | 0.29 | 0.06 | 6.50 | 0.07 | 0.01 | 13.50 | 297 | 294 |
| 3 | 15 | 0.32 | 0.02 | 4.00 | 0.07 | 0.01 | 13.50 | 298 | 294 |

| Test Phase | FID THC _e (ppmC) | CH _{4e} (ppmC) | CO _{2e} (%) | CO _{em} (ppm) | R _a (%) | VMIX (ft ³) | D _{phase n} (mile) | P _B (mmHg) | HCHO _e (ppm) |
|------------|-----------------------------|-------------------------|----------------------|------------------------|--------------------|-------------------------|-----------------------------|-----------------------|-------------------------|
| 1 | 82 | 9 | 1.5 | 250 | 30 | 2834 | 3.581 | 760 | 0.81 |
| 2 | 19 | 5 | 0.7 | 20 | 32 | 4862 | 3.845 | 760 | 0.09 |
| 3 | 22 | 6 | 0.8 | 32 | 29 | 2835 | 3.583 | 760 | 0.10 |

For Phase 1:

$$\begin{aligned}
 \text{Imass}_e &= (\text{Iconc}_{e1} + \text{Iconc}_{e2}) * \text{Dens}_{\text{ROH}} * \text{Ivol}_r \\
 &= (2.24 \mu\text{g/mL} + 0.05 \mu\text{g/mL}) * 0.7914 \text{ g/mL} * \\
 &\quad 15 \text{ mL} * 10^6 \mu\text{g/g} \\
 &= 27.2 \mu\text{g}
 \end{aligned}$$

$$\begin{aligned}
 \text{Mol. Wt. of CH}_3\text{OH} &= (1 * 12.01115) + (4 * 1.00797) + (1 * 15.9994) \\
 &= 32.0428 \text{ g/mole}
 \end{aligned}$$

$$\begin{aligned}
 \text{Ivol}_e &= \text{Ivol}_{em} * (293/16^\circ \text{ K} / \text{Itemp}_e) * (\text{P}_B / 760 \text{ mm Hg}) \\
 &= 3.90 \text{ liter} * (293.16^\circ \text{ K} / 295^\circ \text{ K}) * (760 \text{ mm Hg} / 760 \text{ mm Hg}) \\
 &= 3.88 \text{ liter}
 \end{aligned}$$

$$\begin{aligned}
 \text{ROH}_e &= (\text{Imass}_e / \text{Ivol}_e) * (\text{Mol. Vol.} / \text{Mol. Wt.}) \\
 &= (27.2 * 10^{-6} \text{ g} / 3.88 \text{ liter}) * (24.055 \text{ liter/mole} / 32.0428 \text{ g/mole}) \\
 &= 5.27 \text{ ppmC}
 \end{aligned}$$

$$\text{Imass}_d = (\text{Iconc}_{d1} + \text{Iconc}_{d2}) * \text{Dens}_{\text{ROH}} * \text{Ivol}_r$$

$$= (0.07 \mu\text{g/mL} + 0.01 \mu\text{g/mL}) * 0.7914 \text{ g/ml} * 15 \text{ ml} * 10^6 \mu\text{g/g}$$

$$= 0.95 \mu\text{g}$$

$$\text{Ivol}_d = \text{Ivol}_{dm} * (293.16^\circ \text{K} / \text{Itemp}_d) * (P_B / 760 \text{ mm Hg})$$

$$= 13.50 \text{ liter} * (293.16^\circ \text{K} / 294^\circ \text{K}) * (760 \text{ mm Hg} / 760 \text{ mm Hg})$$

$$= 13.46 \text{ liter}$$

$$\text{ROH}_d = (\text{Imass}_d / \text{Ivol}_d) * (\text{Mol. Vol.} / \text{Mol. Wt.})$$

$$= (0.95 * 10^{-6} \text{ g} / 13.46 \text{ liter}) * (24.055 \text{ liter/mole} / 32.0428 \text{ g/mole})$$

$$= 0.05 \text{ ppmC}$$

$$DF = \frac{12.02}{\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e + \text{ROH}_e + \text{HCHO}_e) * 10^{-4}}$$

(see section 6, DF Calc.)

$$\text{NMHC}_e = \text{FID THC}_e - (r_{\text{CH}_4} * \text{CH}_{4e}) - (r_{\text{CH}_3\text{OH}} * \text{ROH}_e)$$

$$= 82 \text{ ppmC} - (1.04 * 9 \text{ ppmC}) - (0.66 * 5.27 \text{ ppmC})$$

$$= 69 \text{ ppmC}$$

$$\text{CO}_e = (1 - (0.01 + 0.005 * \text{HCR})) * \text{CO}_{2e} - 0.000323 * R_a * \text{CO}_{em}$$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e.

$$= (1 - (0.02705) * 1.5\% - 0.000323 * 30\%) * 250 \text{ ppm}$$

$$= 237 \text{ ppm}$$

$$DF = \frac{12.02}{1.5\% + (69 \text{ ppmC} + 9 \text{ ppmC} + 237 \text{ ppmC} + 5.27 \text{ ppmC} + 0.81 \text{ ppm}) * 10^{-4}}$$

$$\text{ROH}_{\text{conc}} = \text{ROH}_e - (\text{ROH}_d * (1 - (1 / DF)))$$

$$= 5.27 \text{ ppmC} - (0.05 \text{ ppmC} * (1 - (1 / 7.84)))$$

$$= 5.23 \text{ ppmC}$$

$$\text{ROH}_{\text{dens}} = (\text{Mol. Wt.} * \text{conversion of liter to ft}^3) / (\text{Mol. Vol.})$$

$$= (32.0428 \text{ g/mole} * 28.316 \text{ liter/ft}^3) / 24.055 \text{ liter/mole}$$

$$= 37.719 \text{ g/ft}^3$$

$$\text{ROH}_{\text{mass n}} = (\text{ROH}_{\text{conc}} * \text{ROH}_{\text{dens}} * \text{VMIX} * 10^{-6}) / (\text{Carbon No.})$$

$$\text{ROH}_{\text{mass l}} = (5.23 \text{ ppmC} * 37.719 \text{ g/ft}^3 * 2834 \text{ ft}^3 * 10^{-6}) / 1$$

$$= 0.56 \text{ g}$$

Similarly, for Phase 2:
and for Phase 3:

$$\begin{aligned} \text{ROH}_{\text{mass } 2} &= 0.08 \text{ g} \\ \text{ROH}_{\text{mass } 3} &= 0.08 \text{ g} \end{aligned}$$

Therefore,

$$\text{ROH}_{\text{wm}} = 0.43 * \left(\frac{\text{ROH}_{\text{mass1}} + \text{ROH}_{\text{mass2}}}{D_{\text{phase1}} + D_{\text{phase2}}} \right) + 0.57 * \left(\frac{\text{ROH}_{\text{mass3}} + \text{ROH}_{\text{mass2}}}{D_{\text{phase3}} + D_{\text{phase2}}} \right)$$

$$\text{ROH}_{\text{wm}} = 0.43 * \left(\frac{0.56 \text{ mg} + 0.08 \text{ mg}}{3.581 \text{ miles} + 3.845 \text{ miles}} \right) + 0.57 * \left(\frac{0.08 \text{ mg} + 0.08 \text{ mg}}{3.583 \text{ miles} + 3.845 \text{ miles}} \right)$$

$$\text{ROH}_{\text{wm}} = 0.05 \text{ g (methanol weighted mass emissions)}$$

5. CARBONYL MASS EMISSIONS CALCULATIONS

5.1. INTRODUCTION

Vehicular emissions are measured according to the FTP [Ref. 1]. For each of the three phases of the FTP, a set of two impingers (or cartridges) is used to collect carbonyl emissions in the dilute exhaust. A fourth set of two impingers (or cartridges) is used to collect a composite dilution air (background) carbonyl sample from all three phases of the FTP. All impingers (or cartridges) are analyzed according to Method No. 1004 to determine the mass of individual carbonyl compounds in each impinger (or cartridge). The measured carbonyl masses are used in the following equations to calculate the weighted mass emissions of each carbonyl compound.

5.2. CARBONYL MASS EMISSIONS CALCULATION PER TEST PHASE

$$5.2.1 \quad \text{RHO}_{\text{mass } n} = (\text{RHO}_{\text{conc}} * \text{RHO}_{\text{dens}} * \text{VMIX} * 10^{-6})$$

$$5.2.2 \quad \text{RHO}_{\text{conc}} = \text{RHO}_e - (\text{RHO}_d * (1 - (1 / \text{DF})))$$

NOTE: If RHO_{conc} is calculated to be less than zero, then $\text{RHO}_{\text{conc}} = 0$.

$$5.2.3 \quad \text{RHO}_e = (\text{Imass}_e / \text{Ivol}_e) * (\text{Mol. Vol.} / \text{Mol. Wt.})$$

$$5.2.4 \quad \text{Ivol}_e = \text{Ivol}_{\text{em}} * (293.16^\circ\text{K} / \text{Itemp}_e) * (P_B / 760 \text{ mm Hg})$$

$$5.2.5 \quad \text{RHO}_d = (\text{Imass}_d / \text{Ivol}_d) * (\text{Mol. Vol.} / \text{Mol. Wt.})$$

$$5.2.6 \quad \text{Ivol}_d = \text{Ivol}_{\text{dm}} * (293.16^\circ\text{K} / \text{Itemp}_d) * (P_B / 760 \text{ mm Hg})$$

5.3. WEIGHTED CARBONYL MASS EMISSIONS CALCULATION

$$RHO_{wm} = 0.43 * \left(\frac{RHO_{mass1} + RHO_{mass2}}{D_{phase1} + D_{phase2}} \right) + 0.57 * \left(\frac{RHO_{mass3} + RHO_{mass2}}{D_{phase3} + D_{phase2}} \right)$$

5.4. SAMPLE CALCULATION

5.4.1 Carbonyl emissions from a CNG vehicle are collected in three sets of dilute exhaust impingers and one set of dilution air impingers during the FTP. HPLC is used to determine the formaldehyde mass in each impinger. Calculate the weighted formaldehyde mass emissions based on the following data:

| Test Phase | Ivol _r (mL) | Imass _e (µg) | Ivol _{em} (liter) | Imass _d (µg) | Ivol _{dm} (liter) | Itemp _e (°K) | Itemp _d (°K) |
|------------|------------------------|-------------------------|----------------------------|-------------------------|----------------------------|-------------------------|-------------------------|
| 1 | 15 | 2.45 | 8.49 | 0.17 | 31.57 | 295 | 292 |
| 2 | 15 | 0.76 | 14.55 | 0.17 | 31.57 | 298 | 292 |
| 3 | 15 | 0.64 | 4.00 | 0.17 | 31.57 | 298 | 292 |

| Test Phase | FID THC _e (ppmC) | CH _{4e} (ppmC) | CO _{2e} (%) | CO _{em} (ppm) | R _a (%) | VMIX (ft ³) | D _{phase n} (mile) | P _B (mmHg) |
|------------|-----------------------------|-------------------------|----------------------|------------------------|--------------------|-------------------------|-----------------------------|-----------------------|
| 1 | 132 | 108 | 0.9 | 8 | 68 | 2866 | 3.581 | 760 |
| 2 | 4 | 3 | 0.1 | 4 | 67 | 4841 | 3.845 | 760 |
| 3 | 22 | 9 | 0.5 | 5 | 65 | 2837 | 3.583 | 760 |

For Phase 1:

$$\begin{aligned} \text{Mol. Wt. of HCHO} &= (1 * 12.01115) + (2 * 1.00797) + (1 * 15.9994) \\ &= 30.0268 \text{ g/mole} \end{aligned}$$

$$\begin{aligned} Ivol_e &= Ivol_{em} * (293/16^\circ \text{K} / Itemp_e) * (P_B / 760 \text{ mm Hg}) \\ &= 8.49 \text{ liter} * (293.16^\circ \text{K} / 295^\circ \text{K}) * (760 \text{ mm Hg} / 760 \text{ mm Hg}) \\ &= 8.44 \text{ liter} \end{aligned}$$

$$\begin{aligned} RHO_e &= (Imass_e / Ivol_e) * (\text{Mol. Vol.} / \text{Mol. Wt.}) \\ &= (2.45 * 10^{-6} \text{ g} / 8.44 \text{ liter}) * (24.055 \text{ liter/mole} / 30.0268 \text{ g/mole}) \\ &= 233 \text{ ppb} \end{aligned}$$

$$\begin{aligned} \text{Ivol}_d &= \text{Ivol}_{dm} * (293.16^\circ\text{K} / \text{Itemp}_d) * (P_B / 760 \text{ mm Hg}) \\ &= 31.57 \text{ liter} * (293.16^\circ\text{K} / 292^\circ\text{K}) * (760 \text{ mm Hg} / 760 \text{ mm Hg}) \\ &= 31.70 \text{ liter} \end{aligned}$$

$$\begin{aligned} \text{RHO}_d &= (\text{Imass}_d / \text{Ivol}_d) * (\text{Mol. Vol.} / \text{Mol. Wt.}) \\ &= (0.17 * 10^{-6} \text{ g} / 31.70 \text{ liter}) * (24.055 \text{ liter/mole} / 30.0268 \text{ g/mole}) \\ &= 4 \text{ ppb} \end{aligned}$$

$$\text{DF} = 9.77 / [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}^e) * 10^{-4}]$$

(see section 6, DF Calc.)

$$\begin{aligned} \text{NMHC}_e &= \text{FID THC}_e - (r_{\text{CH}_4} * \text{CH}_{4e}) \\ &= 132 \text{ ppmC} - (1.04 * 108 \text{ ppmC}) \\ &= 20 \text{ ppmC} \end{aligned}$$

$$\text{CO}_e = (1 - (0.01 + 0.005 * \text{HCR}) * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e.

$$\begin{aligned} &= (1 - 0.02890 * 0.9\% - 0.000323 * 68\%) * 8 \text{ ppm} \\ &= 7.6 \text{ ppm} \end{aligned}$$

$$\begin{aligned} \text{DF} &= 9.77 / [0.9\% + (20 \text{ ppmC} + 108 \text{ ppmC} + 7.6 \text{ ppm}) * 10^{-4}] \\ &= 10.69 \end{aligned}$$

$$\begin{aligned} \text{RHO}_{\text{conc}} &= \text{RHO}_e - (\text{RHO}_d * (1 - (1 / \text{DF}))) \\ &= 233 \text{ ppb} - (4 \text{ ppb} * (1 - (1 / 10.69))) \\ &= 229 \text{ ppb} \end{aligned}$$

$$\begin{aligned} \text{RHO}_{\text{dens}} &= (\text{Mol. Wt.} * \text{conversion of liter to ft}^3) / (\text{Mol. Vol.}) \\ &= (30.0268 \text{ g/mole} * 28.316 \text{ liter/ft}^3) / 24.055 \text{ liter/mole} \\ &= 35.35 \text{ g/ft}^3 \end{aligned}$$

$$\begin{aligned} \text{RHO}_{\text{mass } n} &= (\text{RHO}_{\text{conc}} * \text{RHO}_{\text{dens}} * \text{VMIX} * 10^{-6}) \\ \text{RHO}_{\text{mass } 1} &= (229 \text{ ppb} * 35.35 \text{ g/ft}^3 * 2866 \text{ ft}^3 * 10^{-6}) \\ &= 23.2 \text{ mg} \end{aligned}$$

Similarly, for Phase 2: $\text{RHO}_{\text{mass } 2} = 6.6 \text{ mg}$
 and for Phase 3: $\text{RHO}_{\text{mass } 3} = 12.7 \text{ mg}$

Therefore,

$$RHO_{wm} = 0.43 * \left(\frac{RHO_{mass1} + RHO_{mass2}}{D_{phase1} + D_{phase2}} \right) + 0.57 * \left(\frac{RHO_{mass3} + RHO_{mass2}}{D_{phase3} + D_{phase2}} \right)$$

$$RHO_{wm} = 0.43 * \left(\frac{23.2 \text{ mg} + 6.6 \text{ mg}}{3.581 \text{ miles} + 3.845 \text{ miles}} \right) + 0.57 * \left(\frac{12.7 \text{ mg} + 6.6 \text{ mg}}{3.583 \text{ miles} + 3.845 \text{ miles}} \right)$$

$$RHO_{wm} = 3.2 \text{ mg/mi (formaldehyde weighted mass emissions)}$$

5.4.2 Carbonyl emissions from a gasoline vehicle are collected in three sets of dilute exhaust cartridges and dilution air cartridges during the FTP. HPLC is used to determine the formaldehyde mass in each cartridge. Calculate the weighted formaldehyde mass emissions based on the following data:

| Test Phase | Iconc _{ce} (µg/mL) | Iconc _{cd} (µg/mL) | Iconc _{blk} (µg/mL) | Ivol _{dm} (liter) | Ivol _{em} (liter) | Itemp _e (°K) | Itemp _d (°K) |
|------------|-----------------------------|-----------------------------|------------------------------|----------------------------|----------------------------|-------------------------|-------------------------|
| 1 | 1.212 | 0.028 | 0.0 | 8.61 | 8.57 | 294.26 | 294.26 |
| 2 | 0.334 | 0.043 | 0.0 | 14.91 | 13.83 | 294.26 | 294.26 |
| 3 | 0.172 | 0.026 | 0.0 | 8.68 | 8.74 | 294.26 | 294.26 |

| Test Phase | FID THC _e (ppmC) | CH _{4e} (ppmC) | CO _{2e} (%) | CO _{em} (ppm) | R _a (%) | VMIX (ft ³) | D _{phase n} (mile) | P _B (mmHg) |
|------------|-----------------------------|-------------------------|----------------------|------------------------|--------------------|-------------------------|-----------------------------|-----------------------|
| 1 | 132 | 108 | 0.9 | 8 | 68 | 2866 | 3.581 | 760 |
| 2 | 4 | 3 | 0.1 | 4 | 67 | 4841 | 3.845 | 760 |
| 3 | 22 | 9 | 0.5 | 5 | 65 | 2837 | 3.583 | 760 |

For all three phases Ivol_c = 4.4 mL

For Phase 1:

$$Imass_d = (Iconc_{cd} - Iconc_{blk}) * Ivol_c$$

$$Imass_d = (0.028 - 0.0) \mu\text{g/mL} * 4.4 \text{ mL} = 0.1232 \mu\text{g}$$

$$Imass_e = (Iconc_{ce} - Iconc_{blk}) * Ivol_c$$

$$\text{I}_{\text{mass}_e} = (1.212 - 0.0) \mu\text{g/mL} * 4.4 \text{ mL} = 5.33 \mu\text{g}$$

$$\begin{aligned} \text{Mol. Wt. of HCHO} &= (1 * 12.01115) + (2 * 1.00797) + (1 * 15.9994) \\ &= 30.0265 \text{ g/mole} \end{aligned}$$

$$\begin{aligned} \text{I}_{\text{vol}_e} &= \text{I}_{\text{vol}_{em}} * (293/16^\circ \text{K} / \text{I}_{\text{temp}_e}) * (\text{P}_B / 760 \text{ mm Hg}) \\ &= 8.57 \text{ liter} * (293.16^\circ\text{K} / 294.26^\circ\text{K}) * (760 \text{ mm Hg} / 760 \text{ mm Hg}) \\ &= 8.54 \text{ liter} \end{aligned}$$

$$\begin{aligned} \text{RHO}_e &= (\text{I}_{\text{mass}_e} / \text{I}_{\text{vol}_e}) * (\text{Mol. Vol.} / \text{Mol. Wt.}) \\ &= (5.33 * 10^{-6} \text{ g} / 8.54 \text{ liter}) * (24.055 \text{ liter/mole} / 30.0265 \text{ g/mole}) \\ &= 500 \text{ ppb} \end{aligned}$$

$$\begin{aligned} \text{I}_{\text{vol}_d} &= \text{I}_{\text{vol}_{dm}} * (293.16^\circ\text{K} / \text{I}_{\text{temp}_d}) * (\text{P}_B / 760 \text{ mm Hg}) \\ &= 8.61 \text{ liter} * (293.16^\circ\text{K} / 294.26^\circ\text{K}) * (760 \text{ mm Hg} / 760 \text{ mm Hg}) \\ &= 8.58 \text{ liter} \end{aligned}$$

$$\begin{aligned} \text{RHO}_d &= (\text{I}_{\text{mass}_d} / \text{I}_{\text{vol}_d}) * (\text{Mol. Vol.} / \text{Mol. Wt.}) \\ &= (0.1232 * 10^{-6} \text{ g} / 8.58 \text{ liter}) * (24.055 \text{ liter/mole} / 30.0265 \text{ g/mole}) \\ &= 11.5 \text{ ppb} \end{aligned}$$

$$\begin{aligned} \text{DF} &= 9.77 / [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e) * 10^{-4}] \\ &(\text{see section 6, DF Calc.}) \end{aligned}$$

$$\begin{aligned} \text{NMHC}_e &= \text{FID THC}_e - (\text{r}_{\text{CH}_4} * \text{CH}_{4e}) \\ &= 132 \text{ ppmC} - (1.04 * 108 \text{ ppmC}) \\ &= 20 \text{ ppmC} \end{aligned}$$

$$\text{CO}_e = (1 - (0.01 + 0.005 * \text{HCR}) * \text{CO}_{2e} - 0.000323 * \text{R}_a) * \text{CO}_{em}$$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e.

$$\begin{aligned} &= (1 - 0.02890 * 0.9\% - 0.000323 * 68\%) * 8 \text{ ppm} \\ &= 7.6 \text{ ppm} \end{aligned}$$

$$\begin{aligned} \text{DF} &= 9.77 / [0.9\% + (20 \text{ ppmC} + 108 \text{ ppmC} + 7.6 \text{ ppm}) * 10^{-4}] \\ &= 10.69 \end{aligned}$$

$$\begin{aligned} \text{RHO}_{\text{conc}} &= \text{RHO}_e - (\text{RHO}_d * (1 - (1 / \text{DF}))) \\ &= 500 \text{ ppb} - (11.5 \text{ ppb} * (1 - (1 / 10.69))) \\ &= 489.6 \text{ ppb (formaldehyde)} \end{aligned}$$

$$\text{RHO}_{\text{dens}} = (\text{Mol. Wt.} * \text{conversion of liter to ft}^3) / (\text{Mol. Vol.})$$

$$= (30.0265 \text{ g/mole} * 28.316 \text{ liter/ft}^3) / 24.055 \text{ liter/mole}$$

$$= 35.35 \text{ g/ft}^3$$

$$\text{RHO}_{\text{mass n}} = (\text{RHO}_{\text{conc}} * \text{RHO}_{\text{dens}} * \text{VMIX} * 10^{-6})$$

$$\text{RHO}_{\text{mass 1}} = (489.6 \text{ ppb} * 35.35 \text{ g/ft}^3 * 2866 \text{ ft}^3 * 10^{-6})$$

$$= 49.6 \text{ mg (formaldehyde)}$$

Similarly, for Phase 2: $\text{RHO}_{\text{mass 2}} = 12.9 \text{ mg}$

and similarly for Phase 3: $\text{RHO}_{\text{mass 3}} = 5.6 \text{ mg}$

Therefore,

$$\text{RHO}_{\text{wm}} = 0.43 * \left(\frac{\text{RHO}_{\text{mass 1}} + \text{RHO}_{\text{mass 2}}}{\text{D}_{\text{phase 1}} + \text{D}_{\text{phase 2}}} \right) + 0.57 * \left(\frac{\text{RHO}_{\text{mass 3}} + \text{RHO}_{\text{mass 2}}}{\text{D}_{\text{phase 3}} + \text{D}_{\text{phase 2}}} \right)$$

$$\text{RHO}_{\text{wm}} = 0.43 * \left(\frac{49.6 \text{ mg} + 12.9 \text{ mg}}{3.581 \text{ miles} + 3.845 \text{ miles}} \right) + 0.57 * \left(\frac{5.6 \text{ mg} + 12.9 \text{ mg}}{3.583 \text{ miles} + 3.845 \text{ miles}} \right)$$

$$\text{RHO}_{\text{wm}} = 5.04 \text{ mg/mi (formaldehyde weighted mass emissions)}$$

6. DILUTION FACTOR CALCULATION

6.1. For Non-Alcohol Fueled Vehicles:

$$6.1.1 \quad \text{DF} = \frac{100 * \left(\frac{x}{x + y/2 + 3.76 * (x + y/4 - z/2)} \right)}{\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e) * 10^{-4}}$$

(where fuel composition is $\text{C}_x\text{H}_y\text{O}_z$ as measured for the fuel used.)

$$6.1.2 \quad \text{CO}_e = (1 - (0.01 + 0.005 * \text{HCR}) * \text{CO}_{2e} - 0.000323 * \text{R}_a) * \text{CO}_{\text{em}}$$

NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e .

a) For gasoline, $\text{CH}_{1.85}$, where $x = 1$, $y = 1.85$, and $z = 0$:

$$\text{DF} = 13.47 / [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e) * 10^{-4}]$$

$$\text{CO}_e = (1 - 0.01925 * \text{CO}_{2e} - 0.000323 * \text{R}_a) * \text{CO}_{\text{em}}$$

b) For Phase 2 gasoline, $\text{CH}_{1.94}$, where $x = 1$, $y = 1.94$ and $z = 0.017$,

$$DF = 13.29 / [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e) * 10^{-4}]$$

$$\text{CO}_e = (1 - 0.01970 * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

c) For LPG, $\text{CH}_{2.64}$, where $x = 1$, $y = 2.64$, $z = 0$:

$$DF = 11.68 / [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e) * 10^{-4}]$$

$$\text{CO}_e = (1 - 0.02320 * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

d) For CNG, $\text{CH}_{3.78}$, where $x = 1$, $y = 3.78$, and $z = 0.016$:

$$DF = 9.83 / [[\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e) * 10^{-4}]]$$

$$\text{CO}_e = (1 - 0.02890 * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

6.2. For Alcohol Fueled Vehicles:

$$6.2.1 \quad DF = \frac{100 * \left(\frac{x}{x + y/2 + 3.76 * (x + y/4 - z/2)} \right)}{\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e + \text{ROH}_e + \text{HCHO}_e) * 10^{-4}}$$

(where fuel composition is $\text{C}_x\text{H}_y\text{O}_z$ as measured for the fuel used.)

6.2.2 $\text{CO}_e = (1 - (0.01 + 0.005 * \text{HCR}) * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$
 NOTE: If a CO instrument which meets the criteria specified in CFR 40, 86.111 is used and the conditioning column has been deleted, CO_{em} must be substituted directly for CO_e .

a) For M100 (100% methanol), CH_3OH , where $x = 1$, $y = 4$, and $z = 1$:

$$DF = 11.57 / [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e + \text{ROH}_e + \text{HCHO}_e) * 10^{-4}]$$

$$\text{CO}_e = (1 - 0.03000 * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

b) For M85 (85% methanol, 15% indolene), $\text{CH}_{3.41}\text{O}_{0.72}$, where $x = 1$, $y = 3.41$, and $z = 0.72$:

$$DF = 12.02 / [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e + \text{ROH}_e + \text{HCHO}_e) * 10^{-4}]$$

$$\text{CO}_e = (1 - 0.02705 * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

c) For E100 (100% ethanol), $\text{C}_2\text{H}_5\text{OH}$, where $x = 1$, $y = 3$, and $z = 0.5$:

$$DF = 12.29 / [\text{CO}_{2e} + (\text{NMHC}_e + \text{CH}_{4e} + \text{CO}_e + \text{ROH}_e + \text{HCHO}_e) * 10^{-4}]$$

$$\text{CO}_e = (1 - 0.02500 * \text{CO}_{2e} - 0.000323 * R_a) * \text{CO}_{em}$$

APPENDIX 1

LIST OF COMPOUNDS

| CAS # | COMPOUND | MIR |
|------------|--|-------|
| | Alcohols | |
| 00067-56-1 | methanol | 0.71 |
| 00064-17-5 | ethanol | 1.69 |
| | Light End and Mid-Range Hydrocarbons (Listed in approximate elution order) | |
| 00074-85-1 | ethene | 9.08 |
| 00074-86-2 | ethyne | 1.25 |
| 00074-84-0 | ethane | 0.31 |
| 00115-07-1 | propene | 11.58 |
| 00074-98-6 | propane | 0.56 |
| 00463-49-0 | 1,2-propadiene | 12.16 |
| 00074-99-7 | 1-propyne | 6.45 |
| 00075-28-5 | 2-methylpropane | 1.35 |
| 00115-11-7 | 2-methylpropene | 6.35 |
| 00106-98-9 | 1-butene | 10.29 |
| 00106-99-0 | 1,3-butadiene | 13.58 |
| 00106-97-8 | n-butane | 1.33 |
| 00624-64-6 | trans-2-butene | 13.91 |
| 00463-82-1 | 2,2-dimethylpropane | 1.68 |
| 00107-00-6 | 1-butyne | 6.20 |
| 00590-18-1 | cis-2-butene | 13.22 |
| 00563-45-1 | 3-methyl-1-butene | 6.99 |
| 00078-78-4 | 2-methylbutane | 1.68 |
| 00503-17-3 | 2-butyne | 16.33 |
| 00109-67-1 | 1-pentene | 7.79 |
| 00563-46-2 | 2-methyl-1-butene | 6.51 |
| 00109-66-0 | n-pentane | 1.54 |
| 00078-79-5 | 2-methyl-1,3-butadiene | 10.69 |
| 00646-04-8 | trans-2-pentene | 10.23 |
| 00558-37-2 | 3,3-dimethyl-1-butene | 6.06 |
| 00627-20-3 | cis-2-pentene | 10.24 |
| 00689-97-4 | 1-buten-3-yne | 11.09 |
| 00513-35-9 | 2-methyl-2-butene | 14.45 |
| 00542-92-7 | 1,3-cyclopentadiene | 7.61 |
| 00075-83-2 | 2,2-dimethylbutane | 1.33 |
| 00142-29-0 | cyclopentene | 7.38 |
| 00691-37-2 | 4-methyl-1-pentene | 6.26 |

| CAS # | COMPOUND | MIR |
|------------|--------------------------------|-------|
| 00760-20-3 | 3-methyl-1-pentene | 6.22 |
| 00287-92-3 | cyclopentane | 2.69 |
| 00079-29-8 | 2,3-dimethylbutane | 1.14 |
| 01634-04-4 | 1-methyl-tert-butyl-ether | 0.78 |
| 00691-38-3 | 4-methyl-cis-2-pentene | 8.44 |
| 00107-83-5 | 2-methylpentane | 1.80 |
| 00674-76-0 | 4-methyl-trans-2-pentene | 8.44 |
| 00096-14-0 | 3-methylpentane | 2.07 |
| 00763-29-1 | 2-methyl-1-pentene | 5.18 |
| 00592-41-6 | 1-hexene | 6.17 |
| 00110-54-3 | n-hexane | 1.45 |
| 13269-52-8 | trans-3-hexene | 8.16 |
| 07642-09-3 | cis-3-hexene | 8.22 |
| 04050-45-7 | trans-2-hexene | 8.44 |
| 00616-12-6 | 3-methyl-trans-2-pentene | 8.44 |
| 00625-27-4 | 2-methyl-2-pentene | 12.28 |
| 01120-62-3 | 3-methylcyclopentene | 8.65 |
| 07688-21-3 | cis-2-hexene | 8.44 |
| 00637-92-3 | 1-ethyl-tert-butyl-ether | 2.11 |
| 00922-62-3 | 3-methyl-cis-2-pentene | 8.44 |
| 00590-35-2 | 2,2-dimethylpentane | 1.22 |
| 00096-37-7 | methylcyclopentane | 2.42 |
| 00108-08-7 | 2,4-dimethylpentane | 1.65 |
| 00464-06-2 | 2,2,3-trimethylbutane | 1.32 |
| 07385-78-6 | 3,4-dimethyl-1-pentene | 4.56 |
| 00693-89-0 | 1-methylcyclopentene | 13.95 |
| 00071-43-2 | benzene | 0.81 |
| 03404-61-3 | 3-methyl-1-hexene | 4.56 |
| 00562-49-2 | 3,3-dimethylpentane | 1.32 |
| 00110-82-7 | cyclohexane | 1.46 |
| 00591-76-4 | 2-methylhexane | 1.37 |
| 00565-59-3 | 2,3-dimethylpentane | 1.55 |
| 00110-83-8 | cyclohexene | 5.45 |
| 00589-34-4 | 3-methylhexane | 1.86 |
| 01759-58-6 | trans-1,3-dimethylcyclopentane | 2.15 |
| 02532-58-3 | cis-1,3-dimethylcyclopentane | 2.15 |
| 00617-78-7 | 3-ethylpentane | 1.63 |
| 00822-50-4 | trans-1,2-dimethylcyclopentane | 1.99 |
| 00592-76-7 | 1-heptene | 4.56 |
| 00540-84-1 | 2,2,4-trimethylpentane | 1.44 |
| 14686-14-7 | trans-3-heptene | 6.96 |
| 00142-82-5 | n-heptane | 1.28 |
| 02738-19-4 | 2-methyl-2-hexene | 6.96 |
| 03899-36-3 | 3-methyl-trans-3-hexene | 6.96 |

| CAS # | COMPOUND | MIR |
|------------|--|------|
| 14686-13-6 | trans-2-heptene | 7.33 |
| 00816-79-5 | 3-ethyl-2-pentene | 6.96 |
| 00107-39-1 | 2,4,4-trimethyl-1-pentene | 3.45 |
| 10574-37-5 | 2,3-dimethyl-2-pentene | 6.96 |
| 06443-92-1 | cis-2-heptene | 6.96 |
| 00108-87-2 | methylcyclohexane | 1.99 |
| 00590-73-8 | 2,2-dimethylhexane | 1.13 |
| 00107-40-4 | 2,4,4-trimethyl-2-pentene | 5.85 |
| 01640-89-7 | ethylcyclopentane | 2.27 |
| 00592-13-2 | 2,5-dimethylhexane | 1.68 |
| 00589-43-5 | 2,4-dimethylhexane | 1.80 |
| 02815-58-9 | 1,2,4-trimethylcyclopentane | 1.75 |
| 00563-16-6 | 3,3-dimethylhexane | 1.57 |
| 00565-75-3 | 2,3,4-trimethylpentane | 1.23 |
| 00560-21-4 | 2,3,3-trimethylpentane | 1.57 |
| 00108-88-3 | toluene | 3.97 |
| 00584-94-1 | 2,3-dimethylhexane | 1.34 |
| 00592-27-8 | 2-methylheptane | 1.20 |
| 00589-53-7 | 4-methylheptane | 1.48 |
| 00589-81-1 | 3-methylheptane | 1.35 |
| 15890-40-1 | (1a,2a,3b)-1,2,3-trimethylcyclopentane | 1.75 |
| 00638-04-0 | cis-1,3-dimethylcyclohexane | 1.72 |
| 02207-04-7 | trans-1,4-dimethylcyclohexane | 1.75 |
| 03522-94-9 | 2,2,5-trimethylhexane | 1.33 |
| 02613-65-2 | trans-1-methyl-3-ethylcyclopentane | 1.75 |
| 16747-50-5 | cis-1-methyl-3-ethylcyclopentane | 1.75 |
| 00111-66-0 | 1-octene | 3.45 |
| 14850-23-8 | trans-4-octene | 5.90 |
| 00111-65-9 | n-octane | 1.11 |
| 13389-42-9 | trans-2-octene | 5.90 |
| 02207-03-6 | trans-1,3-dimethylcyclohexane | 1.72 |
| 07642-04-8 | cis-2-octene | 5.90 |
| 01069-53-0 | 2,3,5-trimethylhexane | 1.33 |
| 02213-23-2 | 2,4-dimethylheptane | 1.48 |
| 02207-01-4 | cis-1,2-dimethylcyclohexane | 1.75 |
| 01072-05-5 | 2,6-dimethylheptane | 1.25 |
| 01678-91-7 | ethylcyclohexane | 1.75 |
| 00926-82-9 | 3,5-dimethylheptane | 1.63 |
| 00100-41-4 | ethylbenzene | 2.79 |
| 03074-71-3 | 2,3-dimethylheptane | 1.25 |
| 00108-38-3 | m-&p-xylene | 8.49 |
| 02216-34-4 | 4-methyloctane | 1.08 |
| 03221-61-2 | 2-methyloctane | 0.96 |
| 02216-33-3 | 3-methyloctane | 1.25 |

| CAS # | COMPOUND | MIR |
|------------|---------------------------------------|-------|
| 00100-42-5 | styrene (ethenylbenzene) | 1.95 |
| 00095-47-6 | o-xylene | 7.49 |
| 00124-11-8 | 1-nonene | 2.76 |
| 00111-84-2 | n-nonane | 0.95 |
| 00098-82-8 | (1-methylethyl)benzene | 2.32 |
| 15869-87-1 | 2,2-dimethyloctane | 1.09 |
| 04032-94-4 | 2,4-dimethyloctane | 1.09 |
| 02051-30-1 | 2,6-dimethyloctane | 1.27 |
| 00103-65-1 | n-propylbenzene | 2.20 |
| 00620-14-4 | 1-methyl-3-ethylbenzene | 6.61 |
| 00622-96-8 | 1-methyl-4-ethylbenzene | 6.61 |
| 00108-67-8 | 1,3,5-trimethylbenzene | 11.22 |
| 00611-14-3 | 1-methyl-2-ethylbenzene | 6.61 |
| 00095-63-6 | 1,2,4-trimethylbenzene | 7.18 |
| 00124-18-5 | n-decane | 0.83 |
| 00538-93-2 | (2-methylpropyl)benzene | 1.97 |
| 00135-98-8 | (1-methylpropyl)benzene | 1.97 |
| 00535-77-3 | 1-methyl-3-(1-methylethyl)benzene | 5.92 |
| 00526-73-8 | 1,2,3-trimethylbenzene | 11.26 |
| 00099-87-6 | 1-methyl-4-(1-methylethyl)benzene | 5.92 |
| 00496-11-7 | 2,3-dihydroindene (indan) | 3.17 |
| 00527-84-4 | 1-methyl-2-(1-methylethyl)benzene | 5.92 |
| 00141-93-5 | 1,3-diethylbenzene | 5.92 |
| 00105-05-5 | 1,4-diethylbenzene | 5.92 |
| 01074-43-7 | 1-methyl-3-n-propylbenzene | 5.92 |
| 01074-55-1 | 1-methyl-4-n-propylbenzene | 5.92 |
| 00135-01-3 | 1,2-diethylbenzene | 5.92 |
| 01074-17-5 | 1-methyl-2-n-propylbenzene | 5.92 |
| 01758-88-9 | 1,4-dimethyl-2-ethylbenzene | 8.86 |
| 00874-41-9 | 1,3-dimethyl-4-ethylbenzene | 8.86 |
| 00934-80-5 | 1,2-dimethyl-4-ethylbenzene | 8.86 |
| 02870-04-4 | 1,3-dimethyl-2-ethylbenzene | 8.86 |
| 01120-21-4 | n-undecane (hendecane) | 0.74 |
| 00933-98-2 | 1,2-dimethyl-3-ethylbenzene | 8.86 |
| 00095-93-2 | 1,2,4,5-tetramethylbenzene | 8.86 |
| 01595-11-5 | 1-methyl-2-n-butylbenzene | 5.35 |
| 00527-53-7 | 1,2,3,5-tetramethylbenzene | 8.86 |
| 01074-92-6 | 1-(1,1-dimethylethyl)-2-methylbenzene | 5.35 |
| 00488-23-3 | 1,2,3,4-tetramethylbenzene | 8.86 |
| 00538-68-1 | n-pentylbenzene | 1.78 |
| 00098-19-1 | 1-(1,1-dimethylethyl)-3,5-DMbenzene | 7.33 |
| 00091-20-3 | naphthalene | 3.26 |
| 00112-40-3 | n-dodecane | 0.66 |

| CAS # | COMPOUND | MIR |
|---------------------------|----------------------------------|------------|
| Carbonyl Compounds | | |
| 00050-00-0 | formaldehyde | 8.97 |
| 00075-07-0 | acetaldehyde | 6.84 |
| 00107-02-8 | acrolein | 7.60 |
| 00067-64-1 | acetone | 0.43 |
| 00123-38-6 | propionaldehyde | 7.89 |
| 00123-72-8 | butyraldehyde | 6.74 |
| 00066-25-1 | hexanaldehyde | 4.98 |
| 00100-52-7 | benzaldehyde | 0.00 |
| 00078-93-3 | methyl ethyl ketone (2-butanone) | 1.49 |
| 00078-85-3 | methacrolein | 6.23 |
| 04170-30-3 | crotonaldehyde | 10.07 |
| 00110-62-3 | valeraldehyde | 5.76 |
| 00620-23-5 | m-tolualdehyde | 0.00 |

**List of Compounds
(Listed by CAS number)**

| | |
|------------|---|
| 00050-00-0 | formaldehyde |
| 00064-17-5 | ethanol |
| 00066-25-1 | hexanaldehyde |
| 00067-56-1 | methanol |
| 00067-64-1 | acetone |
| 00071-43-2 | benzene |
| 00074-84-0 | ethane |
| 00074-85-1 | ethene |
| 00074-86-2 | ethyne |
| 00074-98-6 | propane |
| 00074-99-7 | 1-propyne |
| 00075-07-0 | acetaldehyde |
| 00075-28-5 | 2-methylpropane |
| 00075-83-2 | 2,2-dimethylbutane |
| 00078-78-4 | 2-methylbutane |
| 00078-79-5 | 2-methyl-1,3-butadiene |
| 00078-85-3 | methacrolein |
| 00078-93-3 | methyl ethyl ketone (2-butanone) |
| 00079-29-8 | 2,3-dimethylbutane |
| 00091-20-3 | naphthalene |
| 00095-47-6 | o-xylene |
| 00095-63-6 | 1,2,4-trimethylbenzene |
| 00095-93-2 | 1,2,4,5-tetramethylbenzene |
| 00096-14-0 | 3-methylpentane |
| 00096-37-7 | methylcyclopentane |
| 00098-19-1 | 1-(1,1-dimethylethyl)-3,5-dimethylbenzene |
| 00098-82-8 | (1-methylethyl)benzene |
| 00099-87-6 | 1-methyl-4-(1-methylethyl)benzene |
| 00100-41-4 | ethylbenzene |
| 00100-42-5 | stryrene (ethenylbenzene) |
| 00100-52-7 | benzaldehyde |
| 00103-65-1 | n-propylbenzene |
| 00105-05-5 | 1,4-diethylbenzene |
| 00106-97-8 | n-butane |
| 00106-98-9 | 1-butene |
| 00106-99-0 | 1,3-butadiene |
| 00107-00-6 | 1-butyne |
| 00107-02-8 | acrolein |
| 00107-39-1 | 2,4,4-trimethyl-1-pentene |
| 00107-40-4 | 2,4,4-trimethyl-2-pentene |
| 00107-83-5 | 2-methylpentane |
| 00108-08-7 | 2,4-dimethylpentane |

| | |
|------------|-----------------------------------|
| 00108-38-3 | m- & p-xylene |
| 00108-67-8 | 1,3,5-trimethylbenzene |
| 00108-87-2 | methylcyclohexane |
| 00108-88-3 | toluene |
| 00109-66-0 | n-pentane |
| 00109-67-1 | 1-pentene |
| 00110-54-3 | n-hexane |
| 00110-62-3 | valeraldehyde |
| 00110-82-7 | cyclohexane |
| 00110-83-8 | cyclohexene |
| 00111-65-9 | n-octane |
| 00111-66-0 | 1-octene |
| 00111-84-2 | n-nonane |
| 00112-40-3 | n-dodecane |
| 00115-07-1 | propene |
| 00115-11-7 | 2-methylpropene |
| 00123-38-6 | propionaldehyde |
| 00123-72-8 | butyraldehyde |
| 00124-11-8 | 1-nonene |
| 00124-18-5 | n-decane |
| 00135-01-3 | 1,2-diethylbenzene |
| 00135-98-8 | (1-methylpropyl)benzene |
| 00141-93-5 | 1,3-diethylbenzene |
| 00142-29-0 | cyclopentene |
| 00142-82-5 | n-heptane |
| 00287-92-3 | cyclopentane |
| 00463-49-0 | 1,2-propadiene |
| 00463-82-1 | 2,2-dimethylpropane |
| 00464-06-2 | 2,2,3-trimethylbutane |
| 00488-23-3 | 1,2,3,4-tetramethylbenzene |
| 00496-11-7 | 2,3-dihydroindene (indan) |
| 00503-17-3 | 2-butyne |
| 00513-35-9 | 2-methyl-2-butene |
| 00526-73-8 | 1,2,3-trimethylbenzene |
| 00527-53-7 | 1,2,3,5-tetramethylbenzene |
| 00527-84-4 | 1-methyl-2-(1-methylethyl)benzene |
| 00535-77-3 | 1-methyl-3-(1-methylethyl)benzene |
| 00538-68-1 | n-pentylbenzene |
| 00538-93-2 | (2-methylpropyl)benzene |
| 00540-84-1 | 2,2,4-trimethylpentane |
| 00542-92-7 | 1,3-cyclopentadiene |
| 00558-37-2 | 3,3-dimethyl-1-butene |
| 00560-21-4 | 2,3,3-trimethylpentane |
| 00562-49-2 | 3,3-dimethylpentane |
| 00563-16-6 | 3,3-dimethylhexane |
| 00563-45-1 | 3-methyl-1-butene |

| | |
|------------|--------------------------------|
| 00563-46-2 | 2-methyl-1-butene |
| 00565-59-3 | 2,3-dimethylpentane |
| 00565-75-3 | 2,3,4-trimethylpentane |
| 00584-94-1 | 2,3-dimethylhexane |
| 00589-34-4 | 3-methylhexane |
| 00589-43-5 | 2,4-dimethylhexane |
| 00589-53-7 | 4-methylheptane |
| 00589-81-1 | 3-methylheptane |
| 00590-18-1 | cis-2-butene |
| 00590-35-2 | 2,2-dimethylpentane |
| 00590-73-8 | 2,2-dimethylhexane |
| 00591-76-4 | 2-methylhexane |
| 00592-13-2 | 2,5-dimethylhexane |
| 00592-27-8 | 2-methylheptane |
| 00592-41-6 | 1-hexene |
| 00592-76-7 | 1-heptene |
| 00611-14-3 | 1-methyl-2-ethylbenzene |
| 00616-12-6 | 3-methyl-trans-2-pentene |
| 00617-78-7 | 3-ethylpentane |
| 00620-14-4 | 1-methyl-3-ethylbenzene |
| 00620-23-5 | m-tolualdehyde |
| 00622-96-8 | 1-methyl-4-ethylbenzene |
| 00624-64-6 | trans-2-butene |
| 00625-27-4 | 2-methyl-2-pentene |
| 00627-20-3 | cis-2-pentene |
| 00637-92-3 | 1-ethyl-tert-butyl-ether |
| 00638-04-0 | cis-1,3-dimethylcyclohexane |
| 00646-04-8 | trans-2-pentene |
| 00674-76-0 | 4-methyl-trans-2-pentene |
| 00689-97-4 | 1-buten-3-yne |
| 00691-37-2 | 4-methyl-1-pentene |
| 00691-38-3 | 4-methyl-cis-2-pentene |
| 00693-89-0 | 1-methylcyclopentene |
| 00760-20-3 | 3-methyl-1-pentene |
| 00763-29-1 | 2-methyl-1-pentene |
| 00816-79-5 | 3-ethyl-2-pentene |
| 00822-50-4 | trans-1,2-dimethylcyclopentane |
| 00874-41-9 | 1,3-dimethyl-4-ethylbenzene |
| 00922-62-3 | 3-methyl-cis-2-pentene |
| 00926-82-9 | 3,5-dimethylheptane |
| 00933-98-2 | 1,2-dimethyl-3-ethylbenzene |
| 00934-80-5 | 1,2-dimethyl-4-ethylbenzene |
| 01069-53-0 | 2,3,5-trimethylhexane |
| 01072-05-5 | 2,6-dimethylheptane |
| 01074-17-5 | 1-methyl-2-n-propylbenzene |
| 01074-43-7 | 1-methyl-3-n-propylbenzene |

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| 01074-55-1 | 1-methyl-4-n-propylbenzene |
| 01074-92-6 | 1-(1,1-dimethylethyl)-2-methylbenzene |
| 01120-21-4 | n-undecane (hendecane) |
| 01120-62-3 | 3-methylcyclopentene |
| 01595-11-5 | 1-methyl-2-n-butylbenzene |
| 01634-04-4 | 1-methyl-tert-butyl-ether |
| 01640-89-7 | ethylcyclopentane |
| 01678-91-7 | ethylcyclohexane |
| 01758-88-9 | 1,4-dimethyl-2-ethylbenzene |
| 01759-58-6 | trans-1,3-dimethylcyclopentane |
| 02051-30-1 | 2,6-dimethyloctane |
| 02207-01-4 | cis-1,2-dimethylcyclohexane |
| 02207-03-6 | trans-1,3-dimethylcyclohexane |
| 02207-04-7 | trans-1,4-dimethylcyclohexane |
| 02213-23-2 | 2,4-dimethylheptane |
| 02216-33-3 | 3-methyloctane |
| 02216-34-4 | 4-methyloctane |
| 02532-58-3 | cis-1,3-dimethylcyclopentane |
| 02613-65-2 | trans-1-methyl-3-ethylcyclopentane |
| 02738-19-4 | 2-methyl-2-hexene |
| 02815-58-9 | 1,2,4-trimethylcyclopentane |
| 02870-04-4 | 1,3-dimethyl-2-ethylbenzene |
| 03074-71-3 | 2,3-dimethylheptane |
| 03221-61-2 | 2-methyloctane |
| 03404-61-3 | 3-methyl-1-hexene |
| 03522-94-9 | 2,2,5-trimethylhexane |
| 03899-36-3 | 3-methyl-trans-3-hexene |
| 04032-94-4 | 2,4-dimethyloctane |
| 04050-45-7 | trans-2-hexene |
| 04170-30-3 | crotonaldehyde |
| 06443-92-1 | cis-2-heptene |
| 07385-78-6 | 3,4-dimethyl-1-pentene |
| 07642-04-8 | cis-2-octene |
| 07642-09-3 | cis-3-hexene |
| 07688-21-3 | cis-2-hexene |
| 10574-37-5 | 2,3-dimethyl-2-pentene |
| 13269-52-8 | trans-3-hexene |
| 13389-42-9 | trans-2-octene |
| 14686-13-6 | trans-2-heptene |
| 14686-14-7 | trans-3-heptene |
| 14850-23-8 | trans-4-octene |
| 15869-87-1 | 2,2-dimethyloctane |
| 15890-40-1 | (1a,2a,3b)-1,2,3-trimethylcyclopentane |
| 16747-50-5 | cis-1-methyl-3-ethylcyclopentane |

APPENDIX 2

DEFINITIONS AND COMMONLY USED ABBREVIATIONS

- I. The abbreviations and definitions set forth in this section apply to Parts A through G of these test procedures:
- ASTM = American Society for Testing and Materials
- Carbon No. = number of carbon atoms in the hydrocarbon or organic compound being measured.
- CCR = California Code of Regulations
- CH_3OH_d = the methanol concentration in the dilution air as determined from the dilution air methanol sample using the procedure specified in Method No. 1001, ppmC.
- CH_3OH_e = the methanol concentration in the dilute exhaust as determined from the dilute exhaust methanol sample using the procedure specified in Method No. 1001, ppmC.
- CH_4_d = the methane concentration in the dilution air, ppmC.
- CH_4_e = the methane concentration in the dilute exhaust, ppmC.
- $\text{C}_2\text{H}_5\text{OH}_d$ = the ethanol concentration in the dilution air as determined from the dilution air ethanol sample using the procedure specified in Method No. 1001, ppmC.
- $\text{C}_2\text{H}_5\text{OH}_e$ = the ethanol concentration in the dilute exhaust as determined from the dilute exhaust ethanol sample using the procedure specified in Method No. 1001, ppmC.
- CNG = compressed natural gas
- CO_e = the carbon monoxide concentration in the dilute exhaust corrected for carbon dioxide and water removal, ppm.
- CO_{em} = the carbon monoxide concentration in the dilute exhaust uncorrected for carbon dioxide and water removal, ppm.
- CO_{2e} = the carbon dioxide concentration in the dilute exhaust, %.
- CVS = constant volume sampler

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| $D_{\text{phase } n}$ | = | the distance driven by the test vehicle on a chassis dynamometer during test phase n (where n is either 1, 2, or 3), mile. |
| $Dens_{\text{ROH}}$ | = | density of alcohol, g/mL. |
| DF | = | dilution factor (see Dilution Factor Calculation). |
| FID | = | flame ionization detector |
| FID THC_d | = | the total hydrocarbon concentration including methane and methanol (for methanol-fueled engines) or ethanol (for ethanol-fueled engines) in the dilution air as measured by the FID, ppmC. |
| FID THC_e | = | the total hydrocarbon concentration including methane and methanol (for methanol-fueled engines) or ethanol (for ethanol-fueled engines) in the dilution exhaust as measured by the FID, ppmC. |
| FTP | = | Federal Test Procedure |
| GC | = | gas chromatograph |
| GC/MS | = | gas chromatography/mass spectrometry |
| HC_{conc} | = | net concentration of an HC compound in the dilute exhaust corrected for background per test phase, ppbC. |
| HC_d | = | composite concentration of an HC compound in the dilution air (background) for all three test phases as determined from the composite dilution air sample using the procedure specified in Method No. 1002 and Method No. 1003, ppbC. |
| HC_{dens} | = | mass per unit volume of an HC compound corrected to standard conditions (293.16 K and 760 mm Hg) g/ft^3 . |
| HC_e | = | concentration of an HC compound in the dilute exhaust per test phase as determined from the dilute exhaust sample using the procedure specific in Method No. 1002 and Method No. 1003, ppbC. |
| $\text{HC}_{\text{mass } n}$ | = | mass emissions of an HC compound per test phase n (where n is either 1, 2, or 3), mg. |

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| HC _{wm} | = | total weighted mass of an HC compound per mile, g/mile. |
| HCHO _e | = | formaldehyde concentration in the dilute exhaust as determined from the dilute exhaust carbonyl sample using the procedure specified in Method No 1004, ppm. |
| HCR | = | the hydrogen-to-carbon ratio for the fuel used. |
| HPLC | = | high performance liquid chromatography |
| Iconc _{blk} | = | concentration of the blank cartridge, µg/mL |
| Iconc _{cd} | = | total concentration of carbonyl compound extracted from both cartridges for the dilution air, µg/mL |
| Iconc _{ce} | = | total concentration of carbonyl compound extracted from both cartridges for the diluted exhaust, µg/mL |
| Iconc _{d1} | = | dilution air (background) alcohol concentration in the primary impinger for all three test phases as determined by the procedure specified in Method No. 1001, µg/mL. |
| Iconc _{d2} | = | dilution air (background) alcohol concentration in the secondary impinger for all three test phases as determined by the procedure specified in Method No. 1001, µg/mL. |
| Iconc _{e1} | = | dilute exhaust alcohol concentration in the primary impinger per test phase as determined by the procedure specified in Method No. 1001, µg/mL. |
| Iconc _{e2} | = | dilute exhaust alcohol concentration in the secondary impinger per test phase as determined by the procedure specified in Method No. 1001, µg/mL. |
| Imass _d | = | total mass of an alcohol or carbonyl compound collected from the dilution air (background) in both primary and secondary impingers/cartridges for all three test phases as determined by the procedure specified in Method No. 1001 (alcohol) or Method No. 1004 (carbonyl), µg. |
| Imass _e | = | total mass of an alcohol or carbonyl compound collected from the dilute exhaust in both primary and secondary impingers/cartridges per test phase as determined by the procedure specified in Method No. 1001 (alcohol) or Method No. 1004 (carbonyl), µg. |

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| I_{temp_d} | = | dilution air temperature at the flowmeter inlet for impinger/cartridge sampling, °K. |
| I_{temp_e} | = | dilute exhaust temperature at the flowmeter inlet for impinger/cartridge sampling, °K. |
| I_{vol_c} | = | elution volume of the cartridge, mL (The cartridge is extracted with 5 mL acetonitrile, but 0.6 mL is retained in the cartridge, so the elution volume is 4.4 mL.) |
| I_{vol_d} | = | total volume of dilution air (background) drawn through the impingers/cartridges for all three test phases corrected to standard conditions (293.16°K and 760 mm Hg), liter. |
| $I_{vol_{dm}}$ | = | total volume of dilution air (background) drawn through the impingers/cartridges for all three test phases as measured during testing, liter. |
| I_{vol_e} | = | total volume of dilute exhaust drawn through the impingers/cartridges per test phase corrected to standard conditions (293.16°K and 760 mm Hg), liter. |
| $I_{vol_{em}}$ | = | total volume of dilute exhaust drawn through the impingers/cartridges per test phase as measured during testing, liter. |
| I_{vol_r} | = | volume of the reagent used in an impinger, mL. |
| LOD | = | limit of detection |
| LPG | = | liquified petroleum gas |
| Mol. Vol. | = | molecular volume which is 24.055 liter/mole at standard conditions (293.16°K and 760 mm Hg). |
| Mol. Wt. | = | molecular weight of the compound being measured, g/mole. |
| NIST | = | National Institute of Standards and Technology |
| NMHC | = | non-methane hydrocarbons |
| $NMHC_{conc}$ | = | the non-methane hydrocarbon concentration in the dilute exhaust corrected for background, ppmC. |
| $NMHC_d$ | = | the non-methane hydrocarbon concentration in the dilution air corrected for methane and alcohol removal, ppmC. |

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| NMHC _{dens} | = | the mass per unit volume of non-methane hydrocarbon corrected to standard conditions (16.33 g/ft ³ at 293.16°K and 760 mm Hg assuming a C:H ratio of 1:1.85 for gasoline; 16.78 g/ft ³ at 293.16°K and 760 mm HG assuming a C:H ratio of 1:1.94 for Phase 2 reformulated gasoline; 19.52 g/ft ³ at 293.16°K and 760 mm HG assuming a C:H ratio of 1:3.78 for natural gas; and 17.26 g/ft ³ for LPG at 293.16°K and 760 mm Hg assuming a C:H ratio of 1:2.64), g/ft ³ . |
| NMHC _e | = | non-methane hydrocarbon concentration in the dilute exhaust corrected for methane and alcohol removal, ppmC. |
| NMHC _{mass n} | = | the mass emission of non-methane hydrocarbon per test phase n (where n is either 1, 2, or 3), g. |
| NMHC _{wm} | = | the total weighted mass of non-methane hydrocarbon per mile for all three phases of the FTP, g/mile. |
| NMOG | = | non-methane organic gases |
| P _B | = | barometric pressure during testing, mm Hg. |
| PID | = | photoionization detector |
| PLOT | = | porous layer open tubular |
| R _a | = | the relative humidity of the ambient air, %. |
| r _{CH₃OH} | = | the FID response factor to methanol (see CFR 40, 86.121-90(c)). |
| r _{CH₄} | = | the FID response factor to methane (see Part B, "Determination of NMHC by FID"). |
| r _{C₂H₅OH} | = | the FID response factor to ethanol (same procedure for methanol response factor, see CFR 40, 86.121-90(c)). |
| RHO | = | generic symbol representing a carbonyl compound such as formaldehyde, acetaldehyde, acetone, etc. |
| RHO _{conc} | = | net concentration of a carbonyl compound in the dilute exhaust corrected for background per test phase, ppm. |

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| RHO_d | = | composite concentration of a carbonyl compound in the dilution air (background) for all three test phases, ppm. |
| RHO_{dens} | = | mass per unit volume of a carbonyl compound corrected to standard conditions (293.16°K and 760 mm Hg), g/ft ³ . |
| RHO_e | = | concentration of a carbonyl compound in the dilute exhaust per test phase, ppm. |
| $RHO_{mass\ n}$ | = | mass emissions of a carbonyl compound per test phase n (where n is either 1, 2, or 3), g. |
| RHO_{wm} | = | total weighted mass emissions of a carbonyl compound per mile, g/mile. |
| ROH | = | generic symbol representing an alcohol compound such as methanol or ethanol. |
| ROH_{conc} | = | net concentration of an alcohol compound in the dilute exhaust corrected for background per test phase, ppm. |
| ROH_d | = | composite concentration of an alcohol compound in the dilution air (background) for all three test phases, ppm. |
| ROH_{dens} | = | mass per unit volume of an alcohol compound corrected to standard conditions (293.16°K and 760 mm Hg), g/ft ³ . |
| ROH_e | = | concentration of an alcohol compound in the dilute exhaust per test phase, ppmC. |
| $ROH_{mass\ n}$ | = | mass emissions of an alcohol compound per test phase n (where n is either 1, 2, or 3), g. |
| ROH_{wm} | = | total weighted mass emissions of an alcohol compound per mile, g/mile. |
| SAE | = | Society of Automotive Engineers |
| SRM | = | Standard Reference Material |
| VMIX | = | the total dilute exhaust volume measured per test phase and corrected to standard conditions (293.16°K and 760 mm Hg), ft ³ . |

II. The following list is commonly used measurement abbreviations:

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| g | = | gram |
| μg | = | microgram |
| m | = | meter |
| cm | = | centimeter |
| μm | = | micrometer |
| μ | = | micron |
| L | = | liter |
| mL | = | milliliter |
| μL | = | microliter |
| ppb | = | parts per billion. |
| ppbC | = | parts per billion carbon equivalent. |
| ppm | = | parts per million. |
| ppmC | = | parts per million carbon equivalent. |

APPENDIX 3

REFERENCES

- [1] Code of Federal Regulations, Title 40, Part 86, Subpart B
- [2] SAE J254, "Instrumentation and Techniques for Exhaust Gas Emissions Measurement"
- [3] SAE J1094a, "Constant Volume Sampler System for Exhaust Emissions Measurement"
- [4] SAE 770141, "Optimization of a Flame Ionization Detector for Determination of Hydrocarbons in Diluted Automotive Exhausts". G.D. Reschke, Vehicle Emissions Laboratory, General Motors Proving Ground
- [5] SAE J1154, "Methane Measurement Using Gas Chromatography," (revised December 1991)
- [6] U.S. Environmental Protection Agency, Characterization of Exhaust Emissions from Methanol and Gasoline Fueled Automobiles, EPA 460/3-82-004.
- [7] U.S. Environmental Protection Agency, Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air, (Method T03-15) EPA-600/4-89-017 Research Triangle Park, North Carolina, June, 1989.
- [8] Standard Test Method for C₁ through C₆ Hydrocarbons in the Atmosphere by Gas Chromatography, American Standards for Testing Materials (ASTM) Standards on Chromatography (1981).
- [9] U.S. Environmental Protection Agency, Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air, (Method T03-15) EPA-600/4-84-041 Research Triangle Park, North Carolina, April, 1989.
- [10] Hull, L.A., Procedures for 2,4-Dinitrophenylhydrazone Aldehyde-Ketone Air Analysis, Internal Report at U.S. EPA.
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- [12] Keith, L. H., Taylor, J.K., et al, "Principles of Environmental Analysis", Analytical Chemistry, Vol. 55, No. 14, December 1983.